

Review

THE BONE-IMPLANT CONTACT AND OSSEOINTEGRATION OF DIFFERENT IMPLANT SURFACE TREATMENT: THE FINDINGS FROM A SYSTEMATIC REVIEW OF LITERATURE

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ABSTRACT

The dental implant is associated with high long-term predictability for fixed rehabilitation in edentulous patients. The aim of the present review was to evaluate the state-of-art of dental implant surface treatment and their effect on osseointegration. The Pubmed/Medline, EMBASE, Cochrane Library databases has been screened to identify the histologic studies regarding the dental implant surfaces *in vivo*. The screening process revealed a total of 3173 papers with a total of 24 articles obtained by the manual search. A total of 482 duplicates have been removed and 2691 papers were assessed for the full-text evaluation. A total of 2527 articles were removed after the eligibility process and 149 articles were evaluated for the descriptive analysis. The implant osseointegration process is a complex combination of events that is oriented to an intimate interface between the dental implant surface and the host peri-implant tissues that oriented to produce a functional ankylotic relationship between the components under the masticatory loading.

KEYWORDS: *implant, fixture, surface, osseointegration, bone*

INTRODUCTION

The dental implant osseointegration represents the turning point for edentulous ridge rehabilitations due to the more

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recent advances in titanium biocompatibility, enhanced surface treatment and novel high hydrophilic/bioactive materials (1), with a long-term implant success rates over 90% (2). The osteoconduction process is involved with the recruitment and migration of osteogenic cells to the implant surface determines the early events correlated to the dental implant osseointegration. This phase produces a mineralised osteoid matrix deposition representings the main non-functionalised new bone formation at the level of the bone-implant interface. These events are strictly correlated with several factors including the dental implant microtopography (3). Other key factors are represented by the implant material, macro design, surface chemistry, bone density, surgical technique, and implant loading protocol (4). In literature, the bone-implant contact (BIC) percentage represent one of the most reliable parameter for dental implant osseointegration, while values >50% are considered optimal for a long term stability findings (5). On the contrary, the main disadvantage of this parameter is dynamic and could potentially vary over time. In addition, the BIC% is a bidimensional parameter that could be determined only with retrieved biopsies and is not replicable.

Also, the torque removal force has been suggested as an additional technique to assess the implant anchorage for research purposes evaluating the biomechanical behaviour of osseointegration (6). In this way, the roughness of a surface is one of the major factors contributing to implant stability, based on the assessment of the surface peaks and valleys. For this purpose, the arithmetic mean height deviation from a mean bi-dimensional plane (Ra); the Sa is considered in the case of a three-dimensional evaluation (7). The "osseointegration" concept was introduced by Branemark et al. (1) as the direct contact between living bone and a functionally loaded implant surface without interposed soft tissue at the light microscope level (8).

Today, titanium is the most common material for dental implants due to its low weight, high strength/weight ratio, low elasticity modulus, corrosion and wearing resistance, and biocompatibility (9). The most frequent titanium alloy (Ti6Al4V) is composed of 6% of aluminium and 4% vanadium (10). Lincks et al. (11) reported that the osteoblasts-like cells responded differently to cpTi and Ti6Al4V materials due to the alloy mosaicism and the surface chemistry. A passive surface oxide film around the titanium core (12) determines the interface generation between the titanium surface and the surrounding hard tissue. The oxide layer produces hydroxyl functional groups when exposed to the air environment (13). The hydroxyl functional groups dissociate when exposed to body fluid to generate an electric charge that is correlated to the pH of the fluids (13). In this way, the point of zero charge of rutile is 5.3, while the anatase point of zero charge is 6.2 (14, 15). The TiO2 shows reported a neutral property. The hydroxyl concentration of TiO2 is relatively large, representing an advantage for the proteins and cytokines adsorption promotion (12). The machined surfaces of the implant device are provided only by decontamination after the turning procedure.

Various treatments were proposed to improve the surface properties, taking advantage of rough interfaces with high implant stability and the surface contact area (6, 16, 17). In addition, rough surfaces seem to be effective in improving the osteogenic cell's behaviour (18, 19), proliferation and differentiation (20, 21) due to the release of signal mediators, transforming growth factor beta, and prostaglandin E2 (PGE2) (21-24). The optimal roughness for dental implant surfaces range is approximately 1.5 μ m (25). Several methods have been suggested, such as modified surfaces, additive coating protocols, and subtractive methods, while today, the optimal surface type has not been defined. The present systematic reviews aimed to investigate the recent updates of bone-implant contact (BIC) effectiveness of different implant surface treatments.

MATERIALS AND METHODS

Article search methodology

The screening phase was conducted according to the Standards for Reporting Qualitative Research principles (SRQR) and the PRISMA guidelines (26). The selection was based on a keyword strategy synthetised in Table I.

 Table I. Boolean search and keyword strategy.

	Search Strategies
	Advanced keywords search:
Keywords	((dental AND (implant OR implants OR implantation OR
	implantology) AND (surface OR surfaces OR surface topography)
	AND (Histo*))
Databases	Pubmed/Medline, EMBASE, Cochrane Library

The papers' title and abstracts were assessed for an initial screening, and the manuscripts were limited to histological studies with bone-to-implant contact (BIC) outcomes. The full texts were finally collected and evaluated to assess the eligibility for the descriptive analysis.

Inclusion and exclusion criteria

The inclusion criteria for the eligibility synthesis were limited to histological studies that assessed bone-to-implant contact (BIC) outcomes from 1995 to today. The exclusion criteria were systematic and literature reviews, letters to the editor, *in vitro* and laboratory simulation, pilot studies, preliminary reports, no loading outcomes and early follow-up. The articles written in non-English language were excluded from the review.

RESULTS

Screening process

The electronic database identification process revealed a total of 3173 and 24 articles screened trough a manual search. A total of 482 duplicates have been removed from the articles list, and 2691 articles have been submitted for the full-text screening process. A total of 2527 papers were excluded for the following reasons: 1302 for the wrong outcome, 668 for the wrong device, 259 for wrong study design, 147 for wrong publication type, 101 written in a foreign language, 34 for wrong study duration and 16 for the wrong study population.

Sandblasted surfaces

The sandblasting procedure was proposed by sandblasting the metal surface with gritting agents. The number and rotations speed, the flux pressure, and the granulometry of the agent particles (10, 27) determine the treatment. The sandblasting procedure increases the surface irregularity and the implant biomechanical characteristics. The most common sandblasting agents are aluminium oxide/alumina (Al2O3) and titanium oxide (TiO2). The primary studies concerning the sandblasted surfaces are summarised in Table II. The procedure can influence the adhesion, proliferation, and differentiation of osteoblasts (20, 28).

Moreover, the fibroblasts result in a more difficult adhesion to the implant surface and a lower soft tissue proliferation around the implant in favour of the new bone formation (27, 29). Using surfaces blasted with Al2O3 particles was investigated compared to turned titanium surfaces. In the literature, the sandblasted implants showed higher BIC than the machined (30). In another study, the machined implants with Sa of 0.96 μ m were compared to different blasting sizes, and after 12 weeks, all blasted surfaces demonstrated higher BIC compared with machined surfaces.

The blasting procedure leaves residual particles over the implant's surface, which can modify the bone healing process. Some authors support that the presence of remaining particles may benefit osseointegration, catalysing this process (31); others support that aluminium ions are suspected to impair bone formation by a possible competitive action to calcium (32-35). TiO2 particle blasting was proposed to promote bone contact (27). Dental implants with TiO2 surface were compared to machined implants with a statistically significant higher removal torque compared to machined implants. No differences in BIC were detected (25). A combination of TiO2 blasted surface with fluoride ions has been proposed to improve the early osseointegration of dental implants (36). This method reported a bone-to-implant contact mean of >48% after 2 months of healing, which was higher than the blasting procedure alone (36).

At the same time, the precise nature of multinucleated giant cells is not thoroughly investigated, while a histological study suggested a priming effect on osteoblast activity similar to the hypothetic role of osteoclasts (37). Additional studies focus on sandblasted implants (38-40).

Plasma sprayed and plasma-chemical vapour surface

The plasma-sprayed treatments were studied in orthopaedics (41) and dental implants with no histological evidence of connective tissue infiltration at the interface level (42). Plasma-sprayed implants are obtained by spraying heat molten metal on the implant core, producing irregularly sized and shaped rounded particles and splats with valleys, pores and crevices (43). This treatment improves implant stability, bone growth (44), and higher surface contact area (10).

This treatment has been successfully investigated in rabbits (45), monkeys (46, 47), and humans (48-51), in different functional loading conditions (52). *In vivo*, no significant differences were detected between plasma-spray vs. machined implant, with a BIC percentage ranging between 55.9% and 56.2% (53). An alkali modification of the plasma-spraying

Author	Implant surface	Results	Findings	Experimental design
Piattelli et al. (1998) 30	(1) Al ₂ O ₃ blasted	BIC values	The blasted sites presented BIC	Implants inserted in the
	(2) Turned	(1) 60%±1.4% (2) 51%±1.9%	values statistically higher in comparison to turned.	femoral articulation of rabbits.
Piattelli et al. (1996) ³⁷	(1) Al ₂ O ₃ blasted (2) Turned (3) Plasma-spray	ACP, ALP activity (1), (2) and (3)	No MGS activity was reported for (1) and (2). At 2 weeks, Plasma spray revealed MGS	Healing period: 8 weeks Healing period: 2, 4 and weeks
Wennerberg et al. (1998) ³⁸	(1) Al₂O₃ blasted (25μm, 75μm, and 250μm particles)	BIC values (1) Ranging from 31 to 47%	Blasted surfaces demonstrated more bone in contact to implant surface compared to turned	Implants inserted in the tibia of rabbits.
	(2) Turned	(2) Ranging from 18 to 23%.	surface.	Healing period: 12 weeks
Wennerberg et al. $(1996)^{31}$	(1) Al ₂ O ₃ blasted (25 μm particles)	BIC values (1) 49.2 % (2) 47.6 %	No statistically different values concerning torque removal BIC values between the surfaces	Implants inserted in the tibia of rabbits.
	(2) TiO ₂ blasted (25 μm particles)	Removal	blasted with the same size of particles.	Healing period: 12 weeks
		(1) 26.5 Ncm (2) 24.9 Ncm		
Wennerberg et al. $(1995)^{25}$	(1) TiO ₂ blasted (25 μm particles)	BIC values (1) 40.9 % (2) 34 5 %	BIC values were not significantly different between the implants. However, TiO	Implants inserted in the tibia of rabbits.
	(2) Turned	(2) 34.3 % Removal torque (1) 35.4 Ncm (2) 29.2 Nom	blasted implants demanded a statistically significant greater removal torque force than turned implants.	Healing period: 12 weeks
Gotfredsen et al. (1992) ³⁹	(1) TiO ₂ blasted (10-53 μm particles)	Removal torque (1) 150 Ncm	BIC not significantly difference (data not shown), but, blasted implants presented higher	Implants were immediately placed, in dogs.
	(2) Turned	(2) 60 Ncm	removal torque values in comparison to turned sites.	No prosthetic rehabilitation was performed
Ivanoff et al. (2001) ⁴⁰	(1) TiO ₂ blasted (25 µm particles)	BIC values (1) 37 % (2) 9 %	The analysis of the results revealed a significantly higher BIC for the blasted implants	Healing period: 12 weeks Microimplants were inserted in the ridge of 27 patients.
	(2) Turned		than turned groups.	Mean healing period ranging from 3.9 to 6.3
Rocci et al. (2008) ³⁶	(1) TiO ₂ blasted (25 μm particles)	BIC values (1) 24.8 % (2) 48.3 %	The implant surfaces grit- blasted seems to produce a positive effect on	months. A total of 7 implants positioned in human
	(2) TiO ₂ blasted with fluoride ions		osseointegration, the adding of fluoride ions could produce a	mandible.
			sensible bioactive effect on the integration process.	Mean healing period 8 weeks.

 Table II. Comparative studies which used sandblasted implants.

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technique by sodium hydroxide solutions at 40°C for 24 h can determine an oxide layer $Ra = 17.6 \mu m$) and 20 nm thickness (44). The main reported disadvantage of plasma-spray is the detachment of titanium after implant insertion. Franchi et al. (54) reported the particle detachment of plasma spray, sandblasted and acid-etched, and machined implants in sheep. The authors reported that the titanium particles were detected only in plasma-sprayed implants. This phenomenon can be related to the friction between the implant surface and host bone cavity during implant placement, but its implications are unclear. Recent non-thermal and argon-based plasma applications have been proposed for dental implants, reporting no significant changes in new bone formation compared to sandblasted dental implant (55-58).

On the contrary, a significant increase in argon-based plasma-spray implant-bone contact was reported by Qiao et al. compared to sandblasted and acid-etched fixtures (59). Several studies increased the new bone formation of hybrid titanium-zirconia dental implants obtained through a novel plasma spray technique (60, 61). The microwave plasma-chemical-vapour deposition (MWP-CVD) of diamond-coated Ti-Al6-V4 dental implants compared to Ti-Al6-V4 implants have been investigated (62, 63). No differences in BICs, delamination, or particle-dissociation due to shearing forces have been detected (62).

Acid-etched surfaces

The acid-etch implant was proposed to avoid the residues released from sandblasting, a non-uniform surface modification of the implant body (10). For this purpose, different acid-etching solutions have been proposed, such as chloridic (HCl), sulfuric (H2SO4), hydrofluoric (HF), and nitric (HNO3), in different combinations. The acid-etching process effectiveness is by the baseline roughness, acid composition, temperature, and etching time. The histologic assessment results have been evaluated in Tables III and IV. A study compared two different etching of solution HCl and H2SO4, reporting that the surfaces presented a homogeneous distribution of small 1-2 μ m peaks and valleys and a removal torque 4 times higher for acid etched (6). The dual acid-etched procedure was proposed to obtain a macro- and micro-texture of the titanium surfaces (6) and higher platelet and osteogenic molecular signals (64, 65). Degidi et al. (66) reported histologically a mean BIC percentage of 61.3%, with no gaps or fibrous tissues present at the interface. Similar BIC results were reported after four months of healing on non-loaded implants (67).

In immediate loading protocols, the mean BIC levels ranged between 78% and 85% in vivo in humans (68). In the posterior maxilla after 6 months of healing, the BIC values of dual acid-etched sites were statistically higher than in turned sites (~70%) (69). Different acid concentrations were evaluated by Cho et al. (70), reporting a removal torque for dual acid-etched implants statically higher compared to the machined surface. The removal torque of 2mm diameters triple-etched micro-implants has been investigated by Pontes et al. (71), who reported an increase of the strength resistance >6Ncm after 8 weeks of healing. In a sheep study, Jinno et al. (72) reported that the dual-acid etch technique produces similar BIC findings to dual etching-sandblasting surfaces. Some authors associated the main findings for bone response to the dental implant macro-geometry (72-74). Halldin et al. reported that nano- and microtopography indicted by dual etching can potentiate the initial biomechanical behaviour, while for a more extended osseointegration period, the surface interlocking capacity seems more effective (75). On the contrary, several studies reported that the roughness scale seems to be effective for new bone formation (76, 77).

A similar outcome was reported by Yoo et al. that highlighted higher BICs and removal torque resistance of dual-acid etched implants compared to grit blasted/acid etch with low bone remodelling rates (78). Also, others obtained similar results (79).

Sandblasted and acid-etched surfaces

The combination of sandblasting and acid-etching technique has been suggested to produce uniform scattered gaps and hole distribution and slightly less rough than the plasma-sprayed surface, which is characterised by profoundly irregular micro-texture and less favourable substrate for cell proliferation (80). The histological studies have been summarised in Tables V and VI. Higher torque removal values of sandblasted/acid-etch surfaces have been reported (+75%-125%) compared to acid-etched implants (81). Abrahamsson et al. (82) reported that the BIC values in dogs were significantly higher in sandblasted/acid-etched implants compared to machined surfaces. Similar results were observed in the comparative evaluation of sandblasted/acid-etched compared in plasma-sprayed implants(83). Sandblasted and acid-

Author	Surface treatment	Results	Findings	Experimental design
Klokkevold	(1) Acid-etched	Removal torque	The resistance to torque	Implants were inserted in
et al.	$(HC1 / H_2SO_4)$	(1) 20.50 Ncm	removal was 4 times greater	the femur of rabbits.
$(1997)^{6}$		(2) 4.95 Ncm	for acid etched implants in	
	(2) Turned		comparison to the turned	Healing period: 2 months
			surfaces.	
Cho et al. $(2002)^{70}$	(1) Acid-etched	Removal torque	Dual acid etched implants	Implants were inserted in
$(2003)^{70}$	(HF and HCl /	$(1) 34.7 \text{ Ncm}^*$	required a higher removal	the tibia of rabbits.
	H_2SO_4)	(2) 15.2 Ncm	torque average force than the	
	(1) T		turned surface implants.	Healing period: 12 weeks
Wang at al	(2) Lurned (1) Asid stabad	DIC values	PIC values were	Implants wars inserted in
200267	(1) Actu-etcheu (Ossootite [®])	(1) 62 5 9/	bic values were	aroos with poor hope quality
2003	(Osseottie ⁺)	(1) 02.3 % (2) 20 5 %	significantly night in dual	in the mandible of dogs
	() Turned	(2) 39.3 /0	comparison to turned sites	In the manufole of dogs.
	(2) fulled $(ICF^{\mathbb{R}})$		comparison to turned sites.	Healing period: 1 months
Klokkevold	(1) Acid-etched	Removal torque	Statistically significant	Implants were inserted in
et al	$(HC1/H_2SO_4)$	(1) 27 40 Ncm	differences were observed	the femur of rabbits
$(2001)^{79}$	(11017 112004)	(2) 59 23 Ncm	between acid-etched and	the femal of fusions.
(2001)	(2) Plasma-sprav	(3) 6 73 Ncm	turned implants and	Healing period 3 months**
	(=) I monin sprag		between plasma-spraved and	freeding period. 5 months
	(3) Turned		turned implants.	
			However, differences	
			between acid etched and	
			plasma-sprayed were not	
			statistically different.	
Pontes et al	(1) Triple Acid-	Removal torque	The triple acid etching can	Healing period: 8 weeks
(2015) 71	etched	(1) 3.3 ± 1.7 Ncm	create a promising and	Implants were inserted in
		(2) 2.2 ± 1.3 Ncm	efficient surface for the	rats.
		$(3) 6.7 \pm 1.4$ Ncm	process of osseointegration.	
Rezende de	(1) Acid-etched	BIC values	Bone-to-implant contact and	Implants were inserted in
Jesus et al.	(2) Sandblasted and	2 weeks	BD increased with time in	dogs
(2017) ⁷³	Acid-etched	(1) $19.57 \pm 13.57\%$	both surface treatments	Healing period: 2 and 4
		$(2) 20.33 \pm 7.99\%$	implants	weeks.
		4 weeks		
		$(1) 40.25 \pm 9.45\%$ (2) 42.80 + 4.48%		
		(2) 42.80± 4.48%		
Carr et al	(1) Plasma-snrav	BIC values	No significant differences	Implants were inserted in
$(2000)^{53}$	(1) I lasina-spi ay	(1) 55.9%	could be observed between	the mandible of baboons
(2000)	(2) Turned	(2) 56.2 %	groups concerning the BIC	
	()	(-) , -	percentage.	No prosthetic rehabilitation
			1 C	was performed.
				Healing period: 6 months.
Carr et al. (2000) ⁵³	(1) Plasma-spray (2) Turned	BIC values (1) 55.9 % (2) 56.2 %	No significant differences could be observed between groups concerning the BIC percentage.	Implants were inserted in the mandible of baboons. No prosthetic rehabilitation was performed. Healing period: 6 months.

 Table III. Comparative studies which used acid etched and plasma-sprayed implants.

etched surfaces reported increased osteoconductive cell proliferation characteristics compared to plasma-spray implants (80, 84, 85). The histological findings of sandblasted/acid-etched reported after six months of healing in humans a mean BIC of 76.6 % (86). After 40 months, a 75.4 % BIC mean was observed on retrieved human implants (87).

Some studies reported sufficient bone volume and density that sandblasted/acid-etched surfaces can present a success rate of 99 % after two years (88). The combination of acid-etching and ZrO2 particles sandblasting produces an increased bone deposition compared to plasma-sprayed and machined implants (54). Several authors reported that the depth and distribution of irregularities, the cavity morphology, and contaminating elements derived from the treatment procedures

Author	Surface treatment	Results	Findings	Experimental design
Testori et al. (2001) ⁶⁸	Acid etched (Osseotite [®])	BIC values ranging from 78% to 85%	Implants were successfully used in immediately loaded protocol.	Histologic analysis of two retrieved immediately loaded implants.
Degidi et al. (2003) ⁶⁶	Acid etched (HCl and H ₂ SO ₄)	Mean BIC value 61.3%	No gaps or fibrous tissues were observed at the interface.	Healing period: 4 months. Histologic analysis of two retrieved implants.
Trisi et al. (2002) ⁶⁹	(1) Acid etched (Osseotite [®])	BIC values	BIC values in dual acid- etched sites were	No prosthetic rehabilitation was performed. Healing period: 6 months. Histologic analysis of implants inserted in the
	(2) Turned	(1) 72.35 % (2) 35.32 %	statistically higher than in turned sites.	posterior maxilla of 11 patients. Healing period: 6 months.

Table IV. Histologic studies in which acid etched implants were retrieved from humans.

play an important role in cell behaviour (89). In different animal study models, the sandblasted and acid-etched surfaces seem to produce in animals very similar BICs (~60%) compared to RBM, acid treatments and micro-arc procedures with no significant differences (90-96). At the same time, Marinho et al. reported a significantly higher new bone contact compared to the comparison of machined implant surfaces (97). Similar results were obtained by Buser et al. (98).

Nodized surfaces and micro-arc treatment

The oxidation technique has been proposed to modify the oxide layer properties and the surface biocompatibility (99), avoiding the deposit of grit particles (100). The anodised surfaces are obtained by a voltage application on the titanium surface in an electrolyte bath. The treated surface appeared with micro-pores of variable diameters without cytotoxicity (101). The removal torque of different thicknesses of anodised surfaces was investigated, which was significantly higher than that of smooth surfaces (99).

In the rabbit model, anodised, and sed and hydrothermally treated, and machined implants were investigated, reporting BIC values ranging between 40% and 50% and removal torque differences between the study groups (102). Authors reported that differentiation and calcification occurred on rough and smooth surfaces, indicating that the porous microstructure could enhance cell proliferation (43). In literature, it was demonstrated that the voltage for the anodising technique could produce a sensible influence on osseointegration properties, while the optimal value seems to be at \sim 550 V (103). In this way, the micro-roughness generated by anodic oxidation seems to significantly ameliorate BICs compared to sandblasted surfaces (104, 105) and machined implants (106). Moreover, using a super-hydrophilic surface of anodic oxidation implants has been proposed to potentiate this histological finding (107), while using biologically-derived triterpenoids adjuvant coating seems to produce no significant effect on this parameter 108).

In addition, the electrochemical anion sulphuric acid and phosphoric acid incorporation significantly affect BICs with an increase of $\sim 200\%$ histological bone contact (109). The micro-arc surface oxidation treatment has been proposed to improve the titanium dental implant. The biocompatibility of micro-arc oxidation has been tested by several authors, producing an acceleration and enhancement of the fixture's osseointegration (90, 110–113). Dundar et al. (90) reported similar BIC means ($\sim 60\%$) comparing different surfaces RBM, SLA, micro-arc, and sandblasted-micro-arc treatment with no significant difference.

Hydroxyapatite-coated surfaces and ceramic-coating implants

Hydroxyapatite implants have been studied to improve bone-implant fixation due to an increased osteoblast activity to this contact and adhesion, proliferation, and differentiation (114). Histological findings of hydroxyapatite implant

Author Su	urface treatment	Results	Findings	Experimental design
Abrahamsson (1)) Sand-blasted and	-	BIC values (data not shown)	Implants were inserted in
et al. aci	id-etched		were significantly greater in	the mandible of dogs.
$(2004)^{82}$			sandblasted and acid-etched	
(2)) Turned		sites than in turned surfaces	No prosthetic rehabilitation
				was performed.
				Healing period: 1, 2, 4, 6, 8
Marinha at (1)	Sand blastad and		The SLA surfaces revealed	Implants were inserted in
(1)	j Sanu-Diasteu anu	-	a higher hone response vs	rats
al. (2003) aci	iu-ciciicu		machined surfaces	Healing period: 5, 15, 30
(2)) Turned		indefinied surfaces.	and 60 days
Coelho et al (1)) alumina-blasting	BIC volues	No significant differences of	Implants were inserted in
$(2011)^{57}$ (2)) biologic blasting	(1) 40 13 \pm 2 54%	BIC were detected at 4	dogs.
(3)) plasma	$(1) + 0.13 \pm 2.3 + 70$ (2) 37 23 ± 2 14%	weeks. An higher reoval	Healing period: 4, weeks.
(4)) microblasted RBM	(3) $38.56 \pm 2.49\%$	torque was detected for	
(5)) Sand-blasted and	$(4) 39.65 \pm 2.27\%$	RBM implants.	
aci	id-etched (AB/AE)	(5) 38.72±1.44%		
Cochran et (1)) Sand-blasted and	BIC values	The sandblasted and acid	Implants were inserted in
al. (1998) ³⁴ aci	Id-elched	(1) 71.68 %	cignificantly greater DIC	the mandible of dogs.
(23	rticles and etched	(2) 58.88 %	percentage than did the	Loading period: 12 months
wit	th HCl / H ₂ SO ₄)		plasma-spraved However	Healing period: 15 months
	un men / m2004)		no qualitative differences in	fieuning period. To months
(2)) Plasma-spraved		bone tissue were observed	
	, .		between groups.	
Buser et al. (1)) Sandblasted and	Removal torque	Statistically significant	Implants were inserted in
(1999) ⁹⁸ aci	id-etched	(1) 1.43 Ncm	differences were observed	the maxilla of miniature
(0.1	.25–0.50 μm particles,	(2) 1.54 Ncm	between sandblasted and	pigs.
etc	ched with HCl /	(3) 0.26 Ncm	acid-etched and turned	
H ₂	SO ₄)		implants, and between	No prosthetic rehabilitation
	DI		plasma-sprayed and turned	was performed.
(2)) riasma-sprayed		impiants.	Healing period: 12 weeks*
) Turned		however, unterences	
(3)	j i ui litu		acid etched and plasma-	
			sprayed were not	
			statistically different	

Table V. Comparative studies that used sandblasted and acid-etched implants.

*Data from the 1st and 2nd healing periods were not included in this table.

