



TRICHINELLA SPIRALIS MUST INHIBIT THE IMMUNE SYSTEM TO SURVIVE

I. Frydas* and S. Frydas

Department of Parasitology, Aristotle University, Thessaloniki, Greece.

*Correspondence to:

Dr. Ilias Frydas,
Department of Parasitology,
Aristotle University,
54124 Thessaloniki, Greece.
e-mail: ilias.frydas@gmail.com

ABSTRACT

Trichinella spiralis is a parasitic nematode that infects various mammals, including humans. This tissue-dwelling parasite plays its pathological role by affecting the immune system and causes the disease trichinellosis. *T. spiralis* generates a variety of proteins that protect it from the host's immune system and it releases inhibitory proteins such as proteases, antioxidant enzymes, and immunomodulatory molecules. *T. spiralis* modulates T-cell and macrophage responses, reducing the immunological response launched against it by the organism. By producing proteases, the parasite inhibits the infiltration of immune cells that would help fight the infection. These anti-immune effects help *T. spiralis* survive. In addition, this parasite helps the response shift from Th1 to Th2, an effect that is less effective in eliminating infections. Recently, it has been reported that antigens secreted by *T. spiralis* can interact with immune cells by binding to the toll-like receptor (TLR), reducing the inflammatory response. It can be concluded that *T. spiralis* affects the immune system to survive and carry out its pathological action, but it can also produce immunity-stimulating antigens, an effect that could be useful to fight certain autoimmune diseases that are currently incurable.

KEYWORDS: *Trichinella spiralis*, helminth, parasite, nematode, immunity

INTRODUCTION

Helminths are pathological parasites that act on the host's immune system by activating and modulating it. *Trichinella spiralis* is a parasitic nematode that infects various mammals, including humans, and causes trichinellosis (or trichinosis) when organisms consume raw meat containing its larvae.

T. spiralis includes many species such as *T. native*, *T. nelsoni*, *T. britovi*, *T. murrelli*, and the new species, *T. chanchalensis* (1), and is present in many low income and industrialized countries. Some species of *T. spiralis*, such as *T. pseudospiralis*, *T. papuae*, and *T. zimbabwensis* are not encapsulated (2). The biological cycle is immediate; eggs are released into the external environment and the larva develops. When the climatic conditions are mild, the parasite remains infectious for 5-8 years (3). To establish and maintain infection, it must evade and manipulate the host immune system (4).

T. spiralis is very effective at evading the immune system. Trichinellosis, the infection caused by this parasite, is a disease where there is immune stimulation due to antigen molecules of the parasite. The molecular mechanisms by which *T. spiralis* achieves this are diverse and sophisticated, involving both evasion and active suppression of immune responses.

The parasite can induce both pro-inflammatory and anti-inflammatory cytokines, which can have varying effects on the immune system (5,6). The acute phase of infection is associated with a strong pro-inflammatory Th1 response (7),

Received: 01 February, 2022
Accepted: 19 April, 2022

1972-6945 (2022)

Copyright © by Biolife-Publisher

This publication and/or article is for individual use only and may not be further reproduced without written permission from the copyright holder. Unauthorized reproduction may result in financial and other penalties. Disclosure: all authors report no conflicts of interest relevant to this article.

while the chronic phase often involves a shift toward an anti-inflammatory Th2 response, characterized by the production of cytokines like IL-4, IL-10, and TGF- β (8).

It has been reported that *T. spiralis* might have a protective effect against certain autoimmune diseases (9). However, the relationship between *T. spiralis* and autoimmunity is complex and has multifaceted aspects.

DISCUSSION

T. Spiralis modulates host immune responses in different ways. The parasite secretes a variety of proteins that help it avoid detection and destruction by the host immune system (10). These proteins can interfere with antigen presentation and reduce the host's ability to mount an effective immune response (11). Furthermore, the parasite can directly suppress immune cell function. For example, *T. spiralis* can modulate the activity of T cells and macrophages to reduce their ability to attack the parasite (12,13).