Table V	VI. Histologic	studies in	ı which	sandblasted	and	acid-etched	implant	s were	retrieved	from	humans
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Author	Surface treatment	Results	Findings	Experimental design
Hayakawa et al. (2002) ⁸⁶	Sandblasted and acid-etched (Straumann [®])	BIC value 76.6 %	Bone surrounding the implant was uniformly and maturely structured.	Histologic analysis of one retrieved implant that was inserted in the palatal bone of the maxilla of a patient as anchorage for orthodontic treatment.
Sakakura et al. (2005) ⁸⁷	Sandblasted and acid-etched	BIC value 75.4 %	The surrounding bone healed in a well-organized pattern and could not be differentiated from the original alveolus.	Healing period: 6 months Histologic analysis of one retrieved implant of a patient. Loading period: 40 months
an. (2003)	and actu-ctened	/3.4 %	pattern and could not be differentiated from the original alveolus.	Loading period: 4(

indicated a BIC range between 87.5%-97.4% (115). This coating technique reported high survival rates at medium- and long-term follow-ups (2, 116, 117). After 12 years of loading, the survival rate of hydroxyapatite implants was 93.2%, statistically increasing compared to titanium implants (2). After 10 years of loading, the hydroxyapatite implants reported a BIC range between 70.74%-86.23% (118).

Piattelli et al. (119) reported a localised chronic suppurative bone infection associated with peri-implantitis in a hydroxyapatite-coated implant, where the coating appeared detached from the titanium surface. Different methods can be used for hydroxyapatite coating, such as coating/sintering, electrophoretic deposition, immersion coating, hot isostatic pressing, solution deposition, sputter coating, and thermal spraying techniques (120). Hydroxyapatite plasma-spraying was indicated to combine the hydroxyapatite characteristics and the bone-implant mechanical interlock associated with the plasma-spraying procedure. Higher BIC values were reported for hydroxyapatite implants than titanium plasma-spray implants and machined fixtures (121). The Resorbable Blast Material (122) also known as the technique of ion-beam-assisted deposition (IBAD) (123, 124) has been proposed to improve the coating quality properties.

In vivo, the BIC values were significantly higher in IBAD surfaces compared to blasted and machined implants. The authors suggested that the advantages of the HA-coated implants in the early healing period could be apparent, while the separation or fracture of the coating layer could be prevented. However, the resorption needs to be further investigated123. Svanborg et al. (125) investigated different hydroxyapatite (HA) nanocoating thicknesses on titanium grade Ti-6A1-4V implants of 15 mm in length and 3.85 mm in diameter in rabbits.

The single layer-HA coating reported a mean Sa 0.91 (0.20) μ mm while the double layer-HA coating showed a mean Sa 0.77 (0.19) μ mm125. After 9 weeks of healing, the single layer-HA coating reported higher values of removal torque (p<0.05) and at 2 weeks reported an increase of almost 5% of new bone formation compared with the control and the double layer-HA coating. After 9 weeks, the BIC for both groups was similar (~60%) (125). The advantage of ceramic-coating implants has been described due to the high osteoconductivity of the surfaces, while these techniques can produce a surface biofunctionalisation that can increase the implant osseointegration (126-167).

The surface functionalisation seems to maintain the implant roughness, while Jimbo et al. reported no significant differences between the smooth bioceramic surface and the rough bioceramic coated implants (142). In addition, other studies investigated different ceramic coatings such as calcium carbonate, ceramic brushite, glass fibres, phosphate-containing polymers, magnesium-containing polymers, and calcium-phosphate (126-167). Granato et al. investigated the coating thickness and demonstrated that the optimal Ca- and P-derived bioceramic coating layer ranged between 300-500

Author	Surface treatment	Results	Findings	Experimental design
Sul et al.	(1) Anodized	Removal torque	The preliminary results	Implants were inserted in
$(2002)^{99}$	(oxide thickness	(1) Ranging from	of this study suggest that	the tibia of rabbits.
	approximately 200, 600,	0.113 to 0.129	the oxide thickness	
	800 or 1000 nm)	Nm	influence the bone tissue formation.	Healing period: 6 weeks
	(2) Turned	(2) 0.075 Nm		
	(oxide thickness: 17.4 nm)			
Son et al.	(1) Anodized	Removal torque	Difference between	Implants were inserted in
$(2003)^{*102}$.		(1) 51.35 Ncm	groups was not	the tibia of rabbits.
	(2) Turned	(2) 35.28 Ncm	statistically significant	
			concerning removal	Healing period: 12 weeks
			torque and BIC values	
			(data not shown).	
Ivanoff et al	(1) Anodized	BIC values	BIC values were	Histologic analysis of
$(2003)^{113}$		(1) 34 %	statistically higher in	implants inserted in the
	(2) Turned	(2) 13 %	oxidized than in turned	ridge of 20 patients.
			sites.	
				Mean healing period: 6.6
				months

Table VII. Comparative studies which used anodized implants

* Data from the 1st healing period, and an experimental group were not included in this table.

nm (150). Moreover, the fluorapatite and heated-hydroxyapatite coatings present a decreased respiration rate compared to hydroxyapatite implant surfaces (167).

Thermal oxidation and heat surface treatment

The investigation of innovative procedures able to contrast surface wearing and successful bioactivity and osseointegration represents the current breakthrough in implantology. Thermal oxidation aims to create a highly crystalline oxide coating able to potentiate the interaction between the titanium surface and the host surrounding bone (168, 169). A 700°C exposure for 1 hour by a controlled furnace of Ti6Al4V alloy can induce the formation of a rutile oxide layer that could improve the osteoblast attachment on the implant surface in vitro (170). In addition, the heat treatment at 800°C in the air for 1 minute also seems to increase the BICs in vivo of acid significantly etch Ti6Al4V implants (171). The Al obtained similar results (2) (3) abrasive particle blasting with thermochemical treatment in minipigs compared to SLA (shot blasting surface) (172).

Quameya et al. reported that adding a supplemental fluoridic acid etch to the thermally oxidised surface did not significantly affect osseointegration compared to standard SLA surface implant (173). The heat-derived oxide layer has been studied by Kim et al. (174), which compared different oxide layer thicknesses of 20nm to 80nm and the additional treatment of CaP coating. The same authors detected no significant differences in BICs and ISQ at 5 weeks on dogs (174).

Zirconia implants and acrylic materials

Zirconia (zirconium oxide, ZrO2) is a ceramic material purposed as dental implant material due to its biocompatibility, esthetic properties, and mechanical behaviour, which are better than alumina (60, 61, 175-188). Zirconia is reported to present a bone contact similar to titanium implants189,190. The interface is composed of a proteoglycan layer that is thicker than titanium (191, 192). Zirconia implants are biocompatible, bioinert, and radiopaque, with high corrosion and wearing resistance, flexion and fracture (193-197).

In rabbits, the BIC value of zirconia implants was 68.4% after 4 weeks with no foreign bone reaction and fibrous tissue infiltration at the level of the interfaces (198). Loaded zirconia implants were evaluated in monkeys, with BIC values ranging between 66%-81% (199). Zirconia implants submitted to Al2O3 sandblasting were compared to titanium (Al2O3 sandblasting followed by H2O2 and HF etching reporting BIC values of 67.4% for zirconia, and 72.9% for titanium surfaces with no statistically significant differences (190). Various types of zirconia implants have been investigated in the literature, while the most investigated are yttria-stabilised tetragonal zirconia polycrystalline (3Y-TZP) and ceria-stabilised zirconia-alumina nanocomposite (NanoZr) (176).

Mijhatovic et al. investigated three different roughnesses of zirconia implants compared to sandblasted large grit and acid-etched titanium implants, showing no significant differences in total BICs after 10 weeks on dogs. The hybrid hydrophilic titanium-zirconium alloy (TiZr1317) revealed a lower removal torque at 2 weeks compared to standard titanium implants, while no differences were detected at 4 and 12 weeks. At 4 weeks, hybrid hydrophilic titaniumzirconium alloy (TiZr1317) showed significantly higher BICs in the marrow area of 19.25% (179). Very few studies investigated in vivo the properties of plastic and acrylic resin implants (200). Okamatsu et al. (200) studied the hybrid titanium-plastic implants and evaluated a homogeneous 150- to 250-nm acrylic layer coating. The authors reported new bone formation in the test and control groups, with no direct bone contact with the plastic implant.

UV and biologically functionalised surfaces

In literature, photodynamically functionalised implant surfaces have been investigated (201-204). Mehl et al. (201) reported no significant differences between BICs and ISQ in a split-mouth study model using a high-energy UV-irradiation in epicrestally titanium implants. On the contrary, a significant increase in removal torque and BICs was reported by Shen et al. (203). The authors evaluated a different combination of SLA-surfaces treated by UV-bactericidal irradiation at 15-20W, 0.1mW/Cm2 and 0.2mW/Cm2 (203). The UV photo-functionalisation seems effective, especially in the early phases of osseointegration (202), with a significant increase in bone contact and dental implant stability. This treatment seems to take a significant advantage when combined with biologically functionalised treatment with fibronectin and osteopontin (204, 205) due to a significant increase in the hydrophilicity of the surface.

DISCUSSION

Several authors investigated the biological properties of dental implant surfaces under *in vitro* conditions. At the same time, this kind of research is consistent in investigating the specific cell response, the clinical relevance of these results is discussible, and the development of long-term clinical evaluations is fundamental. Different implant topographies seem to influence the outcome of dental implants, but the magnitude and clinical relevance of this influence are still being investigated.

On the other hand, many studies are being published to investigate the viability of modified surfaces. Regarding the titanium alloy, a study performed in rabbits reported that the removal torque was statistically different after 6 months and 12 months, where the cpTi implants were significantly more stable. The BIC means presented no significant differences between the materials (206). In another investigation, cpTi and Ti6Al4V dental implants were positioned in baboons, reporting that BIC means were significantly higher in cpTi and Ti6Al4V implants, but differences after six months were not significantly different (207).

Even if the use of the alloy represents a mechanical advance compared to cpTi, biomechanical tests revealed that cpTi presented an increased stability. Moreover, the titanium implants, after the air exposure, can form an oxide layer all over the surface of 2–5 nm thickness. The oxide layer (208, 209) plays a key role in corrosion resistance, biocompatibility and implant osseointegration (210-212). The layer is mainly formed by TiO2 (213), and the crystalline structure, the thickness and stability of this layer varies according to the surfaces of the implant (99, 214, 215).

Promising findings for dental implants concerning nitride titanium (TiN), nanostructured texture, laser-treated surfaces, and ceramic materials have been recently reported (188, 216). The nanostructured surfaces (1-100nm) could improve the early interface and bone-implant contact (217, 218). Authors reported that dogs presented a higher percentage of newly formed bone in contact with nanostructured implants than plasma-spray and machined implants (219, 220), and BIC values ranged between 55 and 96% in humans (221).

Nitride titanium (TiN) was proposed to produce a surface less susceptible to the ions release. For this purpose, the physical vapour deposition technique can produce a thin TiN layer ($\sim 1\mu$) for an osseointegration quality similar to standard titanium implants. This layer increases corrosion resistance, lower bacterial adhesion, and a golden aspect of the implant surface (222-227). The laser ablation is a reproducible procedure for a controlled, micron-sized surface with topographical features on the flanks of the threads. Lasered implants demonstrated significantly higher BIC and removal torque peaks than machined implants (228, 229). Calcium phosphate and ceramic coating are correlated to a high chemical bonding property, similar to hydroxyapatite (86). Biphasic calcium phosphate (162, 230, 231) or tricalcium phosphates have been investigated as implant coating (232, 233).

In conclusion, proper long-term studies have been published for TiO2 surfaces, but other surfaces are documented with a medium-term follow-up period (234). While clinicians should consider that several new treatment surfaces are constantly purposed and currently available in the market, long-term findings are necessary to comprehend their long-term biological response.

REFERENCES

- Brånemark PI, Breine U, Adell R, Hansson BO, Lindström J, Ohlsson Å. Intra-Osseous Anchorage of Dental Prostheses: *I. Experimental Studies. Scandinavian Journal of Plastic and Reconstructive Surgery*. 1969;3(2):81-100. doi:10.3109/02844316909036699
- Schwartz-Arad D, Herzberg R, Levin L. Evaluation of Long-Term Implant Success. *Journal of Periodontology*. 2005;76(10):1623-1628. doi:10.1902/jop.2005.76.10.1623
- 3. Davies JE. Understanding peri-implant endosseous healing. Journal of Dental Education. 2003;67(8):932-949.
- Albrektsson T, Br\aanemark PI, Hansson HA, Lindström J. Osseointegrated titanium implants: requirements for ensuring a longlasting, direct bone-to-implant anchorage in man. Acta Orthopaedica Scandinavica. 1981;52(2):155-170.
- 5. Albrektsson T, Johansson C. Quantified bone tissue reactions to various metallic materials with reference to the so-called osseointegration concept. *The bone-biomaterial interface University of Toronto Press, Toronto*. Published online 1991:357-363.
- 6. Klokkevold PR, Nishimura RD, Adachi M, Caputo A. Osseointegration enhanced by chemical etching of the titanium surface. A

torque removal study in the rabbit. Clin Oral Implants Res. 1997;8(6):442-447. doi:10.1034/j.1600-0501.1997.080601.x

- Wennerberg A, Albrektsson T. Suggested guidelines for the topographic evaluation of implant surfaces. *International Journal of* Oral & Maxillofacial Implants. 2000;15(3).
- 8. Jalbout Z, Tabourian G. Glossary of Implant Dentistry. International College of Oral Implantologists; 2004.
- 9. Lautenschlager EP, Monaghan P. Titanium and titanium alloys as dental materials. International dental journal. 1993;43(3):245-253.
- Scarano A, Piattelli M. Superfici implantari. In: Novello G. Implantologia pratica. Coordenons: New Service International S.r.l. 1a ed. 2005, p. 21-32.
- 11. Lincks J. Response of MG63 osteoblast-like cells to titanium and titanium alloy is dependent on surface roughness and composition. *Biomaterials*. 1998;19(23):2219-2232. doi:10.1016/S0142-9612(98)00144-6
- 12. Hanawa T. Titanium-Tissue Interface Reaction and Its Control With Surface Treatment. *Front Bioeng Biotechnol*. 2019;7:170. doi:10.3389/fbioe.2019.00170
- 13. Cai K, Frant M, Bossert J, Hildebrand G, Liefeith K, Jandt KD. Surface functionalised titanium thin films: zeta-potential, protein adsorption and cell proliferation. *Colloids Surf B Biointerfaces*. 2006;50(1):1-8. doi:10.1016/j.colsurfb.2006.03.016
- Scarano A, Piattelli A, Polimeni A, Di Iorio D, Carinci F. Bacterial adhesion on commercially pure titanium and anatasecoated titanium healing screws: an in vivo human study. *Journal of Periodontology*. 2010;81(10):1466-1471. doi:10.1902/ jop.2010.100061
- 15. Scarano A, Tripodi D, Carinci F, Piccolomini R, D'Ercole S. Biofilm formation on titanium alloy and anatase-Bactercline® coated titanium healing screws: An in vivo human study. *Journal of Osseointegration*. 2013;5(1):8-12.
- Kasemo B. Biocompatibility of titanium implants: Surface science aspects. *The Journal of Prosthetic Dentistry*. 1983;49(6):832-837. doi:10.1016/0022-3913(83)90359-1
- Thomas KA, Kay JF, Cook SD, Jarcho M. The effect of surface macrotexture and hydroxylapatite coating on the mechanical strengths and histologic profiles of titanium implant materials. *J Biomed Mater Res.* 1987;21(12):1395-1414. doi:10.1002/ jbm.820211205
- Chehroudi B, Gould TRL, Brunette DM. Effects of a grooved titanium-coated implant surface on epithelial cell behaviorin vitro andin vivo. J Biomed Mater Res. 1989;23(9):1067-1085. doi:10.1002/jbm.820230907
- 19. Brunette DM. The effects of implant surface topography on the behavior of cells. *International Journal of Oral & Maxillofacial Implants*. 1988;3(4).
- Bowers KT, Keller JC, Randolph BA, Wick DG, Michaels CM. Optimisation of surface micromorphology for enhanced osteoblast responses in vitro. *Int J Oral Maxillofac Implants*. 1992;7(3):302-310.
- Boyan BD, Batzer R, Kieswetter K, et al. Titanium surface roughness alters responsiveness of MG63 osteoblast-like cells to 1?,25-(OH)2D3. *J Biomed Mater Res.* 1998;39(1):77-85. doi:10.1002/(SICI)1097-4636(199801)39:1<77::AID-JBM10>3.0.CO;2-L
- Kieswetter K, Schwartz Z, Hummert TW, et al. Surface roughness modulates the local production of growth factors and cytokines by osteoblast-like MG-63 cells. *J Biomed Mater Res.* 1996;32(1):55-63. doi:10.1002/(SICI)1097-4636(199609)32:1<55::AID-JBM7>3.0.CO;2-O
- Doillon CJ, Silver FH, Berg RA. Fibroblast growth on a porous collagen sponge containing hyaluronic acid and fibronectin. *Biomaterials*. 1987;8(3):195-200. doi:10.1016/0142-9612(87)90063-9
- 24. Martin JY, Schwartz Z, Hummert TW, et al. Effect of titanium surface roughness on proliferation, differentiation, and protein synthesis of human osteoblast-like cells (MG63). *J Biomed Mater Res.* 1995;29(3):389-401. doi:10.1002/jbm.820290314
- 25. Wennerberg A, Albrektsson T, Andersson B, Krol JJ. A histomorphometric and removal torque study of screw-shaped titanium implants with three different surface topographies. *Clin Oral Implants Res.* 1995;6(1):24-30. doi:10.1034/j.1600-0501.1995.060103.x
- 26. Hutton B, Salanti G, Caldwell DM, et al. The PRISMA extension statement for reporting of systematic reviews incorporating network meta-analyses of health care interventions: checklist and explanations. *Ann Intern Med.* 2015;162(11):777-784.

doi:10.7326/M14-2385

- 27. Wennerberg A. Experimental study of turned and grit-blasted screw-shaped implants with special emphasis on effects of blasting material and surface topography. *Biomaterials*. 1996;17(1):15-22. doi:10.1016/0142-9612(96)80750-2
- Schwartz Z, Martin JY, Dean DD, Simpson J, Cochran DL, Boyan BD. Effect of titanium surface roughness on chondrocyte proliferation, matrix production, and differentiation depends on the state of cell maturation. *J Biomed Mater Res.* 1996;30(2):145-155. doi:10.1002/(SICI)1097-4636(199602)30:2<145::AID-JBM3>3.0.CO;2-R
- 29. Abron A, Hopfensperger M, Thompson J, Cooper LF. Evaluation of a predictive model for implant surface topography effects on early osseointegration in the rat tibia model. *The Journal of Prosthetic Dentistry*. 2001;85(1):40-46. doi:10.1067/mpr.2001.112415
- Piattelli A, Manzon L, Scarano A, Paolantonio M, Piattelli M. Histologic and histomorphometric analysis of the bone response to machined and sandblasted titanium implants: an experimental study in rabbits. *Int J Oral Maxillofac Implants*. 1998;13(6):805-810.
- 31. Wennerberg A, Albrektsson T, Andersson B. Bone tissue response to commercially pure titanium implants blasted with fine and coarse particles of aluminum oxide. *Int J Oral Maxillofac Implants*. 1996;11(1):38-45.
- 32. Blumenthal NC, Cosma V. Inhibition of apatite formation by titanium and vanadium ions. *Journal of biomedical materials research*. 1989;23(S13):13-22.
- Savarino L, Cenni E, Stea S, et al. X-ray diffraction of newly formed bone close to alumina- or hydroxyapatite-coated femoral stem. *Biomaterials*. 1993;14(12):900-905. doi:10.1016/0142-9612(93)90131-K
- 34. Toni A, Lewis CG, Sudanese A, et al. Bone demineralisation induced by cementless alumina-coated femoral stems. *The Journal of Arthroplasty*. 1994;9(4):435-444. doi:10.1016/0883-5403(94)90055-8
- 35. Darvell BW, Samman N, Luk WK, Clark RKF, Tideman H. Contamination of titanium castings by aluminium oxide blasting. *Journal of Dentistry*. 1995;23(5):319-322. doi:10.1016/0300-5712(94)00003-X
- 36. Rocci M, Rocci A, Martignoni M, Albrektsson T, Barlattani A, Gargari M. Comparing the TiOblast and Osseospeed surfaces. Histomorphometric and histological analysis in humans. *Oral Implantol (Rome)*. 2008;1(1):34-42.
- Piattelli A, Scarano A, Corigliano M, Piattelli M. Presence of multinucleated giant cells around machined, sandblasted and plasma-sprayed titanium implants: a histological and histochemical time-course study in rabbit. *Biomaterials*. 1996;17(21):2053-2058. doi:10.1016/0142-9612(96)00052-x
- Wennerberg A, Hallgren C, Johansson C, Danelli S. A histomorphometric evaluation of screw-shaped implants each prepared with two surface roughnesses: A histomorphommetric evaluation of screw-shaped implants. *Clinical Oral Implants Research*. 1998;9(1):11-19. doi:10.1034/j.1600-0501.1998.090102.x
- Gotfredsen K, Nimb L, Hjörting-hansen E, Jensen JS, Holmén A. Histomorphometric and removal torque analysis for TiO 2 hyphen;blasted titanium implants. An experimental study on dogs.: Torque analysis for TiO 2 -blasted implants. *Clinical Oral Implants Research*. 1992;3(2):77-84. doi:10.1034/j.1600-0501.1992.030205.x
- Ivanoff CJ, Widmark G, Hallgren C, Sennerby L, Wennerberg A. Histologic evaluation of the bone integration of TiO 2 blasted and turned titanium microimplants in humans: Bone integration of titanium implants. *Clinical Oral Implants Research*. 2001;12(2):128-134. doi:10.1034/j.1600-0501.2001.012002128.x
- 41. Hahn H, Palich W. Preliminary evaluation of porous metal surfaced titanium for orthopedic implants. *J Biomed Mater Res.* 1970;4(4):571-577. doi:10.1002/jbm.820040407
- 42. Schroeder A, van der Zypen E, Stich H, Sutter F. The reactions of bone, connective tissue, and epithelium to endosteal implants with titanium-sprayed surfaces. *Journal of Maxillofacial Surgery*. 1981;9:15-25. doi:10.1016/S0301-0503(81)80007-0
- Sammons RL, Lumbikanonda N, Gross M, Cantzler P. Comparison of osteoblast spreading on microstructured dental implant surfaces and cell behaviour in an explant model of osseointegration: A scanning electron microscopic study. *Clinical Oral Implants Research*. 2005;16(6):657-666. doi:10.1111/j.1600-0501.2005.01168.x
- 44. Xue W, Liu X, Zheng X, Ding C. In vivo evaluation of plasma-sprayed titanium coating after alkali modification. *Biomaterials*. 2005;26(16):3029-3037. doi:10.1016/j.biomaterials.2004.09.003
- 45. Piattelli A, Scarano A, Corigliano M, Piattelli M. Effects of alkaline phosphatase on bone healing around plasma-sprayed titanium

implants: a pilot study in rabbits. Biomaterials. 1996;17(14):1443-1449. doi:10.1016/0142-9612(96)87288-7

- Piattelli A, Corigliano M, Scarano A, Costigliola G, Paolantonio M. Immediate Loading of Titanium Plasma-Sprayed Implants: An Histologic Analysis in Monkeys. *Journal of Periodontology*. 1998;69(3):321-327. doi:10.1902/jop.1998.69.3.321
- Scarano A, Iezzi G, Petrone G, Marinho VC, Corigliano M, Piattelli A. Immediate Postextraction Implants: A Histologic and Histometric Analysis in Monkeys. *Journal of Oral Implantology*. 2000;26(3):163-169. doi:10.1563/1548-1336(2000)026<0163:IP IAHA>2.3.CO;2
- Piattelli A, Corigliano M, Scarano A. Microscopical observations of the osseous responses in early loaded human titanium implants: a report of two cases. *Biomaterials*. 1996;17(13):1333-1337. doi:10.1016/S0142-9612(96)80011-1
- Piattelli A, Paolantonio M, Corigliano M, Scarano A. Immediate Loading of Titanium Plasma-Sprayed Screw-Shaped Implants in Man: A Clinical and Histological Report of Two Cases. *Journal of Periodontology*. 1997;68(6):591-597. doi:10.1902/ jop.1997.68.6.591
- Piattelli A, Scarano A, Piattelli M, Bertolai R, Panzoni E. Histologic aspects of the bone and soft tissues surrounding three titanium non-submerged plasma-sprayed implants retrieved at autopsy: a case report. *J Periodontol*. 1997;68(7):694-700. doi:10.1902/ jop.1997.68.7.694
- 51. Piattelli A, Scarano A, Dalla Nora A, De Bona G, Favero GA. Microscopical features in retrieved human Branemark implants: a report of 19 cases. *Biomaterials*. 1998;19(7-9):643-649. doi:10.1016/S0142-9612(97)00158-0
- 52. Piattelli A, Corigliano M, Scarano A, Quaranta M. Bone reactions to early occlusal loading of two-stage titanium plasma-sprayed implants: a pilot study in monkeys. *Int J Periodontics Restorative Dent*. 1997;17(2):162-169.
- Carr AB, Gerard DA, Larsen PE. Histomorphometric analysis of implant anchorage for 3 types of dental implants following 6 months of healing in baboon jaws. *Int J Oral Maxillofac Implants*. 2000;15(6):785-791.
- Franchi M, Bacchelli B, Martini D, et al. Early detachment of titanium particles from various different surfaces of endosseous dental implants. *Biomaterials*. 2004;25(12):2239-2246. doi:10.1016/j.biomaterials.2003.09.017
- 55. YW H, HL C, LT L, KC T, DT B, YK W. Effects of non-thermal plasma on sandblasted titanium dental implants in beagle dogs. *Journal of the Chinese Medical Association : JCMA*. 2018;81(10):920-925.
- Canullo L, Tallarico M, Botticelli D, Alccayhuaman KAA, Martins Neto EC, Xavier SP. Hard and soft tissue changes around implants activated using plasma of argon: A histomorphometric study in dog. *Clin Oral Implants Res.* 2018;29(4):389-395. doi:10.1111/clr.13134
- 57. Coelho PG, Bonfante EA, Pessoa RS, et al. Characterisation of five different implant surfaces and their effect on osseointegration: a study in dogs. *J Periodontol*. 2011;82(5):742-750. doi:10.1902/jop.2010.100520
- Novaes AB, Souza SLS, de Oliveira PT, Souza AMMS. Histomorphometric analysis of the bone-implant contact obtained with 4 different implant surface treatments placed side by side in the dog mandible. *Int J Oral Maxillofac Implants*. 2002;17(3):377-383.
- 59. Qiao S, Cao H, Zhao X, et al. Ag-plasma modification enhances bone apposition around titanium dental implants: an animal study in Labrador dogs. *Int J Nanomedicine*. 2015;10:653-664. doi:10.2147/IJN.S73467
- Mostafa D, Aboushelib M. Bioactive-hybrid-zirconia implant surface for enhancing osseointegration: an in vivo study. Int J Implant Dent. 2018;4(1):20. doi:10.1186/s40729-018-0129-3
- Huang Z, Wang Z, Li C, Yin K, Hao D, Lan J. Application of Plasma Sprayed Zirconia Coating in Dental Implant: Study in Implant. *The Journal of Oral Implantology*. Published online January 5, 2018. doi:10.1563/aaid-joi-D-17-00124
- 62. Metzler P, von Wilmowsky C, Stadlinger B, et al. Nano-crystalline diamond-coated titanium dental implants a histomorphometric study in adult domestic pigs. *J Craniomaxillofac Surg.* 2013;41(6):532-538. doi:10.1016/j.jcms.2012.11.020
- 63. Ballo AM, Bjöörn D, Astrand M, Palmquist A, Lausmaa J, Thomsen P. Bone response to physical-vapour-deposited titanium dioxide coatings on titanium implants. *Clin Oral Implants Res.* 2013;24(9):1009-1017. doi:10.1111/j.1600-0501.2012.02509.x
- 64. Park JY, Gemmell CH, Davies JE. Platelet interactions with titanium: modulation of platelet activity by surface topography. *Biomaterials*. 2001;22(19):2671-2682. doi:10.1016/S0142-9612(01)00009-6
- 65. Ogawa T, Nishimura I. Different bone integration profiles of turned and acid-etched implants associated with modulated expression

of extracellular matrix genes. Int J Oral Maxillofac Implants. 2003;18(2):200-210.

- Degidi M, Petrone G, Iezzi G, Piattelli A. Bone Contact Around Acid-etched Implants: A Histological and Histomorphometrical Evaluation of Two Human-retrieved Implants. *Journal of Oral Implantology*. 2003;29(1):13-18. doi:10.1563/1548-1336(2003)029<0013:BCAAIA>2.3.CO;2
- 67. Weng D, Hoffmeyer M, Hürzeler MB, Richter EJ. Osseotite ® vs. machined surface in poor bone quality: A study in dogs. *Clinical Oral Implants Research*. 2003;14(6):703-708. doi:10.1046/j.0905-7161.2003.00955.x
- 68. Testori T, Szmukler-Moncler S, Francetti L, et al. Immediate loading of Osseotite implants: a case report and histologic analysis after 4 months of occlusal loading. *Int J Periodontics Restorative Dent*. 2001;21(5):451-459.
- 69. Trisi P, Lazzara R, Rao W, Rebaudi A. Bone-implant contact and bone quality: evaluation of expected and actual bone contact on machined and osseotite implant surfaces. *Int J Periodontics Restorative Dent*. 2002;22(6):535-545.
- Cho S. The removal torque of titanium screw inserted in rabbit tibia treated by dual acid etching. *Biomaterials*. 2003;24(20):3611-3617. doi:10.1016/S0142-9612(03)00218-7
- Pontes AEF, de Toledo CT, Garcia VG, Ribeiro FS, Sakakura CE. Torque Analysis of a Triple Acid-Etched Titanium Implant Surface. *ScientificWorldJournal*. 2015;2015:819879. doi:10.1155/2015/819879
- Jinno Y, Jimbo R, Tovar N, Teixeira HS, Witek L, Coelho PG. In Vivo Evaluation of Dual Acid-Etched and Grit-Blasted/ Acid-Etched Implants With Identical Macrogeometry in High-Density Bone. *Implant Dent.* 2017;26(6):815-819. doi:10.1097/ ID.0000000000000672
- 73. de Jesus RNR, Stavropoulos A, Oliveira MTF, Soares PBF, Moura CCG, Zanetta-Barbosa D. Histomorphometric evaluation of a dual acid-etched vs. a chemically modified hydrophilic dual acid-etched implant surface. An experimental study in dogs. *Clin Oral Implants Res.* 2017;28(5):551-557. doi:10.1111/clr.12833
- 74. Bonfante EA, Granato R, Marin C, et al. Early bone healing and biomechanical fixation of dual acid-etched and as-machined implants with healing chambers: an experimental study in dogs. *Int J Oral Maxillofac Implants*. 2011;26(1):75-82.
- 75. Halldin A, Jimbo R, Johansson CB, Gretzer C, Jacobsson M. Improved osseointegration and interlocking capacity with dual acidtreated implants: a rabbit study. *Clin Oral Implants Res.* 2016;27(1):22-30. doi:10.1111/clr.12507
- 76. Fabbro MD, Taschieri S, Canciani E, et al. Osseointegration of Titanium Implants With Different Rough Surfaces: A Histologic and Histomorphometric Study in an Adult Minipig Model. *Implant Dent*. 2017;26(3):357-366. doi:10.1097/ID.000000000000560
- 77. Freitas GP, Lopes HB, Martins-Neto EC, de Oliveira PT, Beloti MM, Rosa AL. Effect of Surface Nanotopography on Bone Response to Titanium Implant. *J Oral Implantol*. 2016;42(3):240-247. doi:10.1563/aaid-joi-D-14-00254
- 78. Yoo D, Marin C, Freitas G, et al. Surface characterisation and in vivo evaluation of dual Acid-etched and grit-blasted/acid-etched implants in sheep. *Implant Dent*. 2015;24(3):256-262. doi:10.1097/ID.0000000000248
- 79. Klokkevold PR, Johnson P, Dadgostari S, Davies JE, Caputo A, Nishimura RD. Early endosseous integration enhanced by dual acid etching of titanium: a torque removal study in the rabbit: Early endosseous integration enhanced by dual acid etching. *Clinical Oral Implants Research*. 2001;12(4):350-357. doi:10.1034/j.1600-0501.2001.012004350.x
- Galli C, Guizzardi S, Passeri G, et al. Comparison of Human Mandibular Osteoblasts Grown on Two Commercially Available Titanium Implant Surfaces. *Journal of Periodontology*. 2005;76(3):364-372. doi:10.1902/jop.2005.76.3.364
- Buser D, Nydegger T, Hirt HP, Cochran DL, Nolte LP. Removal torque values of titanium implants in the maxilla of miniature pigs. *Int J Oral Maxillofac Implants*. 1998;13(5):611-619.
- 82. Abrahamsson I, Berglundh T, Linder E, Lang NP, Lindhe J. Early bone formation adjacent to rough and turned endosseous implant surfaces. An experimental study in the dog. *Clin Oral Implants Res.* 2004;15(4):381-392. doi:10.1111/j.1600-0501.2004.01082.x
- 83. Cochran DL, Schenk RK, Lussi A, Higginbottom FL, Buser D. Bone response to unloaded and loaded titanium implants with a sandblasted and acid-etched surface: A histometric study in the canine mandible. *J Biomed Mater Res.* 1998;40(1):1-11. doi:10.1002/(SICI)1097-4636(199804)40:1<1::AID-JBM1>3.0.CO;2-Q
- 84. Buser D, Schenk RK, Steinemann S, Fiorellini JP, Fox CH, Stich H. Influence of surface characteristics on bone integration of titanium implants. A histomorphometric study in miniature pigs. J Biomed Mater Res. 1991;25(7):889-902. doi:10.1002/

jbm.820250708

- 85. Wilke HJ. The influence of various titanium surfaces on the interface shear strength between implants and bone. *Clinical implant materials Adv Biomater*. 1990;9:309-314.
- 86. Hayakawa T, Kiba H, Yasuda S, Yamamoto H, Nemoto K. A histologic and histomorphometric evaluation of two types of retrieved human titanium implants. *Int J Periodontics Restorative Dent*. 2002;22(2):164-171.
- Sakakura CE, Nociti FH, Mello GPS, de Mello EDA, de Rezende MLR. Histomorphometric Evaluation of a Threaded, Sandblasted, Acid-Etched Implant Retrieved From a Human Lower Jaw: A Case Report. *Implant Dentistry*. 2005;14(3):289-293. doi:10.1097/01.id.0000173641.57293.e4
- 88. Cochran DL, Buser D, Ten Bruggenkate CM, et al. The use of reduced healing times on ITI ® implants with a sandblasted and acid-etched (SLA) surface:: Early results from clinical trials on ITI ® SLA implants. *Clinical Oral Implants Research*. 2002;13(2):144-153. doi:10.1034/j.1600-0501.2002.130204.x
- Guizzardi S, Galli C, Martini D, et al. Different Titanium Surface Treatment Influences Human Mandibular Osteoblast Response. Journal of Periodontology. 2004;75(2):273-282. doi:10.1902/jop.2004.75.2.273
- 90. Dundar S, Yaman F, Bozoglan A, et al. Comparison of Osseointegration of Five Different Surfaced Titanium Implants. *J Craniofac Surg.* 2018;29(7):1991-1995. doi:10.1097/SCS.00000000004572
- Chiang HJ, Hsu HJ, Peng PW, et al. Early bone response to machined, sandblasting acid etching (SLA) and novel surfacefunctionalisation (SLAffinity) titanium implants: characterisation, biomechanical analysis and histological evaluation in pigs. J Biomed Mater Res A. 2016;104(2):397-405. doi:10.1002/jbm.a.35577
- 92. Ernst S, Stübinger S, Schüpbach P, et al. Comparison of two dental implant surface modifications on implants with same macrodesign: an experimental study in the pelvic sheep model. *Clin Oral Implants Res.* 2015;26(8):898-908. doi:10.1111/clr.12411
- Calvo-Guirado JL, Satorres M, Negri B, et al. Biomechanical and histological evaluation of four different titanium implant surface modifications: an experimental study in the rabbit tibia. *Clin Oral Investig.* 2014;18(5):1495-1505. doi:10.1007/s00784-013-1120-2
- 94. Lai HC, Zhuang LF, Zhang ZY, Wieland M, Liu X. Bone apposition around two different sandblasted, large-grit and acidetched implant surfaces at sites with coronal circumferential defects: an experimental study in dogs. *Clin Oral Implants Res.* 2009;20(3):247-253. doi:10.1111/j.1600-0501.2008.01651.x
- Al-Nawas B, Groetz KA, Goetz H, Duschner H, Wagner W. Comparative histomorphometry and resonance frequency analysis of implants with moderately rough surfaces in a loaded animal model. *Clin Oral Implants Res.* 2008;19(1):1-8. doi:10.1111/j.1600-0501.2007.01396.x
- 96. Perrin D, Szmukler-Moncler S, Echikou C, Pointaire P, Bernard JP. Bone response to alteration of surface topography and surface composition of sandblasted and acid etched (SLA) implants. *Clin Oral Implants Res.* 2002;13(5):465-469. doi:10.1034/j.1600-0501.2002.130504.x
- 97. Marinho VC, Celletti R, Bracchetti G, Petrone G, Minkin C, Piattelli A. Sandblasted and acid-etched dental implants: a histologic study in rats. *Int J Oral Maxillofac Implants*. 2003;18(1):75-81.
- 98. Buser D, Nydegger T, Oxland T, et al. Interface shear strength of titanium implants with a sandblasted and acid-etched surface: A biomechanical study in the maxilla of miniature pigs. J Biomed Mater Res. 1999;45(2):75-83. doi:10.1002/(SICI)1097-4636(199905)45:2<75::AID-JBM1>3.0.CO;2-P
- 99. Sul YT, Johansson CB, Petronis S, et al. Characteristics of the surface oxides on turned and electrochemically oxidised pure titanium implants up to dielectric breakdown: *Biomaterials*. 2002;23(2):491-501. doi:10.1016/S0142-9612(01)00131-4
- Xavier SP, Ikuno KE, Tavares MG. Enhanced bone apposition to Brazilian microrough titanium surfaces. *Brazilian dental journal*. 2010;21:18-23.
- 101. Zhu X, Chen J, Scheideler L, Reichl R, Geis-Gerstorfer J. Effects of topography and composition of titanium surface oxides on osteoblast responses. *Biomaterials*. 2004;25(18):4087-4103. doi:10.1016/j.biomaterials.2003.11.011
- 102. Son W woo, Zhu X, Shin H in, Ong JL, Kim K han. In vivo histological response to anodised and anodised/hydrothermally treated titanium implants. *J Biomed Mater Res.* 2003;66B(2):520-525. doi:10.1002/jbm.b.10042

- 103. Choi JW, Heo SJ, Koak JY, et al. Biological responses of anodised titanium implants under different current voltages. J Oral Rehabil. 2006;33(12):889-897. doi:10.1111/j.1365-2842.2006.01669.x
- 104. Yamagami A, Yoshihara Y, Suwa F. Mechanical and histologic examination of titanium alloy material treated by sandblasting and anodic oxidisation. Int J Oral Maxillofac Implants. 2005;20(1):48-53.
- 105. Kang BS, Sul YT, Johansson CB, Oh SJ, Lee HJ, Albrektsson T. The effect of calcium ion concentration on the bone response to oxidised titanium implants. *Clin Oral Implants Res.* 2012;23(6):690-697. doi:10.1111/j.1600-0501.2011.02177.x
- 106. Knobloch L, Larsen PA, Rashid B, Carr AB. Six-month performance of implants with oxidised and machined surfaces restored at 2, 4, and 6 weeks postimplantation in adult beagle dogs. *Int J Oral Maxillofac Implants*. 2004;19(3):350-356.
- 107. Lee HJ, Yang IH, Kim SK, Yeo IS, Kwon TK. In vivo comparison between the effects of chemically modified hydrophilic and anodically oxidised titanium surfaces on initial bone healing. *J Periodontal Implant Sci.* 2015;45(3):94-100. doi:10.5051/ jpis.2015.45.3.94
- 108. Park IP, Kang TJ, Heo SJ, et al. Investigation of anodised titanium implants coated with triterpenoids extracted from black cohosh: an animal study. *J Adv Prosthodont*. 2014;6(1):14-21. doi:10.4047/jap.2014.6.1.14
- 109. Sul YT, Johansson CB, Kang Y, Jeon DG, Albrektsson T. Bone reactions to oxidised titanium implants with electrochemical anion sulphuric acid and phosphoric acid incorporation. *Clin Implant Dent Relat Res.* 2002;4(2):78-87. doi:10.1111/j.1708-8208.2002.tb00156.x
- 110. Li X, Xu H, Zhao B, Jiang S. Accelerated and enhanced osteointegration of MAO-treated implants: histological and histomorphometric evaluation in a rabbit model. *Int J Oral Sci.* 2018;10(2):11. doi:10.1038/s41368-018-0008-z
- 111. Ma W, Wei JH, Li YZ, et al. Histological evaluation and surface componential analysis of modified micro-arc oxidation-treated titanium implants. *J Biomed Mater Res B Appl Biomater*. 2008;86(1):162-169. doi:10.1002/jbm.b.31002
- 112. Sul YT, Johansson CB, Albrektsson T. Oxidized titanium screws coated with calcium ions and their performance in rabbit bone. Int J Oral Maxillofac Implants. 2002;17(5):625-634.
- 113. Ivanoff CJ, Widmark G, Johansson C, Wennerberg A. Histologic evaluation of bone response to oxidised and turned titanium micro-implants in human jawbone. *Int J Oral Maxillofac Implants*. 2003;18(3):341-348.
- 114. Xie J, Baumann MJ, McCabe LR. Osteoblasts respond to hydroxyapatite surfaces with immediate changes in gene expression. J Biomed Mater Res. 2004;71A(1):108-117. doi:10.1002/jbm.a.30140
- 115. Uehara T, Takaoka K, Ito K. Histological evidence of osseointegration in human retrieved fractured hydroxyapatite-coated screwtype implants: a case report. *Clin Oral Implants Res.* 2004;15(5):540-545. doi:10.1111/j.1600-0501.2004.01031.x
- 116. Morris HF, Ochi S. Hydroxyapatite-coated implants: A case for their use. *Journal of Oral and Maxillofacial Surgery*. 1998;56(11):1303-1311. doi:10.1016/S0278-2391(98)90615-2
- 117. Lee JJ, Rouhfar L, Beirne OR. Survival of hydroxyapatite-coated implants: A meta-analytic review. *Journal of Oral and Maxillofacial Surgery*. 2000;58(12):1372-1379. doi:10.1053/joms.2000.18269
- 118. Trisi P, Keith DJ, Rocco S. Human histologic and histomorphometric analyses of hydroxyapatite-coated implants after 10 years of function: a case report. *Int J Oral Maxillofac Implants*. 2005;20(1):124-130.
- 119. Piattelli A, Cosci F, Scarano A, Trisi P. Localised chronic suppurative bone infection as a sequel of peri-implantitis in a hydroxyapatite-coated dental implant. *Biomaterials*. 1995;16(12):917-920. doi:10.1016/0142-9612(95)93116-U
- Sun L, Berndt CC, Gross KA, Kucuk A. Material fundamentals and clinical performance of plasma-sprayed hydroxyapatite coatings: A review. J Biomed Mater Res. 2001;58(5):570-592. doi:10.1002/jbm.1056
- Ong JL, Carnes DL, Bessho K. Evaluation of titanium plasma-sprayed and plasma-sprayed hydroxyapatite implants in vivo. *Biomaterials*. 2004;25(19):4601-4606. doi:10.1016/j.biomaterials.2003.11.053
- 122. Piattelli M, Scarano A, Paolantonio M, Iezzi G, Petrone G, Piattelli A. Bone Response to Machined and Resorbable Blast Material Titanium Implants: An Experimental Study in Rabbits. *Journal of Oral Implantology*. 2002;28(1):2-8. doi:10.1563/1548-1336(2002)028<0002:BRTMAR>2.3.CO;2
- 123. Jung YC, Han CH, Lee IS, Kim HE. Effects of ion beam-assisted deposition of hydroxyapatite on the osseointegration of

endosseous implants in rabbit tibiae. Int J Oral Maxillofac Implants. 2001;16(6):809-818.

- 124. Lee IS, Kim DH, Kim HE, Jung YC, Han CH. Biological performance of calcium phosphate films formed on commercially pure Ti by electron-beam evaporation. *Biomaterials*. 2002;23(2):609-615. doi:10.1016/S0142-9612(01)00147-8
- 125. Svanborg LM, Hoffman M, Andersson M, Currie F, Kjellin P, Wennerberg A. The effect of hydroxyapatite nanocrystals on early bone formation surrounding dental implants. *Int J Oral Maxillofac Surg.* 2011;40(3):308-315. doi:10.1016/j.ijom.2010.10.010
- 126. Scarano A, Piattelli A, Quaranta A, Lorusso F. Bone Response to Two Dental Implants with Different Sandblasted/Acid-Etched Implant Surfaces: A Histological and Histomorphometrical Study in Rabbits. *BioMed Research International*. 2017;2017:8724951. doi:10.1155/2017/8724951
- 127. Chan YH, Lew WZ, Lu E, et al. An evaluation of the biocompatibility and osseointegration of novel glass fiber reinforced composite implants: In vitro and in vivo studies. *Dent Mater*. 2018;34(3):470-485. doi:10.1016/j.dental.2017.12.001
- 128. Trisi P, Berardini M, Falco A, Sandrini E, Vulpiani MP. A New Highly Hydrophilic Electrochemical Implant Titanium Surface: A Histological and Biomechanical In Vivo Study. *Implant Dentistry*. 2017;26(3):429-437. doi:10.1097/ID.00000000000605
- 129. Cardoso MV, de Rycker J, Chaudhari A, et al. Titanium implant functionalisation with phosphate-containing polymers may favour in vivo osseointegration. *J Clin Periodontol*. 2017;44(9):950-960. doi:10.1111/jcpe.12736
- 130. Liu Y, Zhou Y, Jiang T, Liang YD, Zhang Z, Wang YN. Evaluation of the osseointegration of dental implants coated with calcium carbonate: an animal study. *Int J Oral Sci.* 2017;9(3):133-138. doi:10.1038/ijos.2017.13
- 131. Kalemaj Z, Scarano A, Valbonetti L, Rapone B, Grassi FR. Bone Response to Four Dental Implants with Different Surface Topographies: A Histologic and Histometric Study in Minipigs. *The International Journal of Periodontics & Restorative Dentistry*. 2016;36(5):745-754. doi:10.11607/prd.2719
- 132. Song WW, Heo JH, Lee JH, Park YM, Kim YD. Osseointegration of magnesium-incorporated sandblasted acid-etched implant in the dog mandible: Resonance frequency measurements and histomorphometric analysis. *Tissue Eng Regen Med.* 2016;13(2):191-199. doi:10.1007/s13770-016-9126-x
- 133. Mistry S, Roy S, Jyoti Maitra N, et al. Safety and efficacy of additive and subtractive surface modification of Ti6Al4V endosseous implant in goat bone. *J Mech Behav Biomed Mater*. 2016;57:69-87. doi:10.1016/j.jmbbm.2015.11.019
- 134. Armencea G, Berce C, Rotaru H, et al. Micro-CT and histological analysis of Ti6Al7Nb custom made implants with hydroxyapatite and SiO2-TiO2 coatings in a rabbit model. *Clujul Med.* 2015;88(3):408-414. doi:10.15386/cjmed-479
- 135. Im JH, Kim SG, Oh JS, Lim SC. A Comparative Study of Stability After the Installation of 2 Different Surface Types of Implants in the Maxillae of Dogs. *Implant Dent.* 2015;24(5):586-591. doi:10.1097/ID.0000000000292
- 136. Melin Svanborg L, Meirelles L, Franke Stenport V, et al. Evaluation of bone healing on sandblasted and acid etched implants coated with nanocrystalline hydroxyapatite: an in vivo study in rabbit femur. *Int J Dent.* 2014;2014:197581. doi:10.1155/2014/197581
- 137. Bryington MS, Hayashi M, Kozai Y, et al. The influence of nano hydroxyapatite coating on osseointegration after extended healing periods. *Dent Mater*. 2013;29(5):514-520. doi:10.1016/j.dental.2013.02.004
- 138. Eom TG, Jeon GR, Jeong CM, et al. Experimental study of bone response to hydroxyapatite coating implants: bone-implant contact and removal torque test. Oral Surg Oral Med Oral Pathol Oral Radiol. 2012;114(4):411-418. doi:10.1016/j.0000.2011.10.036
- 139. Choi JY, Jung UW, Kim CS, Jung SM, Lee IS, Choi SH. Influence of nanocoated calcium phosphate on two different types of implant surfaces in different bone environment: an animal study. *Clin Oral Implants Res.* 2013;24(9):1018-1022. doi:10.1111/ j.1600-0501.2012.02492.x
- 140. Poulos NM, Rodriguez NA, Lee J, et al. Evaluation of a novel calcium phosphate-coated titanium porous oxide implant surface: a study in rabbits. *Int J Oral Maxillofac Implants*. 2011;26(4):731-738.
- 141. Mano T, Ishikawa K, Harada K, Umeda H, Ueyama Y. Comparison of apatite-coated titanium prepared by blast coating and flame spray methods--evaluation using simulated body fluid and initial histological study. *Dent Mater J.* 2011;30(4):431-437. doi:10.4012/dmj.2010-162
- 142. Jimbo R, Sotres J, Johansson C, Breding K, Currie F, Wennerberg A. The biological response to three different nanostructures applied on smooth implant surfaces. *Clin Oral Implants Res.* 2012;23(6):706-712. doi:10.1111/j.1600-0501.2011.02182.x

- 143. Coelho PG, Granato R, Marin C, et al. Effect of Si addition on Ca- and P-impregnated implant surfaces with nanometer-scale roughness: an experimental study in dogs. *Clin Oral Implants Res.* 2012;23(3):373-378. doi:10.1111/j.1600-0501.2010.02150.x
- 144. Suzuki M, Guimaraes MVM, Marin C, et al. Histomorphologic and bone-to-implant contact evaluation of dual acid-etched and bioceramic grit-blasted implant surfaces: an experimental study in dogs. J Oral Maxillofac Surg. 2010;68(8):1877-1883. doi:10.1016/j.joms.2009.050
- 145. Junker R, Manders PJD, Wolke J, Borisov Y, Jansen JA. Bone-supportive behavior of microplasma-sprayed CaP-coated implants: mechanical and histological outcome in the goat. *Clin Oral Implants Res.* 2010;21(2):189-200. doi:10.1111/j.1600-0501.2009.01819.x
- 146. Lin A, Wang CJ, Kelly J, Gubbi P, Nishimura I. The role of titanium implant surface modification with hydroxyapatite nanoparticles in progressive early bone-implant fixation in vivo. *Int J Oral Maxillofac Implants*. 2009;24(5):808-816.
- Barros RRM, Novaes AB, Papalexiou V, et al. Effect of biofunctionalised implant surface on osseointegration: a histomorphometric study in dogs. *Braz Dent J.* 2009;20(2):91-98. doi:10.1590/s0103-64402009000200001
- Suzuki M, Guimaraes MVM, Marin C, Granato R, Gil JN, Coelho PG. Histomorphometric evaluation of alumina-blasted/acidetched and thin ion beam-deposited bioceramic surfaces: an experimental study in dogs. *J Oral Maxillofac Surg.* 2009;67(3):602-607. doi:10.1016/j.joms.2008.08.021
- 149. Yang G li, He F ming, Hu J an, Wang X xiang, Zhao S fang. Effects of biomimetically and electrochemically deposited nanohydroxyapatite coatings on osseointegration of porous titanium implants. Oral Surg Oral Med Oral Pathol Oral Radiol Endod. 2009;107(6):782-789. doi:10.1016/j.tripleo.2008.12.023
- 150. Granato R, Marin C, Suzuki M, Gil JN, Janal MN, Coelho PG. Biomechanical and histomorphometric evaluation of a thin ion beam bioceramic deposition on plateau root form implants: an experimental study in dogs. *J Biomed Mater Res B Appl Biomater*. 2009;90(1):396-403. doi:10.1002/jbm.b.31298
- 151. Le Guehennec L, Goyenvalle E, Lopez-Heredia MA, Weiss P, Amouriq Y, Layrolle P. Histomorphometric analysis of the osseointegration of four different implant surfaces in the femoral epiphyses of rabbits. *Clin Oral Implants Res.* 2008;19(11):1103-1110. doi:10.1111/j.1600-0501.2008.01547.x
- Marin C, Granato R, Suzuki M, Gil JN, Piattelli A, Coelho PG. Removal torque and histomorphometric evaluation of bioceramic grit-blasted/acid-etched and dual acid-etched implant surfaces: an experimental study in dogs. *J Periodontol*. 2008;79(10):1942-1949. doi:10.1902/jop.2008.080106
- 153. Coelho PG, Cardaropoli G, Suzuki M, Lemons JE. Histomorphometric evaluation of a nanothickness bioceramic deposition on endosseous implants: a study in dogs. *Clin Implant Dent Relat Res*. 2009;11(4):292-302. doi:10.1111/j.1708-8208.2008.00122.x
- 154. Alzubaydi TL, Alameer SS, Ismaeel T, Alhijazi AY, Geetha M. In vivo studies of the ceramic coated titanium alloy for enhanced osseointegration in dental applications. *J Mater Sci Mater Med.* 2009;20 Suppl 1:S35-42. doi:10.1007/s10856-008-3479-1
- 155. Franco R de L, Chiesa R, de Oliveira PT, Beloti MM, Rosa AL. Bone response to a Ca- and P-enriched titanium surface obtained by anodisation. *Braz Dent J.* 2008;19(1):15-20. doi:10.1590/s0103-64402008000100003
- 156. Yeo IS, Han JS, Yang JH. Biomechanical and histomorphometric study of dental implants with different surface characteristics. Journal of Biomedical Materials Research Part B: Applied Biomaterials: An Official Journal of The Society for Biomaterials, The Japanese Society for Biomaterials, and The Australian Society for Biomaterials and the Korean Society for Biomaterials. 2008;87(2):303-311.
- 157. Zagury R, Harari ND, Conz MB, Soares G de A, Vidigal GM. Histomorphometric analyses of bone interface with titaniumaluminum-vanadium and hydroxyapatite-coated implants by biomimetic process. *Implant Dent*. 2007;16(3):290-296. doi:10.1097/ ID.0b013e3180e9d9ed
- 158. Siebers MC, Wolke JGC, Walboomers XF, Leeuwenburgh SCG, Jansen JA. In vivo evaluation of the trabecular bone behavior to porous electrostatic spray deposition-derived calcium phosphate coatings. *Clin Oral Implants Res.* 2007;18(3):354-361. doi:10.1111/j.1600-0501.2006.01314.x
- 159. Hayakawa T, Takahashi K, Yoshinari M, et al. Trabecular bone response to titanium implants with a thin carbonate-containing

apatite coating applied using the molecular precursor method. Int J Oral Maxillofac Implants. 2006;21(6):851-858.