When activated, *T. spiralis* releases excretory and secretory products such as protease inhibitors, antioxidant enzymes, immunomodulatory molecules, and other enzymes. Protease inhibitors are molecules that can inhibit host proteases, which are part of the host's immune defense mechanisms (14). By inhibiting these enzymes, *T. spiralis* can prevent tissue damage and immune cell infiltration that would otherwise help clear the infection (7).

The parasite also produces antioxidant enzymes such as superoxide dismutase and glutathione peroxidase, which neutralize reactive oxygen species (ROS) produced by immune cells (15). This helps the parasite survive the oxidative burst that is part of the host's immune response.

T. spiralis can secrete immunomodulatory molecules that mimic host cytokines or interfere with cytokine signaling (12). This can skew the immune response towards a Th2 type response, which is less effective at clearing helminth infections compared to a Th1 type response (16). Excretory and secretory antigens are specific proteins secreted by *T. spiralis* that can directly interact with immune cells to alter their function. For example, some excretory/secretory antigens can bind to toll-like receptors (TLRs) on immune cells, altering their signaling pathways and reducing inflammatory responses (12,17).

T. spiralis infection can lead to the induction of regulatory T cells (Tregs), expanding these immune cells that suppress other immune responses. This helps create an environment that is more permissive for the parasite's survival. *T. spiralis* can also modify host tissues and affect cells they are in contact with (18). *T. spiralis* larvae form cysts within host muscle tissues, creating a physical barrier that protects them from immune cells (19). The cyst wall is also immunologically inert, meaning it does not provoke a strong immune response.

T. spiralis also modulates gene expression, affecting microRNAs (miRNAs) (20). This helminth can release miRNAs that are taken up by host cells, leading to changes in the host gene expression profile (21). This can downregulate genes involved in the immune response and create a more favorable environment for the parasite (21,22).

Specific molecules and pathways are affected by *T. Spiralis*. The *T. spiralis* serine protease TsSerp is involved in degrading host proteins and modulating immune responses by interfering with cytokine production and immune cell signaling pathways (24). *T. spiralis* excretory/secretory (Ts-ES) antigens include a variety of proteins and glycoproteins that interact with host immune cells (12). They can modulate dendritic cell function, inhibit T cell proliferation, and alter cytokine production (18, 23).

T. spiralis can induce the production of vascular endothelial growth factor (VEGF), which helps in creating a favorable environment for the larvae in the host muscle by promoting angiogenesis and tissue repair processes that may help in cyst formation and maintenance (25).

The antigenic molecules expressed by this parasite can activate innate immunity with consequent stimulation of adaptive immunity. The antigens act on antigen presenting cells (APCs) such as macrophages and dendritic cells which downregulate adaptive immune cells, such as Tregs and regulatory B cells, activating a sort of immunosuppression (18). This immunosuppression is induced by inhibitory cytokines such as IL-10, IL-17, and TGF- β , and this reaction is exerted by *T. spiralis* to avoid the immune response and allow its survival (12,26). This mechanism could be useful for suppressing the overstimulated immune system in chronic inflammatory diseases such as autoimmune diseases, transplant rejection, and allergic diseases such as asthma (27-29).

T. spiralis larvae can invade the intestine and activate epithelial cells with the secretion of cytokines IL-25 and IL-33 (also called alarmin). These cytokines activate type 2 innate lymphoid cells (ILC2), which in turn produce IL-5 and IL-13 cytokines that act on the precursors of epithelial cells to increase the production of immunosuppressive cytokines (30).

The type 2 immune response induced by the antigenic molecules causes the stimulation of IgE, IgG1, and IgG4 antibodies and anti-inflammatory cytokines including IL-4, IL-13, and IL-33 (31,32). These cytokines facilitate the proliferation of basophils, eosinophils, and macrophages, increasing allergic phenomena and mucus secretion, which are elements of the parasitic immune defense (16).