- 160. Manders PJD, Wolke JGC, Jansen JA. Bone response adjacent to calcium phosphate electrostatic spray deposition coated implants: an experimental study in goats. *Clin Oral Implants Res.* 2006;17(5):548-553. doi:10.1111/j.1600-0501.2006.01263.x
- 161. Xiropaidis AV, Qahash M, Lim WH, et al. Bone-implant contact at calcium phosphate-coated and porous titanium oxide (TiUnite)modified oral implants. *Clin Oral Implants Res.* 2005;16(5):532-539. doi:10.1111/j.1600-0501.2005.01126.x
- 162. Schopper C, Moser D, Goriwoda W, et al. The effect of three different calcium phosphate implant coatings on bone deposition and coating resorption: a long-term histological study in sheep. *Clin Oral Implants Res.* 2005;16(3):357-368. doi:10.1111/j.1600-0501.2004.01080.x
- 163. Hayakawa T, Yoshinari M, Kiba H, Yamamoto H, Nemoto K, Jansen JA. Trabecular bone response to surface roughened and calcium phosphate (Ca-P) coated titanium implants. *Biomaterials*. 2002;23(4):1025-1031. doi:10.1016/S0142-9612(01)00214-9
- Strnad Z, Strnad J, Povýsil C, Urban K. Effect of plasma-sprayed hydroxyapatite coating on the osteoconductivity of commercially pure titanium implants. *Int J Oral Maxillofac Implants*. 2000;15(4):483-490.
- 165. Hulshoff JE, van Dijk K, van der Waerden JP, Wolke JG, Kalk W, Jansen JA. Evaluation of plasma-spray and magnetron-sputter Ca-P-coated implants: an in vivo experiment using rabbits. *J Biomed Mater Res.* 1996;31(3):329-337. doi:10.1002/(SICI)1097-4636(199607)31:3<329::AID-JBM6>3.0.CO;2-O
- 166. Yoshinari M, Klinge B, Dérand T. The biocompatibility (cell culture and histologic study) of hydroxy-apatite-coated implants created by ion beam dynamic mixing. *Clin Oral Implants Res.* 1996;7(2):96-100. doi:10.1034/j.1600-0501.1996.070202.x
- 167. Caulier H, van der Waerden JP, Paquay YC, et al. Effect of calcium phosphate (Ca-P) coatings on trabecular bone response: a histological study. J Biomed Mater Res. 1995;29(9):1061-1069. doi:10.1002/jbm.820290906
- 168. OG B, C C, MA A, S AH, ML E, MC A. Osseointegration of TI6Al4V dental implants modified by thermal oxidation in osteoporotic rabbits. *International journal of implant dentistry*. 2016;2(1):18.
- 169. Ozeki K, Okuyama Y, Fukui Y, Aoki H. Bone response to titanium implants coated with thin sputtered HA film subject to hydrothermal treatment and implanted in the canine mandible. *Biomed Mater Eng.* 2006;16(4):243-251.
- García-Alonso MC, Saldaña L, Vallés G, et al. In vitro corrosion behaviour and osteoblast response of thermally oxidised Ti6Al4V alloy. *Biomaterials*. 2003;24(1):19-26. doi:10.1016/s0142-9612(02)00237-5
- 171. Scarano A, Crocetta E, Quaranta A, Lorusso F. Influence of the thermal treatment to address a better osseointegration of Ti6Al4V Dental Implants: Histological and histomorphometrical study in a rabbit model. *BioMed Research International*. 2018;2018. doi:10.1155/2018/2349698
- 172. Herrero-Climent M, Romero Ruiza MM, Calvo PL, Santos JVR, Perez RA, Gil Mur FJ. Effectiveness of a new dental implant bioactive surface: histological and histomorphometric comparative study in minipigs. *Clin Oral Investig.* 2018;22(3):1423-1432. doi:10.1007/s00784-017-2223-y
- 173. Qamheya AHA, Arısan V, Mutlu Z, et al. Thermal oxidation and hydrofluoric acid treatment on the sandblasted implant surface: A histologic histomorphometric and biomechanical study. *Clin Oral Implants Res.* 2018;29(7):741-755. doi:10.1111/clr.13285
- 174. Kim NS, Vang MS, Yang HS, Park SW, Park HO, Lim HP. Comparion of stability in titanium implants with different surface topographies in dogs. *J Adv Prosthodont*. 2009;1(1):47-55. doi:10.4047/jap.2009.1.1.47
- 175. Piconi C, Maccauro G. Zirconia as a ceramic biomaterial. Biomaterials. 1999;20(1):1-25.
- 176. Han JM, Hong G, Lin H, et al. Biomechanical and histological evaluation of the osseointegration capacity of two types of zirconia implant. *Int J Nanomedicine*. 2016;11:6507-6516. doi:10.2147/IJN.S119519
- 177. Martins R, Cestari TM, Arantes RVN, et al. Osseointegration of zirconia and titanium implants in a rabbit tibiae model evaluated by microtomography, histomorphometry and fluorochrome labeling analyses. J Periodontal Res. 2018;53(2):210-221. doi:10.1111/ jre.12508
- 178. Mihatovic I, Golubovic V, Becker J, Schwarz F. Bone tissue response to experimental zirconia implants. *Clinical oral investigations*. 2017;21(2):523-532.
- 179. Jimbo R, Naito Y, Galli S, Berner S, Dard M, Wennerberg A. Biomechanical and Histomorphometrical Evaluation of TiZr Alloy

Implants: An in vivo Study in the Rabbit. Clin Implant Dent Relat Res. 2015;17 Suppl 2:e670-678. doi:10.1111/cid.12305

- Gredes T, Kubasiewicz-Ross P, Gedrange T, Dominiak M, Kunert-Keil C. Comparison of surface modified zirconia implants with commercially available zirconium and titanium implants: a histological study in pigs. *Implant Dent.* 2014;23(4):502-507. doi:10.1097/ID.00000000000110
- 181. Saulacic N, Erdösi R, Bosshardt DD, Gruber R, Buser D. Acid and alkaline etching of sandblasted zirconia implants: a histomorphometric study in miniature pigs. *Clin Implant Dent Relat Res.* 2014;16(3):313-322. doi:10.1111/cid.12070
- 182. Hoffmann O, Angelov N, Zafiropoulos GG, Andreana S. Osseointegration of zirconia implants with different surface characteristics: an evaluation in rabbits. *Int J Oral Maxillofac Implants*. 2012;27(2):352-358.
- 183. Lee J, Sieweke JH, Rodriguez NA, et al. Evaluation of nano-technology-modified zirconia oral implants: a study in rabbits. J Clin Periodontol. 2009;36(7):610-617. doi:10.1111/j.1600-051X.2009.01423.x
- 184. Kohal RJ, Wolkewitz M, Hinze M, Han JS, Bächle M, Butz F. Biomechanical and histological behavior of zirconia implants: an experiment in the rat. *Clin Oral Implants Res.* 2009;20(4):333-339. doi:10.1111/j.1600-0501.2008.01656.x
- Depprich R, Zipprich H, Ommerborn M, et al. Osseointegration of zirconia implants compared with titanium: an in vivo study. *Head Face Med.* 2008;4:30. doi:10.1186/1746-160X-4-30
- 186. Langhoff JD, Voelter K, Scharnweber D, et al. Comparison of chemically and pharmaceutically modified titanium and zirconia implant surfaces in dentistry: a study in sheep. *Int J Oral Maxillofac Surg.* 2008;37(12):1125-1132. doi:10.1016/j.ijom.2008.09.008
- 187. Gahlert M, Gudehus T, Eichhorn S, Steinhauser E, Kniha H, Erhardt W. Biomechanical and histomorphometric comparison between zirconia implants with varying surface textures and a titanium implant in the maxilla of miniature pigs. *Clin Oral Implants Res.* 2007;18(5):662-668. doi:10.1111/j.1600-0501.2007.01401.x
- 188. Sennerby L, Dasmah A, Larsson B, Iverhed M. Bone tissue responses to surface-modified zirconia implants: A histomorphometric and removal torque study in the rabbit. *Clin Implant Dent Relat Res.* 2005;7 Suppl 1:S13-20. doi:10.1111/j.1708-8208.2005.tb00070.x
- 189. Dubruille JH, Viguier E, Le Naour G, Dubruille MT, Auriol M, Le Charpentier Y. Evaluation of combinations of titanium, zirconia, and alumina implants with 2 bone fillers in the dog. *Int J Oral Maxillofac Implants*. 1999;14(2):271-277.
- 190. Kohal RJ, Weng D, Bächle M, Strub JR. Loaded Custom-Made Zirconia and Titanium Implants Show Similar Osseointegration: An Animal Experiment. *Journal of Periodontology*. 2004;75(9):1262-1268. doi:10.1902/jop.2004.75.9.1262
- Albrektsson T, Hansson H, Ivarsson B. Interface analysis of titanium and zirconium bone implants. *Biomaterials*. 1985;6(2):97-101. doi:10.1016/0142-9612(85)90070-5
- 192. Thomsen P, Larsson C, Ericson LE, Sennerby L, Lausmaa J, Kasemo B. Structure of the interface between rabbit cortical bone and implants of gold, zirconium and titanium. *Journal of Materials Science: Materials in Medicine*. 1997;8(11):653-665.
- 193. Ichikawa Y, Akagawa Y, Nikai H, Tsuru H. Tissue compatibility and stability of a new zirconia ceramic in vivo. J Prosthet Dent. 1992;68(2):322-326. doi:10.1016/0022-3913(92)90338-b
- 194. Akagawa Y, Ichikawa Y, Nikai H, Tsuru H. Interface histology of unloaded and early loaded partially stabilised zirconia endosseous implant in initial bone healing. *J Prosthet Dent.* 1993;69(6):599-604. doi:10.1016/0022-3913(93)90289-z
- 195. Covacci V, Bruzzese N, Maccauro G, et al. In vitro evaluation of the mutagenic and carcinogenic power of high purity zirconia ceramic. *Biomaterials*. 1999;20(4):371-376. doi:10.1016/S0142-9612(98)00182-3
- 196. Schultze-Mosgau S, Schliephake H, Radespiel-Tröger M, Neukam FW. Osseointegration of endodontic endosseous conesZirconium oxide vs titanium. Oral Surgery, Oral Medicine, Oral Pathology, Oral Radiology, and Endodontology. 2000;89(1):91-98. doi:10.1016/S1079-2104(00)80022-0
- 197. Josset Y, Oum'Hamed Z, Zarrinpour A, Lorenzato M, Adnet JJ, Laurent-Maquin D. In vitro reactions of human osteoblasts in culture with zirconia and alumina ceramics. *J Biomed Mater Res.* 1999;47(4):481-493. doi:10.1002/(SICI)1097-4636(19991215)47:4<481::AID-JBM4>3.0.CO;2-Y
- 198. Scarano A, Di Carlo F, Quaranta M, Piattelli A. Bone response to zirconia ceramic implants: an experimental study in rabbits. *The Journal of Oral Implantology*. 2003;29(1):8-12. doi:10.1563/1548-1336(2003)029<0008:BRTZCI>2.3.CO;2
- 199. Akagawa Y, Hosokawa R, Sato Y, Kamayama K. Comparison between freestanding and tooth-connected partially stabilised

zirconia implants after two years' function in monkeys: a clinical and histologic study. *J Prosthet Dent*. 1998;80(5):551-558. doi:10.1016/s0022-3913(98)70031-9

- 200. Okamatsu K, Kido H, Sato A, Watazu A, Matsuura M. Ultrastructure of the interface between titanium and surrounding tissue in rat tibiae--a comparison study on titanium-coated and -uncoated plastic implants. *Clin Implant Dent Relat Res.* 2007;9(2):100-111. doi:10.1111/j.1708-8208.2007.00032.x
- 201. Mehl C, Kern M, Neumann F, Bähr T, Wiltfang J, Gassling V. Effect of ultraviolet photofunctionalization of dental titanium implants on osseointegration. *J Zhejiang Univ Sci B*. 2018;19(7):525-534. doi:10.1631/jzus.B1600505
- 202. Yamauchi R, Itabashi T, Wada K, Tanaka T, Kumagai G, Ishibashi Y. Photofunctionalised Ti6Al4V implants enhance early phase osseointegration. *Bone Joint Res.* 2017;6(5):331-336. doi:10.1302/2046-3758.65.BJR-2016-0221.R1
- 203. Shen J, Liu J, Chen X, Wang X, He F, Wang H. The In Vivo Bone Response of Ultraviolet-Irradiated Titanium Implants Modified with Spontaneously Formed Nanostructures: An Experimental Study in Rabbits. *Int J Oral Maxillofac Implants*. 2016;31(4):776-784. doi:10.11607/jomi.4309
- 204. Hirakawa Y, Jimbo R, Shibata Y, Watanabe I, Wennerberg A, Sawase T. Accelerated bone formation on photo-induced hydrophilic titanium implants: an experimental study in the dog mandible. *Clin Oral Implants Res.* 2013;24 Suppl A100:139-144. doi:10.1111/j.1600-0501.2011.02401.x
- 205. Fiorellini JP, Glindmann S, Salcedo J, Weber HP, Park CJ, Sarmiento HL. The Effect of Osteopontin and an Osteopontin-Derived Synthetic Peptide Coating on Osseointegration of Implants in a Canine Model. *Int J Periodontics Restorative Dent*. 2016;36(6):e88-e94. doi:10.11607/prd.2830
- 206. Johansson CB, Han CH, Wennerberg A, Albrektsson T. A quantitative comparison of machined commercially pure titanium and titanium-aluminum-vanadium implants in rabbit bone. *International Journal of Oral & Maxillofacial Implants*. 1998;13(3).
- 207. Carr AB, Larsen PE, Gerard DA. Histomorphometric comparison of implant anchorage for two types of dental implants after 3 and 6 months' healing in baboon jaws. *The Journal of Prosthetic Dentistry*. 2001;85(3):276-280. doi:10.1067/mpr.2001.114821
- Lausmaa J, Mattsson L, Rolander U, Kasemo B. Chemical Composition and Morphology of Titanium Surface Oxides. MRS Proc. 1985;55:351. doi:10.1557/PROC-55-351
- 209. Arys A, Philippart C, Dourov N, He Y, Le QT, Pireaux JJ. Analysis of titanium dental implants after failure of osseointegration: Combined histological, electron microscopy, and X-ray photoelectron spectroscopy approach. J Biomed Mater Res. 1998;43(3):300-312. doi:10.1002/(SICI)1097-4636(199823)43:3<300::AID-JBM11>3.0.CO;2-J
- Zitter H, Plenk H. The electrochemical behavior of metallic implant materials as an indicator of their biocompatibility. J Biomed Mater Res. 1987;21(7):881-896. doi:10.1002/jbm.820210705
- 211. Ducheyne P. Titanium and calcium phosphate ceramic dental implants, surfaces, coatings and interfaces. *The Journal of Oral Implantology*. 1988;14(3):325-340.
- 212. Larsson C, Thomsen P, Aronsson BO, et al. Bone response to surface-modified titanium implants: studies on the early tissue response to machined and electropolished implants with different oxide thicknesses. *Biomaterials*. 1996;17(6):605-616. doi:10.1016/0142-9612(96)88711-4
- 213. Olefjord I, Hansson S. Surface analysis of four dental implant systems. *International Journal of Oral & Maxillofacial Implants*. 1993;8(1).
- 214. Lim YJ, Oshida Y, Andres CJ, Barco MT. Surface characterisations of variously treated titanium materials. *International Journal* of Oral & Maxillofacial Implants. 2001;16(3).
- Sul YT, Johansson CB, Röser K, Albrektsson T. Qualitative and quantitative observations of bone tissue reactions to anodised implants. *Biomaterials*. 2002;23(8):1809-1817. doi:10.1016/S0142-9612(01)00307-6
- 216. Lee TM, Yang CY, Chang E, Tsai RS. Comparison of plasma-sprayed hydroxyapatite coatings and zirconia-reinforced hydroxyapatite composite coatings: In vivo study. *J Biomed Mater Res.* 2004;71A(4):652-660. doi:10.1002/jbm.a.30190
- 217. Tambasco de Oliveira P, Nanci A. Nanotexturing of titanium-based surfaces upregulates expression of bone sialoprotein and osteopontin by cultured osteogenic cells. *Biomaterials*. 2004;25(3):403-413. doi:10.1016/S0142-9612(03)00539-8

- 218. Webster TJ, Ejiofor JU. Increased osteoblast adhesion on nanophase metals: Ti, Ti6Al4V, and CoCrMo. *Biomaterials*. 2004;25(19):4731-4739. doi:10.1016/j.biomaterials.2003.12.002
- 219. Di Iorio D, Traini T, Degidi M, Caputi S, Neugebauer J, Piattelli A. Quantitative evaluation of the fibrin clot extension on different implant surfaces: Anin vitro study. *J Biomed Mater Res.* 2005;74B(1):636-642. doi:10.1002/jbm.b.30251
- 220. Papalexiou V, Novaes AB, Grisi MFM, Souza SSLS, Taba M, Kajiwara JK. Influence of implant microstructure on the dynamics of bone healing around immediate implants placed into periodontally infected sites. A confocal laser scanning microscopic study. *Clin Oral Implants Res.* 2004;15(1):44-53. doi:10.1111/j.1600-0501.2004.00995.x
- 221. Iezzi G, Degidi M, Scarano A, Perrotti V, Piattelli A. Bone Response to Submerged, Unloaded Implants Inserted in Poor Bone Sites: A Histological and Histomorphometrical Study of 8 Titanium Implants Retrieved From Man. *Journal of Oral Implantology*. 2005;31(5):225-233. doi:10.1563/1548-1336(2005)31[225:BRTSUI]2.0.CO;2
- 222. Wisbey A, Gregson PJ, Tuke M. Application of PVD TiN coating to Co-Cr-Mo based surgical implants. *Biomaterials*. 1987;8(6):477-480. doi:10.1016/0142-9612(87)90085-8
- 223. Scarano A, Piattelli M, Vrespa G, Petrone G, Iezzi G, Piattelli A. Bone Healing around Titanium and Titanium Nitride-Coated Dental Implants with Three Surfaces: An Experimental Study in Rats. *Clin Implant Dent Rel Res.* 2003;5(2):103-111. doi:10.1111/j.1708-8208.2003.tb00191.x
- 224. Scarano A, Piattelli M, Vrespa G, Caputi S, Piattelli A. Bacterial adhesion on titanium nitride-coated and uncoated implants: an in vivo human study. *The Journal of Oral Implantology*. 2003;29(2):80-85. doi:10.1563/1548-1336(2003)029<0080:BAOTNA> 2.3.CO;2
- 225. Groessner-Schreiber B, Neubert A, Müller WD, Hopp M, Griepentrog M, Lange KP. Fibroblast growth on surface-modified dental implants: An *in vitro* study: Fibroblasts on Dental Implant Surfaces. *J Biomed Mater Res.* 2003;64A(4):591-599. doi:10.1002/ jbm.a.10417
- 226. Größner-Schreiber B, Griepentrog M, Haustein I, et al. Plaque formation on surface modified dental implants: An *in vitro* study. *Clinical Oral Implants Research*. 2001;12(6):543-551. doi:10.1034/j.1600-0501.2001.120601.x
- 227. Groessner-Schreiber B, Hannig M, Duck A, Griepentrog M, Wenderoth DF. Do different implant surfaces exposed in the oral cavity of humans show different biofilm compositions and activities? *Eur J Oral Sci.* 2004;112(6):516-522. doi:10.1111/j.1600-0722.2004.00171.x
- 228. Park CY, Kim SG, Kim MD, Eom TG, Yoon JH, Ahn SG. Surface Properties of Endosseous Dental Implants After NdYAG and CO2 Laser Treatment at Various Energies. *Journal of Oral and Maxillofacial Surgery*. 2005;63(10):1522-1527. doi:10.1016/j. joms.2005.06.015
- 229. Hallgren C. An in vivo study of bone response to implants topographically modified by laser micromachining. *Biomaterials*. 2003;24(5):701-710. doi:10.1016/S0142-9612(02)00266-1
- 230. Citeau A, Guicheux J, Vinatier C, et al. In vitro biological effects of titanium rough surface obtained by calcium phosphate grid blasting. *Biomaterials*. 2005;26(2):157-165. doi:10.1016/j.biomaterials.2004.02.033
- 231. Schopper C, Ziya-Ghazvini F, Goriwoda W, et al. HA/TCP compounding of a porous CaP biomaterial improves bone formation and scaffold degradation—A long-term histological study. *J Biomed Mater Res.* 2005;74B(1):458-467. doi:10.1002/jbm.b.30199
- Wen HB, de Wijn JR, Cui FZ, de Groot K. Preparation of calcium phosphate coatings on titanium implant materials by simple chemistry. J Biomed Mater Res. 1998;41(2):227-236. doi:10.1002/(SICI)1097-4636(199808)41:2<227::AID-JBM7>3.0.CO;2-K
- 233. Yang Y, Kim KH, Ong JL. A review on calcium phosphate coatings produced using a sputtering process--an alternative to plasma spraying. *Biomaterials*. 2005;26(3):327-337. doi:10.1016/j.biomaterials.2004.02.029
- Albrektsson T, Wennerberg A. Oral implant surfaces: Part 2--review focusing on clinical knowledge of different surfaces. *Int J Prosthodont*. 2004;17(5):544-564.



BPI

Retrospective Study

A RETROSPECTIVE MULTICENTRIC STUDY OF 56 PATIENTS TREATED WITH 92 PTERYGOID IMPLANTS FOR PARTIAL/ FULL ARCH IMPLANT SUPPORTED FIXED REHABILITATION: IMPLANT AND PROSTHESIS SUCCESS RATE

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ABSTRACT

In the case of severe atrophic patients, the search for native bone can be extended beyond the anatomical limits of the oral cavity. So remote anchorage solutions could involve the pterygomaxillary complex composed of the maxillary tuberosity, the pyramidal process of the palatine bone and the pterygoid pillar. Pterygoid implants are typically placed in this zone to rehabilitate patients affected by severe maxillary atrophy. This study's aim consists of the surgical and prosthetic success rate evaluation concerning the pterygoid implants placed to support fixed partial or full arch rehabilitation without a cantilever. All team members designed and conceived this retrospective multicenter study (performed in three different clinical offices) to evaluate the reliability and predictability of this anatomically guided surgical teenique without immediate loading. The study was successful with 100 per cent surgical success and all torque values >=45 N/cm considered as a threshold value. The series comprised 56 people who underwent 92 procedures. The male-to-female ratio

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ISSN: 2038-4106 Copyright © by BIOLIFE 2023 This publication and/or article is for individual use only and may not be further reproduced without written permission from the copyright holder. Unauthorized reproduction may result in financial and other penalties. **Disclosure: All authors report no conflicts of interest relevant to this article.** was close to one (27 men, 29 women). The mean age (\pm SD) was 64.0 \pm 9.3 years (range 41-85 years). Only one prosthetic failure was recorded in a woman aged 67 years receiving a full arch pterygoid implant. Pterygoid implants supported by fixed rehabilitation represent a reliable strategic solution for treating severe atrophic posterior maxilla.

KEYWORDS: pterygoid implants; cantilever free, insertion torque, fixed rehabilitation, atrophic maxilla, graftless surgery

INTRODUCTION

Osseointegrated implantology represents a reliable treatment solution to solve edentulism in jaws (1) in daily clinical practice. Insufficient bone amount and closeness to important anatomical landmarks could prevent implant placement. Each anatomical area is characterized by features and limitations (bone quality and quantity, nerve course, maxillary sinus cavity), which certainly conditioned/impacted this surgical procedure.

Among all, the atrophic posterior maxilla represents a critical and demanding area in the patient's rehabilitation through the insertion of integrated bone implants (2, 3) since it often lacks both in height and in thickness, thus preventing the placement of implants without adjunctive strategies (4).

The presence of the maxillary sinus, an inadequate bone in terms of quality or amount, a large fatty marrow space or the rare presence of cortical bone covering the alveolus represent some of the critical aspects that surgeons could meet during the surgical approach. Regenerative techniques such as maxillary sinus elevation, block grafts, or Customized Bone Regeneration allow bypassing these anatomical criticalities, even if they are not free from long healing periods or donor site morbidity (5-8). In implant surgery, it is mandatory to minimize patients' morbidity, especially if implant patients are getting older. Consequently, therapeutic, surgical procedures must be tailored to them and their ingrained features, systemic diseases, pharmacological therapies, and functional sinus impairment due to sinus lift augmentation (9). According to the current guidelines, daily clinical practice should consider the most cost-effective treatment equal to clinical efficacy.

Although surgical reliability is well documented, there is still disagreement on clinical and prosthetic primacy techniques. Some suggest it could be a good practice to go beyond these critical issues, using shorter and wider diameter implants to reach a high bone implant surface contact (10, 11). Furthermore, biomechanical considerations such as the intense chewing forces acting in the atrophic posterior maxilla should not be forgotten. Ideally, a prosthetic cantilever should be avoided for this aspect (12): several complications could occur, such as screw and framework fracture, marginal bone loss or implant osteointegration loss.

In the case of severe atrophic patients, the search for native bone can be extended beyond the anatomical limits of the oral cavity. So remote anchorage solutions could involve the pterygomaxillary complex composed of the maxillary tuberosity, the pyramidal process of the palatine bone and the pterygoid pillar. Pterygoid implants are typically placed in this zone to rehabilitate patients affected by severe maxillary atrophy (13).

Bone availability in the maxillary tuberosity is highly variable and is based mainly on the adjacent maxillary sinus pneumatization amount. In 1989, Tulasne (14) introduced implant placement in the pterygoid region to overcome anatomical limitations due to atrophic alveolar bone.

The pterygoid implant entails the fixture penetrating three specific osseous structures: maxillary tuberosity, the pyramidal process of the palatine bone and pterygoid pillar, and if it reaches osteointegration successfully, it offers support and stability to the final cantilever-free prosthesis. It significantly differs from tuberosity implant usually placed in the tuberosity region (mainly composed of 3 or 4 types of cancellous bone at the most distal portion of the maxillary alveolar process) and rarely with an angulation above 10 degrees. The pterygoid implants are usually placed with an angulation of 30 - 60 degrees relative to the horizontal maxillary plane, and they could offer support in partial and full arch prosthetic fixed rehabilitation. This anchorage satisfies surgeons and patients due to the time-consuming surgical strategy and favourable cost-benefit ratio.

The aim of this study consists of the surgical and prosthetic success rate evaluation concerning the pterygoid implants placed (with a minimum torque of 45 Ncm) to support fixed partial or full arch rehabilitation without a cantilever. Its proposal consolidates the literature evidence with our shared experience, whose data were analyzed and interpreted according to a characteristic descriptive statistical analysis.

MATERIAL AND METHODS

Study design

All team members designed and conceived this retrospective multicenter study with an enrolled sample of 92 pterygoid implants to evaluate the reliability and predictability of this anatomically guided surgical technique (Noris Medical PteriFit TM) with a 1-year follow-up. It was performed in three different clinical offices:

- 1. Dr Tealdo Tiziano Clinical Office, Alba, Italy;
- 2. Dr Bevilacqua Marco Clinical Office, Boves, Italy;
- 3. Dr Alberti Christian Clinical Office, Rosà, Italy.

Only one type of pterygoid implant (Noris Medical PteriFit [™]) was employed not to introduce further variables. All the patients previously visited after a CBCT 3D scan (Gendex GXDP-700 S) showed clinical and radiological signs of hopeless dentition and severe atrophy. After computer-assisted surgery planning (DTX Studio Clinic software, Nobel Biocare), the implant placement was defined in the pterygoid region. The study was conducted according to the Helsinki Declaration of 1975 principles and revised in 2000 for biomedical research involving human subjects.



Fig. 1a. Initial case of the atrophic patient in the maxillary arch.



Fig. 1b. 2D radiological images and 3D reconstruction of the same atrophic patient.

Since the authors analyzed preexisting and no identifiable data of patients, who were all informed about the nature of the data treatment and their written consent was obtained prior to participation.

Pterygoid rehabilitation protocol

All the patients enrolled in this study had to meet the inclusion criteria or good general health, no contraindications to implant placement or insufficient pterygoid bone amount (Fig. 1a, 1b, Fig. 2). All patients had at least 1 year of follow-up after the prosthesis delivery.

The surgical protocol applied to all the enrolled patients (January 2021 to February 2022) consisted of raising a full-thickness flap to expose the pterygomaxillary synostosis and performing the osteotomy for implant placement according to the manufacturer's guidelines. The implant site



Fig. 2. *Initial case of the patient from the occlusal point of view.*

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preparation sequence included a marking drill, the subsequent passage of a manual osteotome with a 2mm tip to define the implant insertion axis, the use of a 2.3mm diameter twist drill at approximately 1000 RPM, a second 2.8 mm diameter twist drill along the entire working length. The implant insertion was manually performed using a dedicated straight screwdriver.

Manual insertion did not allow the implant insertion torque to be objectively quantified; therefore, a torque wrench was fitted to accept the achievement of a torque of 45 Ncm or greater. Unstable pterygoid implants were immediately removed, and the osteotomy was filled with a hemostatic gelatin sponge (Spongostan -Ethicon). Since the Noris Medical PteriFit TM is soft-tissue level fixture, part of the stained neck of the implant was purposely left with extra bone to contract the relationship with the soft tissues in the tuber area.

The axis of the implants was corrected during the surgery by connecting a pre-angled conical abutment at 30Ncm. Before suturing the flap, a healing cup at 10 Ncm was connected above the stump to achieve non-submerged healing. The one-stage solution offers to screw a 5 mm healing cap on the pterygoid implant immediately after the surgery without any risk of interference with the opposite teeth, as interference during mastication with the chewing forces could prejudge the primary stability, and a surgical failure may occur. Thus, it could be recommended to cover the head of the pterygoid implant with the flap after the implant placement (two-stage) (Fig. 3-5).

After a minimum period of 3 months without prosthetic load, the pterygoid implants were registered with the pick-up and open tray technique for the definitive rehabilitation, which envisaged their union with other implants inserted in the same period.



Fig. 3. Intro-operative picture after implant insertion and Multi Unit Abutment (MUA) connection on different kind of implants.



Fig. 4. *Intraoral picture after a healing period of 4 months.*

A specific quick-setting plaster (BF plaster - Dentaltorino, Italy) was used as impression material for fixed full arch rehabilitation. In the case of partial rehabilitation, in addition to the impression plaster to solidify the implants together, silicone was also used for the remaining fixed dental elements. The final prosthetic frameworks were tightened by a



Fig. 5. Frontal aspect of the provisional prosthesis delivery.



Fig. 6. Frontal aspect of the final prosthesis delivery supported by the pterygoid implants.

motor with a torque of 25Ncm on the pterygoid implants Unigrip[™] connection (Fig. 6, 7).

Study variables

This kind of implant differs from conventional dental implants according to their extra-oral anchorage. For this reason, all the Authors considered only two outcome variables for this study: surgical and prosthetic success rate. Concerning the surgical success rate, only the pterygoid implants that reached a minimum of 45 Ncm insertion torque were considered and maintained in the pterygoid bone (otherwise, they were immediately removed during the surgical phase). The study was successful with 100 per cent surgical success and all torque values > =45 N/cm. The evaluated criteria to meet the prosthetic success rate were overall stability, comfort, function and patient acceptance. This last concept means that after prosthesis delivery, patients met satisfaction in chewing and phonetics without any excessive encumbrance or symptom. All the patients' feedback was collected and recorded during the follow-up dates planned after the prosthesis delivery.

Predictor variables

The following determinant or predictor variable was addressed in this study:

- a) demographic factors (gender, age) (Fig. 8);
- b) dental factors (size, length, diameter, MUA angle, torque insertion, surgical date, one or two-stage, number of implants, nasal implants, zygoma implants, partial/ full arch rehabilitation, prosthesis delivery) (Fig. 9).

RESULTS

Population under study

The series comprised 56 people who underwent 92 procedures. The male-to-female ratio was close to one (27 men, 29 women). The mean age (\pm SD) was 64.0 \pm 9.3 years (range 41-85 years). The primary endpoint was torque.

Surgical technique

The two-stage approach was used in nearly all patients. The one-stage approach was used in just one patient, a woman aged 74 years receiving a full arch pterygoid implant. Zygomatic implants were done in 15 patients (27%) and pasal implants in 10 (19%). Five patients had be

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Fig. 7. *A final Ortopantomography exam after the delivery of the final prosthesis.*



Fig. 8. Distribution of the age range between the genders.



Fig. 9. *Graph describing the distribution of the number of implants over genders (male/female).*

(27%), and nasal implants in 10 (19%). Five patients had both zygomatic and nasal implants.

Variable angulation was never considered. A full arch was used in most subjects (39/56=70%), while a partial arch was used in less than one-third (17/56 = 30%).

Patients in the series received 5.6 ± 1.4 implants overall (mean \pm SD) (range 2-8). Patients receiving partial arch had an

average of 4.3 ± 1.9 implants (median 4, range 2-8), while patients receiving full arch had an average of 6.1 ± 0.7 implants (median 6, range 5-8). Notably, two-thirds of the latter group (26/39) received 6 implants. The length of pterygoid implants ranges from 16 to 28 mm (median value 20.78 mm).

Surgical outcomes

The study succeeded with 100% surgical success and all torque values >=45 N/cm. Only one prosthetic failure was recorded in a woman aged 67 years receiving a full arch pterygoid implant.

DISCUSSION

In the case of severe atrophic posterior maxilla, the search for extraoral implant anchorages could represent a reliable strategy to restore and rehabilitate patients and prevent other alternative regenerative treatments (7, 15); in fact, the pterygoid implants play a crucial role in the reaching of extra oral bony pillar (rescue implants).

This retrospective study shows a success with 100 per cent surgical success and with all torque values > =45 N/ cm, even if other authors reported lower success rates for pterygoid implants (ranging from 80% to 99%) (10-16, 17). However, the surgical success rate we have observed should not mislead us into thinking it is a simple technique. This surgical approach requires operative skills and learning curves. The surgeon should recur to an accurate previous CBCT scan evaluation.

Clinicians should always consider that numerous vascular structures such as maxillary artery, descending palatine artery and pterygoid venous plexus can be detected in this area. Only with a detailed observation of pre-clinical CBCT can the placement of pterygoid implants be relatively safely planned. Up to now, three surgical techniques exist concerning pterygoid implant placement (18). The first is a free-hand surgical technique: we use this to plan and manage the pterygoid region. After a CBCT examination of the area to determine the correct axe insertion of the pterygoid implant, we expose the pterygoid-maxillary synostosis to access and approach the area. The surgeon can alternatively fold up a guided surgery, particularly a static fully guided implant placement (option #2) or a dynamic guided implant placement (option #3). For the static guide surgery, it is very important to consider the opening of the patient's mouth due to the encumbrance of the template and the dedicated drills (19); either technique requires continuous application and a constant learning curve to reach a well-established surgical skill.

This type of retrospective study requires a descriptive statistical analysis: the primary endpoint was the insertion torque; a value equal to or above 45 Ncm was the initial parameter considered. The authors want to underline the important prognostic value of the insertion torque (> =45 N/cm) on the surgical success rate. The primary stability is not always reachable during surgery. Whenever the insertion torque cannot satisfy the minimum of 45 Ncm, it is reccomended removing the implant to place another in another surgery date. An eventual prosthetic connection with nasal implants (10%) or zygomatic implants (27%) does not seem to play a prognostic decisive role. Even if these pterygoid implants differ from conventional intra-oral dental implants, they show a common feature: the importance of primary stability.

Furthermore, the length of pterygoid implants should be enough to allow these fixtures to engage the pterygoid process of the sphenoid bone. In the present study, implants of length ranging from 16 to 28 mm were used (median value 20.78 mm). The length of these implants is very closely related to primary stability and long-term success, as reported in the literature (16-20). Paying attention to all the surrounding anatomical determinants is mandatory in this situation.

It is possible to perform the one-stage surgery (5 mm height for the healing cap) only in safe conditions: at least 5 mm distance from the antagonist teeth. In case of interference during mastication, the chewing forces could prejudge the primary stability, and a surgical failure may occur. Compared to previous studies (20, 21), all the authors decided to redefine the clinical reliability of some parameters, such as:

- angulation of pterygoid implants: it was initially evaluated on an orthopantomography exam. In our opinion, a Cephalometric evaluation could be more indicated to estimate angulation than an Opt evaluation; it gives only an interpretation of the angulation: but would expose patients to further radiological exposure.
- bone loss: the pterygoid region represents a deep area for anchorage. All the authors consider estimating effective bone loss affecting pterygoid implants very challenging. To the best of our knowledge, the literature does not offer

solid support for scientific evidence on the calculation of bone loss around these implants. These are, unfortunately, empirical evaluations (21);

bleeding on probing (BoP): in this deep posterior area, the mucosal tunnel is deeper, and a possible BoP is not a
necessary sign of inflammation. Therefore, we cannot consider this biological parameter as reliable as dental implants;
If we consider this procedure from a prosthetic and biomechanical point of view, unscrewing and cantilever should
be prevented.

The unscrewing may occur if the screw is not tightened with a torque wrench (20 Ncm).

The mobility of the Multi Unit Abutment (MUA) resulting from unscrewing can induce bleeding, suppuration and tenderness and impact the function and satisfaction of the patient. Finally, the cantilever may play an unfavourable role in the overloading and consequent bone loss around the implants (20).

The bone loss was assessed in other studies (22, 23) comparing Opt exams scanned after 1 year of prosthetic loading. We argue that this calculation method is only interpretative but not scientifically reproducible and repeatable.

The postoperative healing phase of each patient did not have any particular signs or events worthy of note: bleeding could occur due to veins of the pterygoid muscles. These events could be stopped with the pterygoid implant placement. Patient acceptance of the distal prosthetic framework was high.

This retrospective study has only one prosthetic failure due to a partial fracture of the framework. As reported in Literature (24), our population under study confirmed high satisfaction with the fixed prostheses. No phonetic problems or speaking problems were referred. Correct and daily hygiene maintenance is mandatory to avoid high levels of plaque index, tissue hyperplasia or mucosal inflammation.

CONCLUSIONS

Fixed maxillary rehabilitations supported by Pterygoid implants represent an alternative reliable treatment solution for atrophic patients in the posterior maxilla; this anchorage allows the time reduction in the surgical procedure and the prosthesis restoration and favourably impacts the quality of the patient's life. This retrospective study met a surgical success of 100% with all torque values > =45 N/cm. Furthermore, these rehabilitation techniques are integrated with the digital flow up from the initial previsualization diagnostic phase, where the patient has real indications of final expectations.

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Conflict of Interest Statement:

All the Authors declare no conflict of interest.

REFERENCES

- Brånemark PI., Breine U, Adell R, Hansson BO, Lindström J, Ohlsson Å. Intra-Osseous Anchorage of Dental Prostheses: I. Experimental Studies. Scandinavian Journal of Plastic and Reconstructive Surgery. 1969;3(2):81-100. doi:https://doi.org/10.3109/02844316909036699
- 2. Adell R, Lekholm U, Rockler B, Brånemark PI. A 15-year study of osseointegrated implants in the treatment of the edentulous jaw. *International Journal of Oral Surgery*. 1981;10(6):387-416. doi:https://doi.org/10.1016/s0300-9785(81)80077-4
- 3. Albrektsson T, Zarb G, Worthington P, Eriksson AR. The long-term efficacy of currently used dental implants: a review and proposed criteria of success. *The International Journal of Oral & Maxillofacial Implants*. 1986;1(1):11-25.
- 4. Barone A, Santini S, Marconcini S, Giacomelli L, Gherlone E, Covani U. Osteotomy and membrane elevation during the maxillary sinus augmentation procedure. *Clinical Oral Implants Research*. 2008;19(5):511-515. doi:https://doi.org/10.1111/j.1600-0501.2007.01498.x
- 5. Adell R, Eriksson BO, Ulf Lekholm, Per-Ingvar Brånemark, Torsten Jemt. Long-term follow-up study of osseointegrated implants

in the treatment of totally edentulous jaws. PubMed. 1990;5(4):347-359.

- De Santis D, Umberto L, Dario D, et al. Custom Bone Regeneration (CBR): An Alternative Method of Bone Augmentation—A Case Series Study. J Clin Med. 2022;11(16):4739-4739. doi:https://doi.org/10.3390/jcm11164739
- 7. De Santis D, Gelpi F, Verlato G, et al. Digital Customized Titanium Mesh for Bone Regeneration of Vertical, Horizontal and Combined Defects: A Case Series. *Medicina (Kaunas)*. 2021;57(1):60-60. doi:https://doi.org/10.3390/medicina57010060
- 8. Wood RM, Moore DL. Grafting of the maxillary sinus with intraorally harvested autogenous bone prior to implant placement. *The International Journal of Oral & Maxillofacial Implants*. 1988;3(3):209-214.
- Schimmel M, Müller F, Suter V, Buser D. Implants for elderly patients. *Periodontology 2000*. 2016;73(1):228-240. doi:https:// doi.org/10.1111/prd.12166
- 10. Langer B, Langer L, Herrmann I, Jorneus L. The wide fixture: a solution for special bone situations and a rescue for the compromised implant. Part 1. *The International Journal of Oral & Maxillofacial Implants*. 1993;8(4):400-408.
- 11. Bahat O. Osseointegrated implants in the maxillary tuberosity: report on 45 consecutive patients. *The International Journal of Oral & Maxillofacial Implants*. 1992;7(4):459-467.
- Bevilacqua M, Tealdo T, Menini M, et al. The influence of cantilever length and implant inclination on stress distribution in maxillary implant-supported fixed dentures. *The Journal of Prosthetic Dentistry*. 2011;105(1):5-13. doi:https://doi.org/10.1016/ s0022-3913(10)60182-5
- Rieger MR. Loading Considerations for Implants. Oral and Maxillofacial Surgery Clinics of North America. 1991;3(4):795-804. doi:https://doi.org/10.1016/s1042-3699(20)30549-5
- 14. Tulasne J. Osseointegrated fixtures in the pterygoid region. In: Advanced Osseointegration Surgery. Applications in the Maxillofacial Region. Quintessence; 1992:182-188.
- 15. Boyne PJ. Augmentation of the posterior maxilla by way of sinus grafting procedures: recent research and clinical observations. *Oral and Maxillofacial Surgery Clinics of North America*. 2004;16(1):19-31. doi:https://doi.org/10.1016/j.coms.2003.10.006
- Bidra AS, May GW, Tharp GK, Chambers MS. Pterygoid Implants for Maxillofacial Rehabilitation of a Patient With a Bilateral Maxillectomy Defect. *J Oral Implantol*. 2013;39(1):91-97. doi:https://doi.org/10.1563/aaid-joi-d-10-00181
- 17. Candel E, Peñarrocha D, Peñarrocha M. Rehabilitation of the Atrophic Posterior Maxilla With Pterygoid Implants: A Review. *Journal of Oral Implantology*. 2012;38(S1):461-466. doi:https://doi.org/10.1563/aaid-joi-d-10-00200
- Bidra AS, Peña-Cardelles J, Iverson M. Implants in the pterygoid region: An updated systematic review of modern roughened surface implants. *Journal of Prosthodontics*. 2022;32(4). doi:https://doi.org/10.1111/jopr.13600
- 19. De Santis D, Graziani P, Castellani R, et al. A New Radiologic Protocol and a New Occlusal Radiographic Index for Computer-Guided Implant Surgery. *Journal of Craniofacial Surgery*. 2016;27(5):e506-e510. doi:https://doi.org/10.1097/scs.00000000002490
- Jaemsuwan S, Arunjaroensuk S, Kaboosaya B, Subbalekha K, Mattheos N, Pimkhaokham A. Comparison of the accuracy of implant position among free-hand implant placement, static and dynamic computer-assisted implant surgery in fully edentulous patients: a non-randomized prospective study. *International Journal of Oral and Maxillofacial Surgery*. 2022;52(2). doi:https:// doi.org/10.1016/j.ijom.2022.05.009
- Curi MM, Cardoso CL, Ribeiro K de CB. Retrospective Study of Pterygoid Implants in the Atrophic Posterior Maxilla: Implant and Prosthesis Survival Rates Up to 3 Years. *The International Journal of Oral & Maxillofacial Implants*. 2015;30(2):378-383. doi:https://doi.org/10.11607/jomi.3665
- 22. Bevilacqua M, Tealdo T, Pera F, et al. Three-dimensional finite element analysis of load transmission using different implant inclinations and cantilever lengths. *The International Journal of Prosthodontics*. 2008;21(6):539-542.
- 23. Peñarrocha M, Carrillo C, Boronat A, Peñarrocha M. Retrospective study of 68 implants placed in the pterygomaxillary region using drills and osteotomes. *The International Journal of Oral & Maxillofacial Implants*. 2009;24(4):720-726.
- 24. Balshi SF, Wolfinger GJ, Balshi TJ. Analysis of 164 titanium oxide-surface implants in completely edentulous arches for fixed prosthesis anchorage using the pterygomaxillary region. *The International Journal of Oral & Maxillofacial Implants*. 2005;20(6):946-952.



Case report

UNDIFFERENTIATED CONNECTIVE TISSUE DISEASE WITH HYPERPLASIA OF YELLOW LIGAMENTS IN L4-L5 CAUSING SEGMENTAL SPINAL STENOSIS

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ABSTRACT

Undifferentiated connective tissue disease (UCTD) is a systemic autoimmune disease characterized by clinical and serological manifestations not fulfilling the criteria for defined connective tissue diseases. Up to 90% of the cases are young women. Usually, UCTD has a mild clinical course with a wide variety of signs and symptoms because it can involve any connective tissue in the body. 40% of patients with UCTD develop the stage of a well-defined systemic autoimmune disease during five years of follow-up, while 60% remain in an undifferentiated stage. The most used drugs in treating UCTD are nonsteroidal anti-inflammatory drugs, corticosteroids, calcium channel blockers, and antimalarial drugs. We report a rare case of a woman with UCTD in corticosteroid treatment, suffering from low back pain refractory to therapy, evidence a computed tomography (CT) of abnormal bone hyperplasia of the yellow ligament conditioning spinal stenosis.

KEYWORDS: undifferentiated connective tissue disease; yellow ligaments; spinal stenosis; systemic autoimmune diseases

INTRODUCTION

Undifferentiated connective tissue disease (UCTD) is a systemic autoimmune disease characterized by clinical and serological manifestations not fulfilling the criteria for defined connective tissue diseases (CTD) such as systemic lupus erythematosus, mixed connective tissue disease, Sjögren syndrome, systemic sclerosis, polymyositis, dermatomyositis, or rheumatoid arthritis (1, 2). Its diagnosis is considered exclusion (3). UCTD is defined if the following criteria are met:

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signs and symptoms suggestive of a CTD, but not fulfilling criteria for a defined CTD, positive antinuclear antibodies on two separate measurement controls, and disease of duration of at least 3 years (4). The incidence is unknown due to the lack of a proper definition of this pathology, but it has been observed that 20-50% of patients presenting in a rheumatology department have a UCTD diagnosis (5).

From an epidemiological point of view, in more than 90% of cases, this pathology mainly affects women, particularly those between 32 and 44 years old (6, 7). There are two forms of UCTD called stable-UCTD and evolving-UCTD (4). The first case represents the forms that remain undifferentiated and are over 60% of the total, while the evolving forms represent about 40% of the UCTD evolve into defined systemic autoimmune disease during five years follow up (8, 9). Like all autoimmune diseases, the aetiology is unknown; what is known is that genetic factors and environmental triggers induce the triggering of these diseases (10). However, UCTD, like other known connective tissue disorders, is characterized by exaggerated immune system activity (11). The latter produces autoantibodies or activates antigen-specific T-lymphocytes that affect connective tissue at every site of the body (12).

Clinically, UCTD has a generally mild course; in most cases, it is characterized by the absence of severe organ damage or involvement, especially in the renal and neurological systems (3). The main symptoms are arthralgia, which can be present in more than 86% of patients; various skin lesions such as livedo, purpura, acrocyanosis, telangiectasias, urticaria (3, 7); Raynaud phenomenon (33%), sicca symptoms (30%), mucocutaneous symptoms included oral ulcers (23%); arthritis (22%) and thyroid disease (7%) (13-18). Constitutional symptoms, such as fever, malaise, and fatigue, are often the initial presentation of the disease (3). From a diagnostic point of view, serological markers are considered essential in the diagnostic criteria for UCTD (3). In particular, anti-Ro/SSA and anti-U1-RNP are considered markers detected in this disease (3, 5).

Regarding imaging studies, chest radiography and computed tomography (CT) can be helpful in studying cardiac and pulmonary involvement (19, 20). Additionally, ultrasonography of the salivary glands was a good test in differentiating between UCTD and other diseases such as Sjögren syndrome (21, 22). The main treatment of UCTD is pharmacological, and the most used drugs are nonsteroidal anti-inflammatory drugs, corticosteroids, calcium channel blockers and antimalarial drugs such as hydroxychloroquine (3, 23, 24). If the disease is not controlled with these drugs or the symptoms are severe, it is necessary to use immunosuppressive agents such as methotrexate and azathioprine (3, 7).

CASE PRESENTATION

A 75-year-old female patient with a medical history that revealed UCTD was receiving corticosteroid treatment for 15 years.

Since the age of 65, the woman had episodes of neck and low back pain investigated with a rachis x-ray showing diffuse spondyloarthritis manifestations; therefore, she was treated with nonsteroidal anti-inflammatory drugs to reduce the pain symptoms.

For about 60 days, she complained of acute left lumbosciatica with paresthesia and weakness of the left lower limb conditioning intermittent claudication. Furthermore, this symptomatology was not responsive to pain-relieving therapy.

The patient underwent lumbosacral computed tomography (CT) (Fig. 1) that showed marked hyperplasia of yellow ligaments at the fourth lumbar disc and



Fig. 1. a), b), c): Marked hyperplasia of the yellow ligaments at L4-L5 (red arrows), which determines important segmental canal stenosis, where the dural sac appears compressed and displaced anteriorly against the posterior wall of L4 and L5. The disc between L4 and L5 appea rs modestly protruded circumferentially (blue arrow) with associated gaseous vacuolar degeneration of the nucleus pulposus (green arrow).

fifth lumbar disc (L4-L5). This hyperplasia determines important segmental canal stenosis, where the dural sac appears compressed and displaced anteriorly against the posterior wall of L4 and L5. Moreover, the intervertebral disc between L4 and L5 appears modestly protruded circumferentially with associated gaseous vacuolar degeneration of the nucleus pulposus. Hyperplasia and subsequent ossification of the yellow ligaments is a rare event that can occur in cases of UCTD.

DISCUSSION

The case report shows a rare hyperplasia of the yellow ligaments that condition root canal stenosis associated with UCTD. As known, the yellow ligament is located inside the spinal canal that connects posterolaterally two laminae of adjacent vertebrae, and it is divided into two portions: capsular portion and interlaminar portion (25, 26). Histologically, connective tissue comprises 80% elastic fibres and 20% collagen fibre (27). It maintains the inherent stability of the spine, controlling intervertebral movement and maintaining a smooth surface of the posterior dural sac (26, 27).

The pathogenesis of the yellow ligament's thickening is unclear (28). Multifactorial agents such as age, mechanical stress, growth factors and systemic disease like connective tissue diseases are known to contribute to hyperplasia development up to ossification of yellow ligaments (28-30). The hypertrophied yellow ligament shows an increase in collagen fibres, calcification, ossification and chondrometaplasia (27); this is due to the production by the cells present in the yellow ligament of a high volume of type II collagen at the expense of elastic fibres (28-30). Subsequently, this collagen is converted to type-1, which can lead to endochondral ossification of the yellow ligament (28-30).

Thickening of the yellow ligaments causes spinal canal narrowing and mechanical compression of the nerve roots, cauda equina and spinal cord (31). This mechanical compression causes low back pain, sciatica, paresthesia, pain and muscle weakness, gait disturbance and bladder-bowel disturbance (26). These symptoms occur even in the absence of bulging *Annulus fibrosus* and herniated nucleus pulposus (26).

Diagnosis is based on the neurological findings, imaging examinations using X-ray, CT and magnetic resonance imaging (MRI) and electrophysiological examinations (32). Generally, the treatment of symptoms brings pain and numbness, and the pain in the lower extremities requires the use of nonsteroidal anti-inflammatory drugs, muscle relaxants and vitamin B12 (26). Physical therapy is recommended at an early stage.

However, dynamic physical therapy, such as massage and stretching of the spine by others, is contraindicated because it increases the risk of hypertrophic yellow ligament injury. Surgical treatment is recommended for patients with ineffective conservative treatment and with severe spastic gait, severe muscle weakness of the lower extremities, and bladder-bowel disturbance. Surgical decompression methods include open-door laminectomy, bulk laminectomy, fenestration, and hemilaminectomy (33-37).

CONCLUSIONS

Hyperplasia of the yellow ligaments and subsequent ossification are rare events of multifactorial aetiology due to advanced age, mechanical stress and systemic disease. Among the pathologies that could cause an alteration of the structural composition and, therefore, the yellow ligaments' functionality are connective tissue diseases, but insufficient data are currently available in the literature. Future studies will be needed to evaluate the relationship between connective tissue diseases such as UCTD and yellow ligament hypertrophy. Studies will then be necessary to make diagnoses with in-depth and targeted imaging techniques to evaluate the spinal column and the subsequent and appropriate medical or surgical therapy.