By implementing these strategies, *T. spiralis* escapes the host's immune system and survives. The infection causes an increase in eosinophilic granulocytes, which is a hallmark characteristic of parasitic infections that protects the parasite (33). Proteins derived from *T. spiralis* larvae are phagocytosed by macrophages, altering their ability to present antigen and activate T cells, a reaction which is mediated by STAT4 (6). The negative regulation of STAT4 and the cytokine IL-12 causes an inhibition of interferon-gamma (IFN- γ) that allows the *T. spiralis* to evade the immune response (34).

CONCLUSIONS

Understanding the biology and the mechanisms by which *T. spiralis* evades the immune response and affects host tissue is crucial for developing strategies to combat the infection and for designing new therapeutic approaches that can overcome the parasite's immune evasion tactics. In addition, immune system activation by helminths is currently being studied for its application in the treatment of diverse pathologies, including autoimmune diseases, that have a poor response to treatment.

Conflict of interest

The authors declare that they have no conflict of interest.

REFERENCES

1. Sharma R, Thompson PC, Hoberg EP, et al. Hiding in plain sight: discovery and phylogeography of a cryptic species of *Trichinella* (Nematoda: Trichinellidae) in wolverine (*Gulo gulo*). *International Journal for Parasitology*. 2020;50(4):277-287. doi:<https://doi.org/10.1016/j.ijpara.2020.01.003>
2. Kapel CMO. Host diversity and biological characteristics of the *Trichinella* genotypes and their effect on transmission. *Veterinary Parasitology*. 2000;93(3-4):263-278. doi:[https://doi.org/10.1016/s0304-4017\(00\)00345-9](https://doi.org/10.1016/s0304-4017(00)00345-9)
3. Gottstein B, Pozio E, Nockler K. Epidemiology, Diagnosis, Treatment, and Control of Trichinellosis. *Clinical Microbiology Reviews*. 2009;22(1):127-145. doi:<https://doi.org/10.1128/cmr.00026-08>
4. Song Y, Xu J, Wang X, et al. Regulation of host immune cells and cytokine production induced by *Trichinella spiralis* infection. *Parasite*. 2019;26:74. doi:<https://doi.org/10.1051/parasite/2019074>
5. Min Kyoung Cho, Mi Kyung Park, Shin Ae Kang, Seon Hee Choi, Soon Cheol Ahn, Hak Sun Yu. *Trichinella spiralis* Infection Suppressed Gut Inflammation with CD4⁺CD25⁺Foxp3⁺T Cell Recruitment. *Korean Journal of Parasitology*. 2012;50(4):385-390. doi:<https://doi.org/10.3347/kjp.2012.50.4.385>
6. Xu N, Bai X, Liu Y, et al. The Anti-Inflammatory Immune Response in Early *Trichinella spiralis* Intestinal Infection Depends on Serine Protease Inhibitor-Mediated Alternative Activation of Macrophages. *The Journal of Immunology*. 2021;206(5):963-977. doi:<https://doi.org/10.4049/jimmunol.2000290>
7. Xu J, Yu P, Wu L, Liu M, Lu Y. Regulatory effect of two *Trichinella spiralis* serine protease inhibitors on the host's immune system. *Scientific Reports*. 2019;9(1). doi:<https://doi.org/10.1038/s41598-019-52624-5>
8. Bruschi F, Gómez-Morales MA. The translational immunology of trichinellosis: from rodents to humans. In: Jirillo E, Magrone T, Miragliotta G, editors. *Immune Response to Parasitic Infections—Immunity to Helminths and Novel Therapeutic Approaches*. Vol. 2. Bantham E-Books; 2014. pp. 125–161.
9. Ilić N, Kosanović M, Gruden-Movsesijan A, et al. Harnessing immunomodulatory mechanisms of *Trichinella spiralis* to design novel nanomedical approaches for restoring self-tolerance in autoimmunity. *Immunology Letters*. 2021;238:57-67. doi:<https://doi.org/10.1016/j.imlet.2021.04.012>
10. Han C, Yu J, Zhang Z, et al. Immunomodulatory effects of *Trichinella spiralis* excretory-secretory antigens on macrophages. *Experimental Parasitology*. 2019;196:68-72. doi:<https://doi.org/10.1016/j.exppara.2018.10.001>
11. Hao C, Wang W, Zhan B, et al. *Trichinella spiralis* Paramyosin Induces Colonic Regulatory T Cells to Mitigate Inflammatory Bowel Disease. *Frontiers in Cell and Developmental Biology*. 2021;9. doi:<https://doi.org/10.3389/fcell.2021.695015>
12. Sun XM, Guo K, Hao CY, Zhan B, Huang JJ, Zhu X. *Trichinella spiralis* Excretory-Secretory Products Stimulate Host Regulatory T Cell Differentiation through Activating Dendritic Cells. *Cells*. 2019;8(11):1404. doi:<https://doi.org/10.3390/cells8111404>