REFERENCES

- Antunes M, Scirè CA, Talarico R, et al. Undifferentiated connective tissue disease: state of the art on clinical practice guidelines. *RMD Open*. 2019;4(Suppl 1):e000786. doi:https://doi.org/10.1136/rmdopen-2018-000786
- 2. Dyball S, Rodziewicz M, Mendoza-Pinto C, Bruce IN, Parker B. Predicting progression from undifferentiated connective tissue

disease to definite connective tissue disease: A systematic review and meta-analysis. *Autoimmunity Reviews*. 2022;21(11):103184. doi:https://doi.org/10.1016/j.autrev.2022.103184

- 3. Marwa K, Anjum F. Undifferentiated Connective Tissue Disease. *StatPearls*. Published online April 27, 2023. https://www.statpearls.com/point-of-care/87459
- Rubio J, Kyttaris VC. Undifferentiated Connective Tissue Disease: Comprehensive Review. *Curr Rheumatol Rep.* 2023;25(5). doi:https://doi.org/10.1007/s11926-023-01099-5
- Mosca M, Tani C, Bombardieri S. Undifferentiated connective tissue diseases (UCTD): a new frontier for rheumatology. *Best Practice & Research Clinical Rheumatology*. 2007;21(6):1011-1023. doi:https://doi.org/10.1016/j.berh.2007.09.004
- 6. Spinillo A, Beneventi F, Caporali R, Ramoni V, Montecucco C. Undifferentiated connective tissue diseases and adverse pregnancy outcomes. An undervalued association? *Am J Reprod Immunol*. 2017;78(6):e12762-e12762. doi:https://doi.org/10.1111/aji.12762
- Serena C, Clemenza S, Simeone S, et al. Undifferentiated Connective Tissue Disease in Pregnancy: A Topic Yet to be Explored. Frontiers in Pharmacology. 2022;13. doi:https://doi.org/10.3389/fphar.2022.820760
- 8. Mosca M, Tani C, Vagnani S, Carli L, Bombardieri S. The diagnosis and classification of undifferentiated connective tissue diseases. *Journal of Autoimmunity*. 2014;48-49:50-52. doi:https://doi.org/10.1016/j.jaut.2014.01.019
- 9. García-González M, Rodríguez-Lozano B, Bustabad S, Ferraz-Amaro I. Undifferentiated connective tissue disease: predictors of evolution into definite disease. *Clinical and Experimental Rheumatology*. 2017;35(5):739-745.
- 10. Mosca M, Tani C, Talarico R, Bombardieri S. Undifferentiated connective tissue diseases (UCTD): Simplified systemic autoimmune diseases. *Autoimmunity Reviews*. 2011;10(5):256-258. doi:https://doi.org/10.1016/j.autrev.2010.09.013
- 11. Nakken B, Bodolay E, Szodoray P. Cytokine Milieu in Undifferentiated Connective Tissue Disease: a Comprehensive Review. *Clin Rev Allergy Immunol* . 2014;49(2):152-162. doi:https://doi.org/10.1007/s12016-014-8452-9
- Cordiali-Fei P, Mussi A, D'Agosto G, et al. Assessment of T Regulatory Cells and Expanded Profiling of Autoantibodies May Offer Novel Biomarkers for the Clinical Management of Systemic Sclerosis and Undifferentiated Connective Tissue Disease. *Clin Dev Immunol.* 2013;2013:1-7. doi:https://doi.org/10.1155/2013/390563
- 13. De Angelis R, Cerioni A, Del Medico P, Blasetti P. Raynaud's phenomenon in undifferentiated connective tissue disease (UCTD). *Clin Rheumatol.* 2004;24(2):145-151. doi:https://doi.org/10.1007/s10067-004-0988-2
- 14. Vaz CC, Couto M, Medeiros D, et al. Undifferentiated connective tissue disease: a seven-center cross-sectional study of 184 patients. *Clinical Rheumatology*. 2009;28(8):915-921. doi:https://doi.org/10.1007/s10067-009-1175-2
- 15. Bodolay E, Csiki Z, Szekanecz Z, et al. Five-year follow-up of 665 Hungarian patients with undifferentiated connective tissue disease (UCTD). *Clin Exp Rheumatol*. 2003;21(3):313-320.
- 16. Danieli MG, Fraticelli P, Franceschini F, et al. Five-year follow-up of 165 Italian patients with undifferentiated connective tissue diseases. *Clinical and Experimental Rheumatology*. 1999;17(5):585-591.
- 17. Alarcón GS, Willkens RF, Ward JR, et al. Early undifferentiated connective tissue disease. IV. Musculoskeletal manifestations in a large cohort of patients with undifferentiated connective tissue diseases compared with cohorts of patients with well-established connective tissue diseases: Follow-up analyses in patients with unexplained polyarthritis and patients with rheumatoid arthritis at baseline. *Arthritis & Rheumatism.* 1996;39(3):403-414. doi:https://doi.org/10.1002/art.1780390308
- Danieli MG, Rossetti L, Fraticelli P, Malcangi G, Testa I, Danieli G. Autoimmune Thyroid Diseases in Patients with Undifferentiated Connective Tissue Disease. *Clinical Rheumatology*. 2000;19(1):42-46. doi:https://doi.org/10.1007/s100670050009
- 19. Kim HC, Ji W, Kim MY, et al. Interstitial Pneumonia Related to Undifferentiated Connective Tissue Disease: Pathologic Pattern and Prognosis. *Chest*. 2015;147(1):165-172. doi:https://doi.org/10.1378/chest.14-0272
- 20. Yoo H, Hino T, Hwang J, et al. Connective tissue disease-related interstitial lung disease (CTD-ILD) and interstitial lung abnormality (ILA): Evolving concept of CT findings, pathology and management. *European Journal of Radiology Open*. 2022;9:100419-100419. doi:https://doi.org/10.1016/j.ejro.2022.100419
- Luciano N, Baldini C, Tarantini G, et al. Ultrasonography of major salivary glands: a highly specific tool for distinguishing primary Sjögren's syndrome from undifferentiated connective tissue diseases. *Rheumatology (Oxford)*. 2015;54(12):kev253kev253. doi:https://doi.org/10.1093/rheumatology/kev253
- La Paglia GMC, Sanchez-Pernaute O, Alunno A, et al. Ultrasound salivary gland involvement in Sjogren's syndrome vs. other connective tissue diseases: is it autoantibody and gland dependent? *Clinical Rheumatology*. 2019;39(4):1207-1215. doi:https:// doi.org/10.1007/s10067-019-04780-2
- 23. Yang SY, Ni R, Lu Y, et al. A three-arm, multicenter, open-label randomized controlled trial of hydroxychloroquine and lowdose prednisone to treat recurrent pregnancy loss in women with undifferentiated connective tissue diseases: protocol for the Immunosuppressant regimens for LIving FEtuses (ILIFE) trial. *Trials*. 2020;21(1). doi:https://doi.org/10.1186/s13063-020-04716-1
- 24. Mosca M, Tani C, Bombardieri S. A case of undifferentiated connective tissue disease: is it a distinct clinical entity? *Nature Clinical Practice Rheumatology*. 2008;4(6):328-332. doi:https://doi.org/10.1038/ncprheum0799
- 25. Liang H, Liu G, Lu S, et al. Epidemiology of ossification of the spinal ligaments and associated factors in the Chinese population: a cross-sectional study of 2000 consecutive individuals. *BMC Musculoskeletal Disorders*. 2019;20(1). doi:https://doi.org/10.1186/ s12891-019-2569-1
- Hirabayashi S. Ossification of the ligamentum flavum. Spine Surgery and Related Research. 2017;1(4):158-163. doi:https://doi. org/10.22603/ssrr.1.2016-0031
- 27. Salimi H, Suzuki A, Habibi H, et al. Biglycan expression and its function in human ligamentum flavum. *Sci Rep.* 2021;11(1). doi:https://doi.org/10.1038/s41598-021-84363-x
- 28. Sun C, Zhang H, Wang X, Liu X. Ligamentum flavum fibrosis and hypertrophy: Molecular pathways, cellular mechanisms, and future directions. *FASEB J*. 2020;34(8):9854-9868. doi:https://doi.org/10.1096/fj.202000635r
- 29. Amudong A, Muheremu A, Abudourexiti T. Hypertrophy of the ligamentum flavum and expression of transforming growth factor beta. *J Int Med Res.* 2017;45(6):2036-2041. doi:https://doi.org/10.1177/0300060517711308
- Ikuta M, Kaito T, Fujimori T, et al. Review of Basic Research about Ossification of the Spinal Ligaments Focusing on Animal Models. J Clin Med. 2023;12(5):1958-1958. doi:https://doi.org/10.3390/jcm12051958
- Uchida K, Nakajima H, Yayama T, Sato R, Baba H. [Updates on ossification of posterior longitudinal ligament. Ossification front of posterior longitudinal ligament and cellular biological assessment of chronic mechanical compressed spinal cord]. *Clinical Calcium*. 2009;19(10):1472-1479.
- Takahashi T, Junya Hanakita, Minami M. Pathophysiology of Calcification and Ossification of the Ligamentum Flavum in the Cervical Spine. *Neurosurg Clin N Am.* 2018;29(1):47-54. doi:https://doi.org/10.1016/j.nec.2017.09.016
- Hirabayashi S, Kitagawa T, Yamamoto I, Yamada K, Kawano H. Development and Achievement of Cervical Laminoplasty and Related Studies on Cervical Myelopathy. *Spine Surgery and Related Research*. 2020;4(1):8-17. doi:https://doi.org/10.22603/ ssrr.2019-0023
- Hirabayashi S, Kitagawa T, Yamamoto I, Yamada K, Kawano H. Surgical Treatment for Ossification of the Posterior Longitudinal Ligament (OPLL) at the Thoracic Spine: Usefulness of the Posterior Approach. *Spine Surg Relat Res.* 2018;2(3):169-176. doi:https://doi.org/10.22603/ssrr.2017-0044
- 35. Zhong ZM, Wu Q, Meng T, et al. Clinical outcomes after decompressive laminectomy for symptomatic ossification of ligamentum flavum at the thoracic spine. *J Clin Neurosci*. 2016;28:77-81. doi:https://doi.org/10.1016/j.jocn.2015.09.023
- Ando K, Imagama S, Ito Z, et al. Predictive Factors for a Poor Surgical Outcome With Thoracic Ossification of the Ligamentum Flavum by Multivariate Analysis. *Spine*. 2013;38(12):E748-E754. doi:https://doi.org/10.1097/brs.0b013e31828ff736
- Aizawa T, Sato T, Sasaki H, Kusakabe T, Morozumi N, Kokubun S. Thoracic myelopathy caused by ossification of the ligamentum flavum: clinical features and surgical results in the Japanese population. *Journal of Neurosurgery: Spine*. 2006;5(6):514-519. doi:https://doi.org/10.3171/spi.2006.5.6.514





Case reports

SUB-CRESTAL IMPLANTS WITH PLATFORM-SWITCHING AND ONE TIME ABUTMENT

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ABSTRACT

The use of dental implants in the rehabilitation of partially or fully edentulous patients is a treatment that has been validated over the last 40 years, with a high success rate. The introduction of platform switching, i.e. the use of abutments with a smaller diameter than the implant neck, has also resulted in an important benefit in terms of biomechanical behaviour, influence on crestal bone and peri-implant soft tissue response. A series of cases using BioPlatform GTB implants in different situations is presented.

KEYWORDS: *short implants, platform switching, one time abutment*

INTRODUCTION

The basis of medium- and long-term rehabilitation success in all implant systems is the integration of the abutmentfixture complex with the surrounding bone tissue. An integration that must be of sufficient quality and quantity and remain stable over time (1).

In particular, the clinical and radiological evaluation of marginal bone loss is considered one of the key factors for the stability and longevity of dental implants, as well as the maintenance of peri-implant soft tissue.

The establishment of a pathogenic microflora at the abutment-fixture interface, with the possible onset of mucositis, the increase in pocket depth and progressive bone resorption, as well as the role of excessive biomechanical stress due to incorrect occlusal loading, are related factors implicated in the loss of marginal bone around dental implants (2-5).

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Over the years, attention has been focused on the role of the position of the neck of the fixture in relation to the marginal ridge, the type, the geometry and the timing of the abutment-fixture connection. Several studies have focused attention on the role of the type of implant-abutment connection that can contribute to the stability of the peri-implant bone level; in fact, the geometry of the connection influences possible bacterial colonisation within the implants (6, 7).

The internal connection seems to show better results in terms of prevention of microbial penetration, resulting in a tight marginal seal and implant stability, thus preventing marginal bone loss (8-10).

Nowadays, the evaluation of those treatments in which abutments with a smaller diameter than the fixture have been used has revealed a better preservation of hard and soft tissues compared to treatments using abutments with similar diameters to the implant (2, 11, 12).

In recent years, developments in the macro-geometries of dental implants and prosthetic components have allowed a considerable increase in the biological performance of dental implants, with a paradigm shift in the surgical approach and implant-prosthetic rehabilitation (1).

The use of 'short' implants, < or = 6 mm in length, has thus become a predictable therapeutic alternative, capable at times of avoiding bone regeneration procedures that are certainly more complex, of longer duration and with a more uncertain or operator-dependent prognosis.

Research interest is therefore focusing on the comparison of marginal, heavily loaded bone with unfavourable levers and crown-radicular ratios (2:1 or more) (10-12).

Case 1

A female patient, 55 years old, non-smoker, with good oral hygiene control was admitted to our department. She had monoedentulous first upper right molar for more than 6 months. On radiological evaluation with TCCB, she had 7 mm of bone thickness in the buccal vestibular direction and a distance of 6 mm from the lower sinus floor. The proposed treatment plan was the insertion of a 4.3 mm diameter and 6 mm long GTB implant fixture, after transcrestal sinus elevation.

The surgical planning was carried out according to the surgical protocol for GTB implants, which provides for the eventual reduction of the 'knifeedge' ridge, the placement of fixtures according to the prosthetic axis, and the placement of the implant at a subcrestal level of at least 1.5 to 2 mm.

This planning was then performed surgically according to protocol, achieving a screwing stability of 25 Ncm. At the same time, the healing abutment GFA, with a height of 4.5 mm, was placed over the fixture.

After 60 days, and radiographic control, the impression was obtained was taken by unscrewing the healing abutment of the GFA according to the "one time abutment" protocol, screwed with a torque of 25 Ncm according to the GTB prosthetic protocol. The definitive polyether impression by using a transfer screwed directly onto the GFA abutment; a metal ceramic crown was then delivered.

At check-up 6 months after definitive loading, the bone closure on the neck of the abutment is complete, visible both radiologically and by the absence of probing.





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Case 2

A 54-year-old male patient presented with mono-edentulous zone 2.4, an endosseous implant was inserted with a diameter of 3.3 and a length of 10 mm, but when the prosthetic abutment was tightened to about 25 N the implant fixture rotated. It was decided to carry out a new osseointegration but after 3 weeks a mucositis and peri-implant resorption process appeared. It was decided to remove the implant and proceed with a new contextual insertion. We chose the insertion of a GTB 3.3 10 mm implant with subcrestal placement, to achieve primary stability and optimal healing of the peri-implant mucosal tissue thanks to the use of the GFA component and the one-time abutment. A good healing of the hard tissues and the peri-implant marginal mucosal tissues with satisfactory pink aesthetics and bone stability was obtained at the time of the final radiographic check (70 days).

Case 3

A 60-year-old female patient presented with periodontal compromised tooth 4.7 such that extraction was necessary. After three months a GTB 4.3 x 7.5 mm length implant fixture was inserted and a 3.5 mm GFA placed. Two months after the surgical phase an impression was taken and a definite crown was inserted, following the indications for a pontic crown that maintains the stability and quality of the keratinized gingiva. There is excellent integration both with the surrounding teeth (white aesthetics) and in the stability of the peri-implant soft tissue (pink aesthetics).



Case 2.

Case 3.

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Case 4

A 30-year-old patient with mono-edentulous zone 1.2 due to a previous root crown fracture presented to us for prosthetic rehabilitation. He had a thin biotype and the smile-gum which makes both implant insertion and the aesthetic result very difficult. It is decided to insert a 3.3-diameter and 9-mm-high implant with 3.5-mm GFA. At the end of the surgical phase, a corrected positioning of the GFA level with the gingival margin but after waiting for the osseointegration phenomenon and at the moment of taking the impression the gingival margin seemed to have migrated apically requiring a substitution of a 2 mm GFA abutment.

After the correct selection of the GFA the temporary crown has integrated correctly with the peri-implant soft tissue with an aesthetic result satisfactory.



Case 4.

DISCUSSION

Marginal bone loss around dental implants has been attributed to several factors. It may be the result of the establishment of a pathogenic microflora, which promotes the onset of peri-implant disease with mucosal inflammation, increased pocket depth and progressive bone resorption. Other studies have suggested that changes in marginal bone level may be the result of biomechanical stress due to incorrect occlusal design (6, 9, 13, 14).

Crestal bone loss may be the physiological result of incorrect three-dimensional positioning of the fixture. The coronal portion of the bone may tend to resorb if the fixture is placed too close to adjacent teeth/implants or a thin residual buccal wall (10, 15).

Subcrestal placement of the implant platform may negatively influence the stability of the peri-implant marginal bone. Crestal bone resorption is also related to the presence of a microgap between implant and abutment and the position of this microgap in relation to the crestal bone level.

The microgap, the micromovement between the fixture and the abutment, and the presence or absence of the switching platform are therefore considered the main factors in marginal bone resorption (2, 5, 9, 12).

Respect of the surgical protocol with a subcrestal positioning and a screwing torque of the implants that is not excessive and the control of the implant insertion axis, which must be as coincident as possible with the prosthetic axis, are key factors in the long-term success of implant rehabilitation (1, 10).

CONCLUSIONS

A conical or cono-morse implant-abutment connection allows subcrestal placement of the fixture, with a substantial reduction in the risk of microbial colonisation and/or micromovement, both negative prognostic factors for marginal bone maintenance.

It is even more evident how the use of abutments with a smaller diameter than the fixture (platform switching) and their early and single insertion (one-time abutment) both contribute to preserving the mucosal bone tissue complex around the implant, positively influencing the prognosis and success of therapy (1, 16, 17).

REFERENCES

1. Wang Q, Dai R, Cao CY, Fang H, Han M, Li QL. One-time versus repeated abutment connection for platform-switched implant:

A systematic review and meta-analysis. Bencharit S, ed. PLOS ONE. 2017;12(10):e0186385. doi:https://doi.org/10.1371/journal. pone.0186385

- Alves C, Muñoz F, González Cantalapiedra A, Ramos I, Neves M, Blanco JA. Marginal bone and soft tissue behavior following platform switching abutment connection/disconnection- a dog model study. *Clin Oral Implants Res.* 2014;26(9):983-991. doi:https://doi.org/10.1111/clr.12385
- Baggi L, Di Girolamo M, Mirisola C, Calcaterra R. Microbiological Evaluation of Bacterial and Mycotic Seal in Implant Systems With Different Implant-Abutment Interfaces and Closing Torque Values. *Implant Dentistry*. 2013;22(4):344-350. doi:https://doi. org/10.1097/id.0b013e3182943062
- Vairo G, Sannino G. Comparative Evaluation of Osseointegrated Dental Implants Based on Platform-Switching Concept: Influence of Diameter, Length, Thread Shape, and In-Bone Positioning Depth on Stress-Based Performance. *Computational and Mathematical Methods in Medicine*. 2013;2013:1-15. doi:https://doi.org/10.1155/2013/250929
- 5. Meynardi F. Implant dentistry: monitoring of bacteria along the transmucosal passage of the healing screw in absence of functional load. *Oral & Implantology*. 2016;9(Suppl. 1):10. doi:https://doi.org/10.11138/orl/2016.9.1s.010
- Becker K, Mihatovic I, Golubovic V, Schwarz F. Impact of abutment material and dis-/re-connection on soft and hard tissue changes at implants with platform-switching. *Journal of Clinical Periodontology*. 2012;39(8):774-780. doi:https://doi.org/10.1111/j.1600-051x.2012.01911.x
- Rodríguez X, Vela X, Méndez V, Segalà M, Calvo-Guirado JL, Tarnow DP. The effect of abutment dis/reconnections on periimplant bone resorption: A radiologic study of platform-switched and non-platform-switched implants placed in animals. *Clinical Oral Implants Research*. 2011;24(3):305-311. doi:https://doi.org/10.1111/j.1600-0501.2011.02317.x
- Baggi L, Cappelloni I, Di Girolamo M, Maceri F, Vairo G. The influence of implant diameter and length on stress distribution of osseointegrated implants related to crestal bone geometry: A three-dimensional finite element analysis. *The Journal of Prosthetic Dentistry*. 2008;100(6):422-431. doi:https://doi.org/10.1016/s0022-3913(08)60259-0
- 9. Di Girolamo M. 3D X-ray microscopic analysis on a prosthetically loaded implant with platform-switching and conical connection: a case report. *Oral & Implantology*. 2017;10(3):241. doi:https://doi.org/10.11138/orl/2017.10.3.241
- Iezzi G, Iaculli F, Calcaterra R, Piattelli A, Di Girolamo M, Baggi L. Histological and Histomorphometrical Analysis on a Loaded Implant With Platform-Switching and Conical Connection: A Case Report. *Journal of Oral Implantology*. 2017;43(3):180-186. doi:https://doi.org/10.1563/aaid-joi-d-16-00182
- DI Girolamo M, Calcaterra R, DI Gianfilippo R, Arcuri C, Baggi L. Bone level changes around platform switching and platform matching implants: a systematic review with meta-analysis. ORAL & implantology. 2016;9(1):1-10. doi:https://doi.org/10.11138/ orl/2016.9.1.001
- 12. Rocci A, Calcaterra R, Rocci M, Rocci C, Baggi L. The influence of micro and macro-geometry in term of bone-implant interface in two implant systems: an histomorphometrical study. *PubMed*. 2017;8(4):87-95. doi:https://doi.org/10.11138/orl/2015.8.4.087
- 13. Rocci A, Calcaterra R, Rocci M, Rocci C, Michele Di Girolamo, L Baggi. Different performance of platform switching in equicrestal position implant: an histological study. *PubMed*. 2017;9(1):11-16. doi:https://doi.org/10.11138/orl/2016.9.1.011
- Calcaterra R, Girolamo M, Mirisola C, Baggi L. Effects of Repeated Screw Tightening on Implant Abutment Interfaces in Terms of Bacterial and Yeast Leakage in Vitro: One-Time Abutment Versus the Multiscrewing Technique. *The International Journal of Periodontics & Restorative Dentistry*. 2016;36(2):275-280. doi:https://doi.org/10.11607/prd.2082
- Meleo D, Baggi L, Di Girolamo M, Di Carlo F, Pecci R, Bedini R. Fixture-abutment connection surface and micro-gap measurements by 3D micro-tomographic technique analysis. *DOAJ (DOAJ: Directory of Open Access Journals)*. 2012;48(1):53-58. doi:https://doi.org/10.4415/ann_12_01_09
- Degidi M, Nardi D, Piattelli A. One abutment at one time: non-removal of an immediate abutment and its effect on bone healing around subcrestal tapered implants. *Clinical Oral Implants Research*. 2011;22(11):1303-1307. doi:https://doi.org/10.1111/j.1600-0501.2010.02111.x
- 17. Hansson S. A conical implant-abutment interface at the level of the marginal bone improves the distribution of stresses in the supporting bone. *Clinical Oral Implants Research*. 2003;14(3):286-293. doi:https://doi.org/10.1034/j.1600-0501.2003.140306.x



Case report

SURGICAL APPROACH OF AN ECTOPIC THIRD MOLAR IN THE MAXILLARY SINUS

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ABSTRACT

Ectopia of third molars within the maxillary sinus is uncommon. Few cases have been reported in the literature. Generally, the diagnosis of upper third molar ectopia at the level of the maxillary sinus can be made following a routine diagnostic examination such as panoramic X-ray, or CBCT in which any lesions created by the element itself can additionally be detected. Our case presents a third molar included in the left upper maxilla of a 60-year-old male patient. The element was removed under general anesthesia, and after twelve months of follow-up, new panoramic X-ray and CBCT were requested to assess the healing of the compromised area.

KEYWORDS: ectopia, molar, maxillary sinus

INTRODUCTION

Dental eruption is a physiological process by which the tooth element in formation achieves its functional position within the oral cavity. Development of the element and proper intraoral positioning depend on complex cellular interactions and molecular processes that may be subject to variation determining the development of an ectopic tooth.

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Ectopic eruption of a tooth element is frequently encountered in clinical practice and the etiology is multifactorial: cleft palate, trauma, odontogenic or rhinogenic infections, genetic factors, cysts, and dental crowding can all contribute to the onset of the phenomenon (1).

This altered process is frequently seen in dental areas, but less common in non-toothed areas such as the mandibular condyle, coronary process, orbit, palate, and nasal cavity. Occasionally, a tooth may erupt within the maxillary sinus.

In the English literature, patients were observed with higher prevalence of ectopic teeth in third molars, 21 cases, followed by unspecified molars. The lowest prevalence of ectopic teeth was found in the first molar, second premolar, and first incisor (2-6).

Generally these elements remain asymptomatic for years and their diagnosis is made only after routine diagnostic exams are performed; sometimes they may cause recurrent sinusitis.

Case report

A male patient aged 60 years came to our observation, reporting pain in the left upper maxillary area, retronasal purulent discharge and halitosis for about three

months.

On intraoral clinical examination, there was mild swelling in the left upper vestibule at the level of the molar apices. The area was painful on palpation with discharge of purulent material at the intrasulcular level of elements 26 and 27 and from the left nasal choana.

A panoramic X-ray was done and it showed a radiotransparent area involving the region of the left upper maxilla and the upper third molar within the maxillary sinus (Fig. 1).

A chronic purulent sinusitis associated with a maxillary odontogenic cyst from 28 in ectopic position was suspected. Then a CBCT scan has been prescribed, showing a hypodense and wellcircumscribed lesion measuring 20 mm x 30 mm in the posterior region of the maxilla, surrounding the crown of the left third molar in an ectopic position. The right maxillary sinus showed mucosal thickening and filling of the alveolar recess, indicative of chronic maxillary sinusitis (Fig. 2).

The patient was admitted to the hospital for surgery under general anesthesia. Before surgery, informed consent was signed by the patient. Under general anesthesia, we proceeded to do Caldwell-Luc surgery with removal of the cyst and removal of the associated tooth and extracted the severely compromised elements 26 and 27. A thick purulent creamy material, due to the



Fig. 1. *Panoramic X-ray showing the presence of ectopic tooth element 2.8 at the level of the left maxillary sinus.*



Fig. 2. From the CBCT, a hypodense and well-circumscribed lesion surrounds the crown of the third molar.

infectious process, was found within the cavity. The surgery was accompanied by a counter-opening performed in the medial sinus wall, introduction of PVC canula for drainage and endosinus lavage with physiologic saline. The canula was removed on day 5. Periodic nasal sinus lavages with physiologic were performed every 10 days for a period of 2 months.

The specimen was sent for histological examination, which confirmed the diagnosis of odontogenic cyst. In the postoperative course there were no severe complications. The final diagnosis of maxillary sinusitis caused by dentigerous cyst with ectopic third molar was confirmed. Severe and disabling symptomatology disappeared after surgery.

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After 14 days, sutures have been removed. The healing was found to be good, and the mucosa appeared healthy and pink. On an objective examination performed 30 days after surgery, the patient was asymptomatic and optimal tissue healing was evident. Approximately 12 months after surgery, a panoramic X-ray and a CBCT were prescribed to the patient to assess the healing and bone regeneration of the cavity (Fig 3, 4).

DISCUSSION

Dental tissue development begins in the intrauterine phase, during the sixth week, through interaction between oral epithelium and mesenchymal tissue. Abnormal tissue interactions, embryological pathologies, such as fusion defects or cyst formations, during this stage, can generate dental ectopias. In addition, the same phenomenon could be caused by displacement of the dental gems, by expanding dentigerous cysts or displacement during eruption of the third molar, malpositioning related to trauma and supernumerary teeth (2).

Certainly, the ectopic condition in an area that physiologically does not involve the presence of dental elements, such as the maxillary sinus, is not frequently encountered in clinical practice. The elements most susceptible to ectopia seem to be the third molars and canines, which generally take a longer time to erupt.

The discovery of a tooth element in ectopic position may or may not be accompanied by the presence of symptoms such as sinusitis and purulent rhinorrhoea, that cannot be treated with antibiotic prophylaxis. In addition, patients often report swelling, pain, headache, and nasal obstruction. In some cases, symptoms like



Fig. 3, 4. X-rays demonstrating successful bone healing.

infraorbital nerve hypoesthesia, epiphora and hemoptysis have been described. Regarding possible infections, cases of oroantral fistulas and purulent discharge have been reported (6-9). Our patient reported pain in the left upper maxillary area, retronasal purulent discharge and halitosis for about three months.

The diagnosis of the lesion associated with ectopic element was made following a panoramic X-ray. Subsequently, the patient underwent a CBCT to better highlight the location of the tooth and the margins of the hypodense lesion identified in the previous X-ray. The CBCT gives a better representation of the sinuses; it also allows us to have more details regarding the position, in this case, of the ectopic element and the size and extension of the associated lesion (10). After viewing the CBCT we opted for enucleation of the cyst removing the associated element with Cadwell-Luc surgery, by which the operators ensured a direct view of the element during the surgical procedure.

CONCLUSIONS

In conclusion, dentigerous cysts associated with ectopic maxillary third molars are rare and poorly documented. They can involve the maxillary sinus and cause chronic maxillary sinusitis (11, 12). Treatment involves surgical removal of the tooth and of the associated lesion using the Cadwell-Luc procedure. If there is unilateral maxillary pain, or hemifacial headache, odontogenic sinusitis should be suspected. A careful oral and radiographic examination is essential as evidenced by the case reported in this article, in which a dentigerous cyst associated with an ectopic third molar caused a maxillary odontogenic sinusitis.

REFERENCES

- 1. Capelli M, Lombroni L, Farronato G, Santamaria G, Lombroni D, Gatti P. Ectopic teeth in the maxillary sinus: A case report and literature review. *Indian Journal of Dental Research*. 2018;29(5):667. doi:https://doi.org/10.4103/ijdr.ijdr_347_17
- 2. Cohen C. Diagnosis and Treatment of Ectopic Eruption of Permanent Molars. Oral Health Group. 2013;1(3).
- 3. Shandilya Ramanojam, Rajshekhar Halli, Manjula Hebbale, Smita Singh Bhardwaj. Ectopic tooth in maxillary sinus: Case series. *Annals of maxillofacial surgery*. 2013;3(1):89-89. doi:https://doi.org/10.4103/2231-0746.110075
- 4. James O, Suleiman IK, Ahmad MM, Olasoji HO. Management of rare ectopic teeth eruption: case series. *Journal of the Korean Association of Oral and Maxillofacial Surgeons*. 2023;49(2):86-90. doi:https://doi.org/10.5125/jkaoms.2023.49.2.86
- Elif Soğur, Meltem Ozden, Tayfun Günbay, Zuhal Tuğsel. Cone beam computed tomography findings of ectopic tooth in the maxillary sinus associated with dentigerous cyst: A report of two cases and review of the literature. *Journal of Oral and Maxillofacial Radiology*. 2015;3(2):70-70. doi:https://doi.org/10.4103/2321-3841.157529
- 6. Guruprasad Y, Chauhan DS, Kura U. Infected Dentigerous Cyst of Maxillary Sinus Arising from an Ectopic Third Molar. *Journal of Clinical Imaging Science*. 2013;3(7):7. doi:https://doi.org/10.4103/2156-7514.117461
- 7. Srinivasa Prasad T, Sujatha G, Niazi T, Rajesh P. Dentigerous cyst associated with an ectopic third molar in the maxillary sinus: A rare entity. *Indian Journal of Dental Research*. 2007;18(3):141. doi:https://doi.org/10.4103/0970-9290.33793
- 8. Saleem T, Khalid U, Hameed A, Ghaffar S. Supernumerary, ectopic tooth in the maxillary antrum presenting with recurrent haemoptysis. *Head & Face Medicine*. 2010;6(1). doi:https://doi.org/10.1186/1746-160x-6-26
- 9. Büyükkurt MC, Tozoglu S, Aras MH, Yolcu Ü. Ectopic Eruption of a Maxillary Third Molar Tooth in the Maxillary Sinus: A Case Report. *The Journal of Contemporary Dental Practice*. 2005;6(3):104-110. doi:https://doi.org/10.5005/jcdp-6-3-104
- 10. Han MH, Chang KH, Lee CH, Na DG, Yeon KM, Han MC. Cystic expansile masses of the maxilla: differential diagnosis with CT and MR. *AJNR American journal of neuroradiology*. 1995;16(2):333-338.
- 11. Candotto V, Gallusi G, Piva A, Baldoni M, Di Girolamo M. Complications in sinus lift. *Journal of Biological Regulators and Homeostatic Agents*. 2020;34(1 Suppl. 1):139-142. DENTAL SUPPLEMENT.
- 12. Di Girolamo S, Martino F, Guerrieri M, et al. Odontogenic Maxillary Sinusopathies: a Radiological Classification. *Journal of Maxillofacial and Oral Surgery*. 2020;21(1). doi:https://doi.org/10.1007/s12663-020-01329-8



Case report

TREATMENT MANAGEMENT IN A YOUNG PATIENT WITH TEMPOROMANDIBULAR DISORDER AND MALOCCLUSION: A CASE REPORT

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ABSTRACT

Signs and symptoms of temporomandibular disorders (TMD) are observed in a percentage ranging from 7.3 to 30.4% of children and adolescents. The purpose of this work is to report a clinical case of a young patient suffering from TMD and malocclusion and who was treated with a gnathological occlusal splint and fixed orthodontic appliance. The patient, a girl aged 10 years and 10 months, had a slight tendency to skeletal Class III malocclusion, 6 mm overbite, 1.4-1.5 crossbite, multiple rotations and lower crowding. A gnathological occlusal splint was made to alleviate the acute symptoms and a gnathological retention splint at the finishing stage of fixed appliance was applied to achieve the functional occlusion. Observation after 2 years out of orthodontic treatment revealed a stable occlusion and improved of TMD symptoms.

KEYWORDS: temporomandibular disorder, malocclusion, treatment, management, adolescent patients

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INTRODUCTION

In everyday clinical practice, it is common to find patients reporting orofacial pain. Thus, a routine temporomandibular disorder (TMD) examination prior to the beginning of the orthodontic therapy is essential (1). For orthodontists and general dentists, it is mandatory to carry out a complete medical history and a comprehensive temporomandibular joint (TMJ) exam to evaluate the presence of any TMD. This allows to recognize patients suffering from orofacial pain conditions and thus to exclude them from the orthodontic treatment until the pain sensation is managed (2).

TMD is a collective term, including several clinical problems involving muscles, TMJ or both. Even if TMD more frequently affects adults, signs and symptoms are observed in a percentage ranging from 7.3 to 30.4% of children and adolescents. The prevalence is higher in females than males and increased with pubertal development (3–5).

The TMD etiopathogenesis of growing patients includes systemic, pathological, psychosocial traumatic, hormonal, genetical, skeletal and occlusal factors (3, 6). The diagnosis of TMD is based on anamnestic collection, clinical examination and instrumental diagnosis.

Clinical and physical assessment of the patient may include the history and determination of joint sounds, evaluation of the mandibular range of motion, appraisal of pain, evaluation for signs of inflammation and a correct clinical and radiographic examination (7, 8). For clinical diagnosis, the Diagnostic Criteria for Temporomandibular Disorders (DC/TMD) use, by Schiffman et al (2014), are strongly recommended (9).

TMD treatment goals include restoration of function, pain decrease, control of any aggravating or contributing factors and improvement of life quality.

Treatment of TMD can be divided into reversible and irreversible. It has been suggested that simple, conservative and reversible types of therapy are effective in reducing most TMD symptoms in children (10–13), including patient education, physical therapy [e.g., jaw exercises or transcutaneous electrical nerve stimulation [TENS)], behavioral therapy, prescription medication (e.g., non-steroidal anti-inflammatory drugs and muscle relaxers) and occlusal splints (3).

The goal of an occlusal appliance is to provide orthopedic stability to the TMJ. These may be used to decrease parafunctional activity and pain (14–16). Occlusal splints are made of hard acrylic. The stabilization type of splint covers all teeth on either the maxillary or mandibular arch and is balanced to allow the occlusion of all teeth when the jaw is in a musculoskeletal stable position.

The aim of an occlusal appliance is to provide orthopedic stability to the TMJ before starting an orthodontic treatment in permanent dentition. These alter the patient's occlusion temporarily and may be used to decrease the parafunctional activity and pain.

Every comprehensive dental history and examination should include a TMJ history and assessment (17). The history should include questions concerning the presence of head and neck pain and mandibular dysfunction, previous orofacial trauma and current illness with an account of the symptoms. In the presence of a positive history and/or signs and symptoms of TMD, a more comprehensive examination (e.g., palpation of masticatory and associated muscles and the TMJ's, documentation of joint sounds, occlusal analysis, and assessment of range of mandibular movements including maximum opening, protrusion, and lateral excursions) (18), together with general dental and medical assessments (19, 20) should be performed.

Thus, the purpose of this work is to report a clinical case of a young patient accompanied with temporomandibular, treated with gnathological occlusal splint, fixed orthodontic appliance and transcutaneous electrical nerve stimulation (TENS).

CLINICAL CASE

Diagnosis and etiology

The patient came with their parents to the Orthodontic Program of the Multidisciplinary Department of Medical-Surgical and Dental Specialties of the University of Campania Luigi Vanvitelli in Naples. They were worried about her orofacial pain and temporal headache and were unsatisfied with her smile.

The patient was a 10 years and 10 months old Caucasian girl who was particularly anxious. A full visit with occlusal and functional examinations was performed by an orthodontist. The clinical evaluation revealed mouth opening reduced

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(36 mm), normal lateral mandibular movements (9 mm), joint pain during functions and no clicking sounds in the TMJ. About the symptoms, she referred daily temporal headache, weakness upon weaking, myofascial pain, difficulty in mouth opening, parafunctional activities and anxiety.

Moreover, the patient presented a slight tendency to class III skeletal malocclusion, increased lower third of the face, irregular smile arch, crossbite of 1.4-1.5, increased overbite (6 mm), moderate lower crowding and multiple rotations. She did not receive any previous gnathological or orthodontic treatment.

In frontal view, the patient presented a symmetric face while in lateral view, the profile was retruded with incompetent lips at rest. There was no history of trauma of craniofacial complex. The panoramic radiograph showed the presence of all permanent teeth and the symmetric condyles without any pathological alterations (Fig. 1).

The cephalometric morphological assessment of the lateral skull radiograph showed a slight tendency of skeletal Class III (ANB = 1° ; AoBo = -1 mm) with hyperdivergency (S-N / Go-Gn = 39° ; FMA = 30°) and the lower incisors presented lingual inclination (IMPA = 82°).

Treatment objectives

Based on the patient's age and diagnosis, the best treatment option was an initial gnathological phase followed by orthodontic treatment.

The treatment of choice seemed to be the most rational option considering the patient and her parents' anxiety and expectations, the TMD and the occlusal features showing unilateral crossbite and lower crowding. The main treatment objectives are described below.

First phase: gnathological treatment to reduce muscle contraction, parafunctional activity and pain. The appliances used were gnathological occlusal splint, TENS and physical therapy.

Second phase: orthodontic treatment with crossbite correction, vertical growth pattern control, lower crowding correction, alignment, leveling and arch form coordination and overbite correction. The appliances used were a 7-7 multibracket fixed appliance in the upper and lower arch (0.022x0.028 MBT prescription) and cusp seating elastics. Thus, a two-phase gnathological-orthodontic treatment was proposed and accepted by the patient and her parents.

Treatment progress

The first phase of treatment started one month later the initial check-up, in October 2016. The first gnathological phase



Fig. 1. Pre-treatment records

consisted of a treatment with occlusal splint in the upper arch (functionalized one a month), TENS (two a month) and physical therapy (two a month) for 8 months (Fig. 2).

After this period of time, the patient underwent a re-evaluation with interim records (both photographic and radiographic) in July 2017. During the re-evaluation was found an improvement of temporal headache, weakness upon weaking, myofascial pain, difficulty in mouth opening, parafunctional activities and anxiety. Then the patient was ready to proceed towards the second phase, controlling constantly any recurrence of temporo-mandibular symptoms.

The second phase of treatment started in September 2017. The upper and lower arches were fully bonded with 0.022x0.025" MBT multibracket fixed appliances.

The following archwire sequencing was used: .016 nickel-

titanium for alignment, .019x.025 nickel-titanium for leveling, .019x.025 stainless- steel for arch coordination and .018 AJ Wilcock Australian wire with refinement bends and cusp seating elastics for the finishing stage. This phase of treatment lasted 1 years and 6 months.

The retention phase included a Hawley retainer appliance in the upper arch and a cuspid-to- cuspid fixed retainer in the lower arch. The patient was repeatedly advised to report any case of recurrence of orofacial pain during the orthodontic phase.

Treatment results

The treatment goals were achieved (Fig. 3). The occlusal, functional and esthetic results were satisfactory, the patient and her parents were happy of her smile. TMD symptoms were improved, the smile arch was good with no buccal corridors, however the profile appears still biretruded.

Oral hygiene during orthodontic treatment was quite good, periodontal tissues were healthy. There were no decayed elements or signs of enamel decalcification and the panoramic radiograph did not show any sign of bone loss or root resorption.



Fig. 3. Final records





Fig. 2. Gnathological occlusal splint and TENS

Intra-oral photographs and dental casts showed a good alignment of marginal ridges, while leveling and arch coordination were achieved: the crossbite and the lower crowding were corrected. The overjet was maintained and the overbite was corrected.

The final static occlusion was satisfactory also on the lingual side and no prematurity was present during protrusive and lateral mandibular movements. Panoramic radiograph revealed that good roots angulation was achieved.

The lateral skull radiograph showed the control of vertical skeletal relations between pre-treatment and post-treatment cephalograms and the incisor inclination.

The panoramic radiograph showed no signs of condylar resorption or periodontal disease. The third molars were present and impacted within the jaw bones.

No clear signs of root resorption can be noted. Root angulations were parallel.

In the final lateral cephalogram assessment the hyperdivergent pattern was controlled (S-N/Go-Gn from 39° to 38°).

The upper and lower incisors inclinations were improved (I/SN from 98° to 102°; IMPA from 82° to 92°). Overjet has remained relatively unchanged while the overbite was corrected.

DISCUSSION

The reported case of a child with TMD pain shows how treatment goals were achieved. The symptomatic, functional, occlusal, esthetic and psychological results were satisfactory. The outcome was rewarding for the clinicians and appreciated by the patient and her parents The key points determining the success of the treatment were good interdisciplinary cooperation (orthodontist, physiotherapist and mental health specialist) and the parent's and patient's collaboration, as reported previously in the international literature (21, 22). Further possible medical correlations should be always checked and monitored in these patients. It is important to prioritize the patient's symptoms, evaluating not only the occlusion but the entire orofacial area, trying not to minimize any signs of TMD. In the field of clinical dentistry, TMD are one of the major diseases. TMD pain in adolescents' patients is frequent and has a clear impact on daily living. In the present study we have used a splint in the upper arch (functionalized one a month), and TENS. Splint therapy has been used to help a majority of young patient with TMD pain and was also a treatment approach in this study (23). TENS is a non-invasive treatment modality for acute and chronic pain. TENS has a positive effect in treatment of TMD patient.

CONCLUSIONS

Treatment of adolescent patients with combined TMD and severe dento-skeletal malocclusions is among the most difficult challenges for orthodontists. In fact, the orthodontists cannot simply aim an occlusal correction but also have to treat all the orofacial complex trying to keep TMD under control, trying to prioritize TMJ signs and symptoms improvement.

Therefore, the treatment in a patient with TMD and malocclusion should be an interdisciplinary treatment that aimed to improve the function, the occlusion and consequently the patient's quality of life. The present study shows also the immediate effects of TENS treatment on TMD-related muscle pain.

REFERENCES

- Festa F, Rotelli C, Scarano A, Navarra R, Caulo M, Macri M. Functional Magnetic Resonance Connectivity in Patients with Temporomadibular Joint Disorders. *Frontiers in Neurology*. 2021;12:629211. doi:10.3389/fneur.2021.629211
- Michelotti A, Rongo R, D'Antò V, Bucci R. Occlusion, orthodontics, and temporomandibular disorders: Cutting edge of the current evidence. *Journal of the World Federation of Orthodontists*. 2020;9(3):S15-S18. doi:10.1016/j.ejwf.2020.08.003
- 3. American Academy of Pediatric Dentistry. Acquired Temporomandibular Disorders in Infants, Children, and Adolescents. *Pediatric Dentistry*. 2017;39(6):354-360.
- 4. Song YL, Yap AU, Türp JC. Association between temporomandibular disorders and pubertal development: A systematic review. *Journal of Oral Rehabilitation*. 2018;45(12):1007-1015. doi:10.1111/joor.12704

- Christidis N, Lindström Ndanshau E, Sandberg A, Tsilingaridis G. Prevalence and treatment strategies regarding temporomandibular disorders in children and adolescents-A systematic review. *Journal of Oral Rehabilitation*. 2019;46(3):291-301. doi:10.1111/joor.12759
- 6. Atsü SS, Güner S, Palulu N, Bulut AC, Kürkçüoğlu I. Oral parafunctions, personality traits, anxiety and their association with signs and symptoms of temporomandibular disorders in the adolescents. *African Health Sciences*. 2019;19(1):1801. doi:10.4314/ahs.v19i1.57
- Inchingolo AD, Ferrara I, Viapiano F, et al. Rapid Maxillary Expansion on the Adolescent Patient: Systematic Review and Case Report. *Children*. 2022;9(7):1046. doi:10.3390/children9071046
- Scarano A, Inchingolo F, Rapone B, Festa F, Rexhep Tari S, Lorusso F. Protective Face Masks: Effect on the Oxygenation and Heart Rate Status of Oral Surgeons during Surgery. *International Journal of Environmental Research and Public Health*. 2021;18(5):2363. doi:10.3390/ijerph18052363
- Schiffman E, Ohrbach R, Truelove E, et al. Diagnostic Criteria for Temporomandibular Disorders (DC/TMD) for Clinical and Research Applications: Recommendations of the International RDC/TMD Consortium Network* and Orofacial Pain Special Interest Group†. *Journal of Oral & Facial Pain and Headache*. 2014;28(1):6-27. doi:10.11607/jop.1151
- 10. Rongo R, Ekberg E, Nilsson I, et al. Diagnostic criteria for temporomandibular disorders (DC/TMD) for children and adolescents: An international Delphi study—Part 1-Development of Axis I. *Journal of Oral Rehabilitation*. 2021;48(7):836-845. doi:10.1111/joor.13175
- Rongo R, Ekberg E, Nilsson I, et al. Diagnostic criteria for temporomandibular disorders in children and adolescents: An international Delphi study-Part 2-Development of Axis II. *Journal of Oral Rehabilitation*. 2022;49(5):541-552. doi:10.1111/ joor.13301
- 12. Scrivani SJ, Khawaja SN, Bavia PF. Nonsurgical Management of Pediatric Temporomandibular Joint Dysfunction. Oral and Maxillofacial Surgery Clinics of North America. 2018;30(1):35-45. doi:10.1016/j.coms.2017.08.001
- Bodner L, Miller VJ. Temporomandibular joint dysfunction in children: evaluation of treatment. *International Journal of Pediatric Otorhinolaryngology*. 1998;44(2):133-137. doi:10.1016/s0165-5876(98)00055-x
- Wahlund K, List T, Larsson B. Treatment of temporomandibular disorders among adolescents: a comparison between occlusal appliance, relaxation training, and brief information. *Acta Odontologica Scandinavica*. 2003;61(4):203-211. doi:10.1080/00016350310003891
- 15. Wahlund K, Larsson B. Long-term treatment outcome for adolescents with temporomandibular pain. *Acta Odontologica Scandinavica*. 2017;76(3):153-160. doi:10.1080/00016357.2017.1394490
- Simmons III HC, Gibbs SJ. Anterior Repositioning Appliance Therapy for TMJ Disorders: Specific Symptoms Relieved and Relationship to Disk Status on MRI. *CRANIO* (2005;23(2):89-99. doi:10.1179/crn.2005.014
- 17. Fujii T, Torisu T, Nakamura S. A Change of Occlusal Conditions After Splint Therapy for Bruxers With and Without Pain in the Masticatory Muscles. *CRANIO* [®]. 2005;23(2):113-118. doi:10.1179/crn.2005.016
- 18. American Academy of Pediatric Dentistry. Best Practices: Record-keeping. The Reference Manual of Pediatric Dentistry. *American Academy of Pediatric Dentistry*. Published online 2021:484-491.
- Scarano A, Mortellaro C, Brucoli M, Lucchina AG, Assenza B, Lorusso F. Short Implants: Analysis of 69 Implants Loaded in Mandible Compared with Longer Implants. *Journal of Craniofacial Surgery*. 2018;29(8):2272-2276. doi:10.1097/ scs.000000000004518
- Scarano A, Murmura G, Mastrangelo F, Lorusso F, Greco Lucchina A, Carinci F. A novel technique to prevent sinus membrane collapse during maxillary sinus floor augmentation without bone graft: technical note. *Journal of Biological Regulators and Homeostatic Agents*. 2018;32(6):1589-1592.
- Moccia S, Nucci L, Spagnuolo C, d'Apuzzo F, Piancino MG, Minervini G. Polyphenols as Potential Agents in the Management of Temporomandibular Disorders. *Applied Sciences*. 2020;10(15):5305. doi:10.3390/app10155305
- 22. Inchingolo F, Tatullo M, Marrelli M, et al. Combined occlusal and pharmacological therapy in the treatment of temporomandibular disorders. *Eurpean Review for Medical and Pharmacological Sciences*. 2011;15(11):1296-1300.
- 23. Nilsson IM, Willman A. Treatment Seeking and Self-Constructed Explanations of Pain and Pain Management Strategies Among Adolescents with Temporomandibular Disorder Pain. *Journal of Oral & Facial Pain and Headache*. 2016;30:127-133. doi:10.11607/ofph.1450





Letter to the Editor

INTERVENTIONAL TREATMENT OF SACROILIAC JOINT DISEASE

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ABSTRACT

Sacroiliac joint (SIJ) disease is a common cause of lower back and buttock pain. It poses a significant diagnostic and therapeutic challenge due to its complex anatomy and varied clinical presentation. Interventional treatments have emerged as effective options for managing SIJ disease, offering potential pain relief and improved quality of life for patients. This comprehensive review explores the interventional treatment modalities available for sacroiliac joint disease, including diagnostic techniques, minimally invasive procedures, and emerging therapies. We delve into the evidence-based literature, discuss the efficacy and safety profiles of these interventions, and highlight key considerations for their implementation. By examining the interventional armamentarium for SIJ disease, this review aims to provide clinicians and patients with a thorough understanding of the available options and inform decision-making in the management of this challenging condition.

KEYWORDS: sacroiliac joint, fixation, injection

INTRODUCTION

The sacroiliac joint (SIJ) plays a crucial role in load transfer and stability of the pelvis, linking the spine to the lower

Received: 14 August 2023 Accepted: 13 September 2023 ISSN: 2038-4106 Copyright © by BIOLIFE 2023 This publication and/or article is for individual use only and may not be further reproduced without written permission from the copyright holder. Unauthorized reproduction may result in financial and other penalties. **Disclosure: All authors report no conflicts of interest relevant to this article.** extremities; while the SIJ disease refers to a range of pathologies including inflammation, degeneration, and instability the SI pain refers to discomfort or pain in the sacroiliac joint (1). SI pain can manifest as pain, tenderness, or discomfort in the lower back, buttocks, hips, or groin area that may be exacerbated by sitting, standing, walking, or climbing stairs (2).

The pain may be localized to one side or can radiate down the leg, resembling sciatica leading to difficulties in differential diagnosis (1-2). The exact prevalence of SI joint-related LBP is challenging to determine due to diagnostic difficulties and varying definitions of SI joint dysfunction. However, studies suggest that the SI joint is a potential source of LBP in approximately 15% to 30% of individuals with chronic low back pain without significant gender difference (3-4).

Despite its prevalence, diagnosis and treatment of SIJ disease remain challenging due to its complex anatomy and the lack of specific clinical and radiographic findings. The advent of interventional techniques has revolutionized the management of SIJ disease, providing targeted therapies and enhancing patient outcomes.

Diagnosis of sacroiliac pain

Diagnosing sacroiliac (SI) pain can be challenging because the symptoms may overlap with other conditions affecting the lower back and hips. The diagnosis of sacroiliac (SI) pain typically involves a comprehensive evaluation that includes a combination of medical history, physical examination including and diagnostic tests; once the diagnosis is confirmed, long-term solutions may be considered.

- 1. Medical History: several factors can increase the risk of developing SI pain, including:
- pregnancy and childbirth: The hormonal changes and increased stress on the SI joints during pregnancy can contribute to SI pain. It is estimated that up to 60% of pregnant women may experience SI joint pain (5);
- trauma or injury: accidents, falls, or repetitive activities that strain the SI joint can lead to SI pain (2). SIJ disease is present in 45%-75% patients undergone posterior fixation treatments (fig. 1a, b);



Fig. 1. Chronic right-side pain and sacro-ileitis in a patient undergone posterior fixation. Coronal CT 2D recon demonstrates transpeduncolare screws at the level of L4, L5 and S1, in a patient treated 5 years before with surgical posterior fixation. No significant bone abnormality can be detected on CT scan (**1a**). On SPECT-CT scan, evident Tc99 uptake can be detected at the level of right SIJ area as well as right iliac bone, secondary to posterior fixation (**1b**).