13. Bai X, Wu X, Wang X, et al. Regulation of cytokine expression in murine macrophages stimulated by excretory/secretory products from *Trichinella spiralis* in vitro. *Molecular and Cellular Biochemistry*. 2011;360(1-2):79-88. doi:<https://doi.org/10.1007/s11010-011-1046-4>
14. Xu N, Liu X, Tang B, et al. Recombinant *Trichinella pseudospiralis* Serine Protease Inhibitors Alter Macrophage Polarization In Vitro. *Frontiers in Microbiology*. 2017;8. doi:<https://doi.org/10.3389/fmicb.2017.01834>
15. Derda M, Wandurska-Nowak E, Hadaś E. Changes in the level of antioxidants in the blood from mice infected with *Trichinella spiralis*. *Parasitology Research*. 2004;93(3):207-210. doi:<https://doi.org/10.1007/s00436-004-1093-9>
16. Motran CC, Silvano L, Chiapello LS, et al. Helminth Infections: Recognition and Modulation of the Immune Response by Innate Immune Cells. *Frontiers in Immunology*. 2018;9. doi:<https://doi.org/10.3389/fimmu.2018.00664>
17. Yu YR, Deng MJ, Lu WW, Jia MZ, Wu W, Qi YF. Systemic cytokine profiles and splenic toll-like receptor expression during *Trichinella spiralis* infection. *Experimental Parasitology*. 2013;134(1):92-101. doi:<https://doi.org/10.1016/j.exppara.2013.02.014>
18. Sofronic-Milosavljevic L, Ilic N, Pinelli E, Gruden-Movsesijan A. Secretory Products of *Trichinella spiralis* Muscle Larvae and Immunomodulation: Implication for Autoimmune Diseases, Allergies, and Malignancies. *Journal of Immunology Research*. 2015;2015:1-14. doi:<https://doi.org/10.1155/2015/523875>
19. Wu Z, Sofronic-Milosavljevic L, Nagano I, Takahashi Y. *Trichinella spiralis*: nurse cell formation with emphasis on analogy to muscle cell repair. *Parasites & Vectors*. 2008;1(1):27. doi:<https://doi.org/10.1186/1756-3305-1-27>
20. Buck AH, Coakley G, Simbari F, et al. Exosomes secreted by nematode parasites transfer small RNAs to mammalian cells and modulate innate immunity. *Nature Communications*. 2014;5(1). doi:<https://doi.org/10.1038/ncomms6488>
21. Taylor PJ, Hagen J, Faruqu FN, et al. *Trichinella spiralis* secretes abundant unencapsulated small RNAs with potential effects on host gene expression. *International Journal for Parasitology*. 2020;50(9):697-705. doi:<https://doi.org/10.1016/j.ijpara.2020.05.008>
22. Maizels RM, Smits HH, McSorley HJ. Modulation of Host Immunity by Helminths: The Expanding Repertoire of Parasite Effector Molecules. *Immunity*. 2018;49(5):801-818. doi:<https://doi.org/10.1016/j.immuni.2018.10.016>
23. Bruschi F, Chiumiento L. Immunomodulation in Trichinellosis: Does *Trichinella* Really Escape the Host Immune System? *Endocrine, Metabolic & Immune Disorders-Drug Targets*. 2012;12(1):4-15. doi:<https://doi.org/10.2174/187153012799279081>
24. Yue X, Sun XY, Liu F, et al. Molecular characterization of a *Trichinella spiralis* serine proteinase. *Veterinary Research*. 2020;51(1). doi:<https://doi.org/10.1186/s13567-020-00847-0>
25. Shariati F, Pérez-Arellano JL, López-Abán J, Arefi M, Martínez-Fernández AR, Muro A. *Trichinella*: Differential expression of angiogenic factors in macrophages stimulated with antigens from encapsulated and non-encapsulated species. *Experimental Parasitology*. 2009;123(4):347-353. doi:<https://doi.org/10.1016/j.exppara.2009.08.016>
26. Zheng W, Ma Z, Sun X, et al. Exposure time determines the protective effect of *Trichinella spiralis* on experimental colitis. *Microbial Pathogenesis*. 2020;147:104263-104263. doi:<https://doi.org/10.1016/j.micpath.2020.104263>
27. Ilic N, Gruden-Movsesijan A, Sofronic-Milosavljevic L. *Trichinella spiralis*: shaping the immune response. *Immunologic Research*. 2012;52(1-2):111-119. doi:<https://doi.org/10.1007/s12026-012-8287-5>
28. Kim SE, Kim JH, Min BH, Bae YM, Hong ST, Choi MH. Crude Extracts of *Caenorhabditis elegans* Suppress Airway Inflammation in a Murine Model of Allergic Asthma. *PLOS ONE*. 2012;7(4):e35447-e35447. doi:<https://doi.org/10.1371/journal.pone.0035447>
29. Gruden-Movsesijan A, Ilic N, Mostarica-Stojkovic M, Stosic-Grujicic S, Milic M, Sofronic-Milosavljevic Lj. *Trichinella spiralis*: Modulation of experimental autoimmune encephalomyelitis in DA rats. *Experimental Parasitology*. 2008;118(4):641-647. doi:<https://doi.org/10.1016/j.exppara.2007.12.003>
30. Angkasekwinai P, Srimanote P, Wang YH, et al. Interleukin-25 (IL-25) Promotes Efficient Protective Immunity against *Trichinella spiralis* Infection by Enhancing the Antigen-Specific IL-9 Response. Urban JF, ed. *Infection and Immunity*. 2013;81(10):3731-3741. doi:<https://doi.org/10.1128/iai.00646-13>
31. Anthony RM, Rutitzky LI, Urban JF, Staderker MJ, Gause WC. Protective immune mechanisms in helminth infection. *Nature Reviews Immunology*. 2007;7(12):975-987. doi:<https://doi.org/10.1038/nri2199>
32. Watanabe N, Bruschi F, Korenaga M. IgE: a question of protective immunity in *Trichinella spiralis* infection. *Trends in Parasitology*. 2005;21(4):175-178. doi:<https://doi.org/10.1016/j.pt.2005.02.010>

-
33. Huang L, Gebreselassie NG, Gagliardo LF, et al. Eosinophils mediate protective immunity against secondary nematode infection. *Journal of Immunology (Baltimore, Md: 1950)*. 2015;194(1):283-290. doi:<https://doi.org/10.4049/jimmunol.1402219>
34. Kobpornchai P, Tiffney EA, Adisakwattana P, Flynn RJ. Trichinella spiralis cystatin, TsCstN, modulates STAT4/IL-12 to specifically suppress IFN- γ production. *Cellular Immunology*. 2021;362:104303-104303. doi:<https://doi.org/10.1016/j.cellimm.2021.104303>