- inflammatory conditions: certain inflammatory diseases, such as ankylosing spondylitis and psoriatic arthritis, can affect the SI joints and lead to pain (6);
- degenerative conditions: conditions like osteoarthritis or degenerative joint disease can affect the SI joints and cause pain (7). Transitional lumbar vertebra is another condition commonly associated to the SIJ disease (Fig. 2a, b).
- 2. Physical examination: clinical evaluation involves (2):
- posture assessment;
- range of motion in order to assess the mobility and stability of the SI joint using maneuvers such as the FABER (flexion, abduction, external rotation) test, Gaenslen's test, and the thigh thrust test aid in identifying SIJ pathology;
- provocative tests able to reproduce SI joint pain stressing the SI joint in various positions to determine if it is the source of pain.
- Imaging tests: no specific radiological findings for the diagnosis of sacroiliac joint-related pain however diagnostic imaging tests are often used to help confirm the diagnosis and rule out other possible causes of pain. These may include:
- plain films: X-rays can provide a basic view of the SI joint and can help identify fractures, degenerative changes, or abnormalities in the joint structure;
- It is important to remember that the SI joint has a complex three-dimensional structure, and plain film X-rays provide a two-dimensional representation. This limitation can make it challenging to accurately assess the joint's full extent, especially regarding subtle changes or early-stage pathology. Neverless, radiographic features such as erosions, sclerosis, and ankylosis are typically seen in advanced inflammatory sacroiliitis and are graded from 0 (normal) to 4 (ankylosis) according to the modified New York criteria (8).
- Magnetic Resonance Imaging (MRI): an MRI scan can provide more detailed images of the SI joint, soft tissues, and surrounding structures. It can help detect inflammation, joint abnormalities, or other potential causes of pain and it has been introduced for the evaluation of axial spondylarthritis due to contrast resolution and 3D according to sacrum plane (9).
- Computed Tomography (CT) scan: a CT scan may be ordered to provide a detailed, cross-sectional view of the SI joint and surrounding structures, particularly if there is a suspected bony abnormality.



Fig. 2. *Right L5 emisacralization in a patient with transitional vertebra and bilateral sacroiliac pain. On 3D CT recons there's evident fusion between the right L5 hemivertebra and the ipsilateral sacral wing, concurring to the asymmetrical loadstress and SIJ disease development (2a). Bilateral SIJ fixation putting 2 screws on regular left side, one at the S1 level and the second at S2, and a third contralateral screw at S1 level were introduced, resolving the clinical symptoms related to the disease (2b).*

- CT Sensitivity, accuracy and detailed information compared to plain radiography. However, due to higher radiation exposure, it is not advisable to use CT for diagnosis or follow-up purposes.
- nuclear medicine is not typically used as a first-line imaging modality for evaluating SI pain, it can be considered in certain cases to assess specific underlying conditions.

No comprehensive guidelines for SI pain have been provided yet. Routinely, conventional radiography represents the first-line modality in most instances and serves as a useful baseline for future comparison; however, the absence of radiographic changes does not exclude an underlying process and many patients with suspected inflammatory back pain usually proceed to further imaging, in particular MRI (9-10). In patients with suspected infection, contrast-enhanced MRI (CE-MRI) or planar or SPECT-CT isotope bone scintigraphy are the modalities of choice, with MRI offering better assessment of anatomical changes and periarticular soft tissue structures over SPECT-CT without ionizing radiation exposure (9-10).

MRI, CT, and isotope bone scintigraphy are all useful in the detection of stress fractures of the sacrum and pelvis. CT is helpful in situations in which there is a contraindication to MRI and provides excellent delineation of periarticular erosions, sclerosis, or osseous metastasis (9-10).

4. Diagnostic Injections: diagnostic injections, such as intra-articular anesthetic blocks or provocative SIJ injections, are considered the gold standard for confirming the diagnosis of SIJ-related pain; in fact, controlled injections into the SIJ can provide temporary pain relief, aiding in the accurate identification of the pain source.

These injections involve injecting an anesthetic (eg lidocaine) or a combination of anesthetic and anti-inflammatory medication into the SI joint in order to temporarily numb the joint and assess its involvement in the patient's pain symptoms: if the injection provides temporary relief of pain, it suggests that the SI joint is the source of the pain (11). The diagnostic injections are performed under imaging (fluoroscopy and/or CT rarely under US or MRI) guidance in order to drive accurately the needle at the level of SI joint with patient in prone position; usually a local anesthesia is performed before needle insertion at the level of SI joints (11).

No more than 2.5 mL of injectate are recommended during an intra-articular diagnostic injection; in fact, extravasation of local anesthetic onto nearby neural structures theoretically compromises the specificity of the diagnostic injection (12-13).

Minimally invasive procedures

There are various nonsurgical treatment options available for sacroiliac joint SI pain, including pain medications such as nonsteroidal anti-inflammatory drugs (NSAIDs), physical therapy (PT), steroid injections into the SIJ, and radiofrequency ablation (RFA) targeting the sacral nerves.

For acute or subacute SI pain, a combination of NSAIDs, icing, and activity modification can be helpful in reducing pain (11). However, it's important to note that NSAIDs do not address the underlying disease process. Moreover, opioids have not been proven to be safe and effective for treating chronic SIJ pain, and their potential for addiction remains a significant public health concern.

The effectiveness of PT for treating chronic SIJ dysfunction and pain has not been demonstrated probably because of a paucity of high-level literature secondary to the great variability in the functional biomechanical deficit in patients with SI pain.

In this scenario, minimally invasive techniques can play a pivot role in SI pain management. Minimally invasive treatments aim to alleviate discomfort and improve functionality in the sacroiliac joint and enhance the overall quality of life for affected individuals. These techniques offer a targeted and minimally invasive alternative to surgical interventions, reducing morbidity, and optimizing resource utilization.

Sacroiliac joint injections

Sacroiliac joint injections involve the injection of local anesthetics, corticosteroids, or a combination of both into the SIJ. These injections aim to provide pain relief, reduce inflammation, and facilitate functional improvement. Various approaches, such as fluoroscopy-guided, CT-guided or ultrasound-guided injections, can be employed to ensure accurate needle placement. There is no high-level evidence supporting the short- or long-term effectiveness of this treatment option.

Since there is no conclusive evidence supporting corticosteroid injections as superior to a placebo, the usefulness of trials using corticosteroid injections as an active control group is uncertain. No improvement in pain or function beyond 1 month with injections in 3 randomized control trials (RCT) evaluating SIJ injection versus radiofrequency (14-16). The cost-effectiveness of sacroiliac joint steroid injections has not been established.

There is a lack of evidence demonstrating long-term pain relief from this procedure, and the benefits of repeated injections have not been confirmed through studies.

Radiofrequency ablation

Radiofrequency ablation (RFA) involves the use of thermal energy to create lesions on the nerves supplying the SIJ, thereby interrupting pain signals. This minimally invasive procedure offers prolonged pain relief and has shown promising outcomes in patients with SIJ pain refractory to conservative management.

The analysis of RF ablation literature is constrained by the inconsistencies in patient selection criteria, the specific nerves chosen for ablation, and the diversity of RF ablation technologies and techniques employed. Four randomized trials, aiming to explain the effectiveness of radiofrequency (RF) ablation compared to sham procedures, have been published. Two studies indicate that RF ablation of the lateral branches of sacral nerve roots can provide temporary relief from SI pain (17-18). A one-year follow-up from one of the cooled RF ablation trials showed a moderate reduction in pain (19). In a smaller trial conducted by Mehta et al. (with a sample size of 30), RF ablation strip lesioning was compared to a sham procedure, resulting in significant improvement in Visual Analog Scale (VAS) and EuroQOL-5D scores at 3 months (20). A more recent study comparing heated RF ablation to a sham procedure demonstrated no significant difference in pain level or patient satisfaction at 1 or 3 months (21).

Additionally, there are three pragmatic RCTs comparing RF ablation to SIJ steroid injection demonstrating better clinical results in RFG groups (14-16). Moreover, SIJ RF ablation randomized against PT, the authors demonstrated no significant differences in pain level or patient satisfaction at 3, 6, 9, or 12 months (22). In the context of the Dutch healthcare system, RF ablation was determined to lack cost-effectiveness from a societal standpoint for patients experiencing chronic pain originating from the sacroiliac joint (23).

Prolotherapy and PRP injection

Prolotherapy involves the injection of biological substances, such as dextrose, into ligamentous tissue is believed to trigger a series of activities, from the influx of granulocytes, macrophages, and fibroblasts to the release of growth factors, finally leading to collagen deposition.

PRP injections utilize the patient's own concentrated platelets to promote tissue regeneration, reduce inflammation, and alleviate pain. PRP therapy has gained popularity as an adjunctive treatment for SIJ disease, particularly in cases of ligamentous laxity and degeneration. There are not RCT nor cost analysis related to those techniques.

A recent case series demonstrated that concentrated dextrose prolotherapy combined with platelet-rich plasma (PRP) injections has been successfully employed to treat lumbo-sacral spine osteoarthritis (OA) in elderly patients who had previously experienced ineffective results with conventional treatment approaches (24).

Minimally invasive fusion techniques

Minimally invasive fusion techniques, such as SIJ fusion using implants or bone grafts, provide long-term stabilization and pain relief for patients with severe SIJ dysfunction. These procedures aim to restore joint stability while minimizing tissue trauma and accelerating recovery:

- SI Joint Fusion with Implants: this technique involves the use of implants or devices designed to stabilize the SIJ. It typically requires a small incision and the insertion of screws, rods, or plates to fuse the joint. The implants help provide stability while the joint heals.
- SI Joint Fusion with Bone Grafting: in this approach, bone graft material is used to promote fusion between the sacrum and ilium. The graft material may be obtained from the patient's own body (autograft) or from a donor (allograft).

Minimally invasive techniques involve small incisions and the use of specialized instruments to prepare the joint and place the bone graft.

 SI Joint Fusion with Percutaneous Screws: percutaneous or minimally invasive screw fixation involves the placement of screws across the SIJ to provide stability and promote fusion. This technique requires small incisions and the use of image guidance to accurately position the screws (Fig. 3).

The lateral approach has been demonstrated that minimally invasive lateral sacroiliac joint fusion (MIS SIJF) generally causes minimal changes in motion or stress at the opposite sacroiliac joint (contralateral SIJ), minimal increase in motion at the L4-L5 or L5-S1 motion segment, and a limited (5%) increase in stress at the hip joint (25-28).

In 2008, SI-BONE, Inc., obtained FDA clearance to market a porous-surfaced transiliac transfixing implant (TTI) for sacroiliac joint fusion (SIJF). Since then, different lateral transiliac transfixing devices have also received FDA clearance for minimally invasive



Fig. 3. Right side SIJ fixation, AP radiographic view: the lower screw inserted at the level of S2 shows fenestration, to facilitate bone integration.

lateral SIJF. The clinical evidence supporting the use of these devices has significantly expanded over the past decade. However, the majority of high-level clinical evidence regarding the safety, effectiveness, durability, and economic benefits of lateral minimally invasive SIJF is primarily derived from the use of the iFuse implant system (29-31).

These studies present compelling evidence supporting the safety and effectiveness of lateral transiliac minimally invasive sacroiliac joint fusion (MIS SIJF) using lateral transfixing devices. The findings consistently show significant improvements in pain levels, functional abilities, and quality of life (QOL). In both randomized trials, patients who underwent SIJF experienced considerably higher levels of pain relief, reduced disability, and improved QOL compared to those who received non-surgical treatment (32-40).

According to the International Society for the Advancement of Spine Surgery, Policy 2020 Update the MIS SIJF is not indicated in the case of (11):

- Less than 6 months of SIJ pain and/or functional impairment.
- Failure to pursue conservative treatment of the SIJ (unless contraindicated).
- Pain not confirmed with a diagnostic SIJ block.
- Presence of other pathology that would substantially prevent the patient from deriving benefit from SIJF.

EMERGING THERAPIES

Peripheral nerve stimulation

Peripheral nerve stimulation (PNS) involves the placement of electrodes near the nerves supplying the SIJ to modulate pain signals.

PNS is believed to provide pain relief by engaging the gate-control theory of pain, as originally described by Melzack and Wall (41). According to this theory, the excitation of inhibitory dorsal horn interneurons occurs through the stimulation of large-diameter, low-threshold, non-nociceptive A β fibers (42). These interneurons play a role in processing and transmitting nociceptive information from A δ and C nerve fibers, effectively inhibiting the transmission of pain signals from the spinal cord to higher centers in the central nervous system (CNS). PNS also acts to reduce central sensitization and hyperalgesia by diminishing excessive peripheral nociceptive activity within the spinal cord. It achieves this by inhibiting wide dynamic range neurons in the dorsal horn and reducing A β fiber-induced activity within the medial lemniscal pathway in the brain. Additionally, animal studies have indicated that the analgesic effects of PNS may involve various pathways, including the serotonergic (5HT2, 5HT3), GABAergic, and glycinergic systems (43).

In a study involving patients with sacroiliac joint pain that did not respond to conservative measures and injection therapy, PNS was implemented, and the patients were followed for up to four years. The study observed significant reductions in average pain scores at one year (measured on the Visual Analog Scale) from 8.8 to 1.6, at two years from 8.8 to 1.9, and at three years from 8.8 to 2.0. By the fourth year, two out of three patients reported satisfaction with the placement of PNS (44). This emerging therapy offers a reversible and adjustable option for pain management, particularly for patients who have failed conventional treatments (40).

Biologic agents and stem cell therapy

Biologic agents, such as anti-inflammatory cytokines, growth factors, or inhibitors of pain mediators, hold promise for the treatment of SIJ disease. These agents target specific pathways involved in inflammation and pain, providing a potential disease-modifying approach.

Among the different biologic agents, adult stem cells, often known as 'medical signaling cells' or 'mesenchymal stem cells' (MSCs), have been extensively studied. MSCs do not express major histocompatibility complex Class II (MHC class II) proteins, which makes them adaptable to various cell types and reduces the risk of treatment rejection. Their remarkable capacity to differentiate into specific cell types plays a crucial role in the healing process by providing the cells necessary for regeneration (45).

Stem cell therapy explores the regenerative potential of stem cells to repair damaged tissues and promote joint healing. Early preclinical and clinical studies have shown encouraging results, suggesting that stem cell therapy may have a role in the future treatment of SIJ disease. While there are a limited number of studies on the utilization of prolotherapy and biologics for treating axial spine pain, further research with stronger evidence is needed to determine the effectiveness of these therapies (45).

Endoscopic radioablation

Recently invented, Endoscopic radioablation seems to demonstrate more effectiveness in comparison to conventional single-needle RF ablation. The procedure consists in introducing two small working cannulas at the level of the lateral border of both S1 and S1 posterior sacral foramina (Fig. 4a), introducing through the cannula an extremely powerful electroknife together with optic fiber, scratching the lateral margin of the sacral foramina from where the SIJ nerve networks projects to the iliac bone (Fig. 4b), disconnecting the SIJ innervation (46-47).



Fig. 4. Endoscopic radioablation of the SIJ. Under CT-guidance, a working cannula is placed at the lateral margin of the posterior first sacral foramen (4a) and a RF probe is then inserted into, emerging at the level of the sacral bone (4b), performing strong ablation of the SIJ nerve network at the emerging area.

CONCLUSIONS

Interventional treatments for SIJ disease aim to alleviate pain, improve functional capacity, and enhance the overall quality of life for affected individuals. These techniques offer a minimally invasive alternative to surgical interventions, reducing morbidity, and optimizing resource utilization. By precisely targeting the source of pain and providing therapeutic interventions, interventional treatments have become integral to the comprehensive management of SIJ disease.

Ongoing research into innovative therapies and technologies, such as targeted drug delivery systems, nanomedicine, and regenerative medicine, holds promise for the future management of SIJ disease. These advancements may offer novel approaches to pain relief, tissue regeneration, and joint stabilization.

Interventional treatments have revolutionized the management of sacroiliac joint disease, providing targeted approaches to pain relief, functional improvement, and joint stabilization. A comprehensive understanding of diagnostic techniques, minimally invasive procedures, emerging therapies, and their efficacy and safety profiles is crucial for informed decision-making and optimizing patient outcomes.

REFERENCES

- 1. Cohen SP, Chen Y, Neufeld NJ. Sacroiliac joint pain: a comprehensive review of epidemiology, diagnosis and treatment. *Expert review of neurotherapeutics*. 2013;13(1):99-116. doi:https://doi.org/10.1586/ern.12.148
- Gartenberg A, Nessim A, Cho W. Sacroiliac joint dysfunction: pathophysiology, diagnosis, and treatment. *European Spine Journal*. 2021;30(10). doi:https://doi.org/10.1007/s00586-021-06927-9
- Sembrano JN, Polly DW. How Often Is Low Back Pain Not Coming From the Back? Spine. 2009;34(1):E27-E32. doi:https://doi. org/10.1097/brs.0b013e31818b8882
- 4. Lingutla KK, Pollock R, Ahuja S. Sacroiliac joint fusion for low back pain: a systematic review and meta-analysis. *European Spine Journal*. 2016;25(6):1924-1931. doi:https://doi.org/10.1007/s00586-016-4490-8
- 5. Fiani B, Sekhon M, Doan T, et al. Sacroiliac Joint and Pelvic Dysfunction Due to Symphysiolysis in Postpartum Women. *Cureus*. 2021;13(10). doi:https://doi.org/10.7759/cureus.18619
- Baronio M, Sadia H, Paolacci S, et al. Etiopathogenesis of sacroiliitis: implications for assessment and management. *The Korean Journal of Pain*. 2020;33(4):294-304. doi:https://doi.org/10.3344/kjp.2020.33.4.294
- O'Shea FD, Boyle E, Salonen DC, et al. Inflammatory and degenerative sacroiliac joint disease in a primary back pain cohort. *Arthritis Care & Research*. 2010;62(4):447-454. doi:https://doi.org/10.1002/acr.20168
- van der Linden S, Valkenburg HA, Cats A. Evaluation of diagnostic criteria for ankylosing spondylitis. A proposal for modification of the New York criteria. *Arthritis and Rheumatism*. 1984;27(4):361-368. doi:https://doi.org/10.1002/art.1780270401
- Mascarenhas V, Sudol-Szopinska I, Boutry N, et al. Imaging and Interpretation of Axial Spondylarthritis: The Radiologist's Perspective—Consensus of the Arthritis Subcommittee of the ESSR. *Seminars in Musculoskeletal Radiology*. 2014;18(03):265-279. doi:https://doi.org/10.1055/s-0034-1375569
- 10. Kok HK, Mumtaz A, O'Brien C, Kane D, Torreggiani WC, Delaney H. Imaging the Patient With Sacroiliac Pain. *Canadian* Association of Radiologists Journal. 2016;67(1):41-51. doi:https://doi.org/10.1016/j.carj.2015.08.001
- Lorio M, Kube R, Araghi A. International Society for the Advancement of Spine Surgery Policy 2020 Update—Minimally Invasive Surgical Sacroiliac Joint Fusion (for Chronic Sacroiliac Joint Pain): Coverage Indications, Limitations, and Medical Necessity. *International Journal of Spine Surgery*. 2020;14(6):860-895. doi:https://doi.org/10.14444/7156
- Kennedy DJ, Engel A, Kreiner DS, Nampiaparampil D, Duszynski B, MacVicar J. Fluoroscopically Guided Diagnostic and Therapeutic Intra-Articular Sacroiliac Joint Injections: A Systematic Review. *Pain Medicine*. 2015;16(8):1500-1518. doi:https://doi.org/10.1111/pme.12833
- Jung MW, Schellhas K, Johnson B. Use of Diagnostic Injections to Evaluate Sacroiliac Joint Pain. International Journal of Spine Surgery. 2020;14(s1):S30-S34. doi:https://doi.org/10.14444/6081

- Salman OH, Gad GS, Mohamed AA, Rafae HH, Abdelfatah AM. Randomized, controlled blind study comparing sacroiliac intraarticular steroid injection to radiofrequency denervation for sacroiliac joint pain. *Egyptian Journal of Anaesthesia*. 2016;32(2):219-225. doi:https://doi.org/10.1016/j.egja.2015.07.005
- 15. Dutta K, Dey S, Bhattacharyya P, Agarwal S, Dev P. Comparison of Efficacy of Lateral Branch Pulsed Radiofrequency Denervation and Intraarticular Depot Methylprednisolone Injection for Sacroiliac Joint Pain. *Pain Physician*. 2018;21(5):489-496.
- 16. Cánovas Martínez L, Orduña Valls J, Paramés Mosquera E, Lamelas Rodríguez L, Rojas Gil S, Domínguez García M. Sacroiliac joint pain: prospective, randomised, experimental and comparative study of thermal radiofrequency with sacroiliac joint block. *Revista Española de Anestesiología y Reanimación*. 2016;63(5):267-272. doi:https://doi.org/10.1016/j.redar.2015.08.003
- Cohen SP, Hurley RW, Buckenmaier CC, Kurihara C, Morlando B, Dragovich A. Randomized Placebo-controlled Study Evaluating Lateral Branch Radiofrequency Denervation for Sacroiliac Joint Pain. *Anesthesiology*. 2008;109(2):279-288. doi:https://doi. org/10.1097/aln.0b013e31817f4c7c
- Patel N, Gross AJ, Lora Beth Brown, Gennady Gekht. A Randomized, Placebo-Controlled Study to Assess the Efficacy of Lateral Branch Neurotomy for Chronic Sacroiliac Joint Pain. Pain Med. 2012;13(3):383-398. doi:https://doi.org/10.1111/j.1526-4637.2012.01328.x
- Patel N. Twelve-Month Follow-Up of a Randomized Trial Assessing Cooled Radiofrequency Denervation as a Treatment for Sacroiliac Region Pain. *Pain Practice*. 2015;16(2):154-167. doi:https://doi.org/10.1111/papr.12269
- Mehta V, Poply K, Husband M, Anwar S, Langford R. The Effects of Radiofrequency Neurotomy Using a Strip-Lesioning Device on Patients with Sacroiliac Joint Pain: Results from a Single-Center, Randomized, Sham-Controlled Trial. *Pain Physician*. 2018;21(6):607-618.
- van Tilburg CWJ, Schuurmans FA, Stronks DL, Groeneweg JG, Huygen FJPM. Randomized Sham-controlled Double-Blind Multicenter Clinical Trial to Ascertain the Effect of Percutaneous Radiofrequency Treatment for Sacroiliac Joint Pain. *The Clinical Journal of Pain*. 2016;32(11):921-926. doi:https://doi.org/10.1097/ajp.00000000000351
- Juch JNS, Maas ET, Ostelo RWJG, et al. Effect of Radiofrequency Denervation on Pain Intensity Among Patients With Chronic Low Back Pain. JAMA. 2017;318(1):68. doi:https://doi.org/10.1001/jama.2017.7918
- 23. Maas ET, Juch JNS, Ostelo RWJG, et al. Cost-Effectiveness of Radiofrequency Denervation for Patients With Chronic Low Back Pain: The MINT Randomized Clinical Trials. *Value in Health*. 2020;23(5):585-594. doi:https://doi.org/10.1016/j.jval.2019.12.009
- Devika Roy GT, Walker V. Ultrasound Guided Dextrose Prolotherapy and Platelet Rich Plasma Therapy in Chronic Low Back Pain: Three Case Reports. *International Journal of Physical Medicine & Rehabilitation*. 2013;01(06). doi:https://doi. org/10.4172/2329-9096.1000149
- 25. Chip Routt ML, Meier MC, Kregor PJ, Mayo KA. Percutaneous iliosacral screws with the patient supine technique. *Operative Techniques in Orthopaedics*. 1993;3(1):35-45. doi:https://doi.org/10.1016/s1048-6666(06)80007-8
- Lindsey DP, Parrish R, Gundanna M, Leasure J, Yerby SA, Kondrashov D. Biomechanics of unilateral and bilateral sacroiliac joint stabilization: laboratory investigation. *Journal of Neurosurgery: Spine*. 2018;28(3):326-332. doi:https://doi. org/10.3171/2017.7.spine17499
- Lindsey DP, Kiapour A, Yerby SA, Goel VK. Sacroiliac Joint Fusion Minimally Affects Adjacent Lumbar Segment Motion: A Finite Element Study. *International Journal of Spine Surgery*. 2015;9:64. doi:https://doi.org/10.14444/2064
- Joukar A, Chande RD, Carpenter RD, et al. Effects on hip stress following sacroiliac joint fixation: A finite element study. JOR SPINE. 2019;2(4). doi:https://doi.org/10.1002/jsp2.1067
- 29. Frank C, Dimitriy Kondrashov, S Craig Meyer, et al. Work intensity in sacroiliac joint fusion and lumbar microdiscectomy. *ClinicoEconomics and Outcomes Research*. 2016;Volume 8:367-376. doi:https://doi.org/10.2147/ceor.s112006
- Garber T, Ledonio CGT, Polly DW. How Much Work Effort is Involved in Minimally Invasive Sacroiliac Joint Fusion? International Journal of Spine Surgery. 2015;9:58. doi:https://doi.org/10.14444/2058
- Lorio M, Martinson M, Ferrara L. Paired Comparison Survey Analyses Utilizing Rasch Methodology of the Relative Difficulty and Estimated Work Relative Value Units of CPT[®]Code 27279. *International Journal of Spine Surgery*. 2016;10:40. doi:https:// doi.org/10.14444/3040

- Sturesson B, Kools D, Pflugmacher R, Gasbarrini A, Prestamburgo D, Dengler J. Six-month outcomes from a randomized controlled trial of minimally invasive SI joint fusion with triangular titanium implants vs conservative management. *European* Spine Journal. 2016;26(3):708-719. doi:https://doi.org/10.1007/s00586-016-4599-9
- 34. Kapural L, Provenzano D, Narouze S. RE: Juch JNS, et al. Effect of Radiofrequency Denervation on Pain Intensity Among Patients With Chronic Low Back Pain: The Mint Randomized Clinical Trials. JAMA 2017;318(1):68-81. Neuromodulation: Technology at the Neural Interface. 2017;20(8):844-844. doi:https://doi.org/10.1111/ner.12729
- Dengler J, Kools D, Pflugmacher R, et al. Randomized Trial of Sacroiliac Joint Arthrodesis Compared with Conservative Management for Chronic Low Back Pain Attributed to the Sacroiliac Joint. *The Journal of Bone and Joint Surgery*. 2019;101(5):400-411. doi:https://doi.org/10.2106/jbjs.18.00022
- Duhon BS, Bitan F, Lockstadt H, Kovalsky D, Cher D, Hillen T. Triangular Titanium Implants for Minimally Invasive Sacroiliac Joint Fusion: 2-Year Follow-Up from a Prospective Multicenter Trial. *International Journal of Spine Surgery*. 2016;10:13. doi:https://doi.org/10.14444/3013
- Whang PG, Darr E, Meyer SC, et al. Long-Term Prospective Clinical And Radiographic Outcomes After Minimally Invasive Lateral Transiliac Sacroiliac Joint Fusion Using Triangular Titanium Implants. *Medical Devices: Evidence and Research*. 2019;Volume 12:411-422. doi:https://doi.org/10.2147/mder.s219862
- Block J, Miller, Reckling. Analysis of postmarket complaints database for the iFuse SI Joint Fusion System[®]: a minimally invasive treatment for degenerative sacroiliitis and sacroiliac joint disruption. *Medical Devices: Evidence and Research*. 2013;6:77. doi:https://doi.org/10.2147/mder.s44690
- 39. Cher D, Reckling WC, Capobianco R. Implant survivorship analysis after minimally invasive sacroiliac joint fusion using the iFuse Implant System[®]. *Medical Devices: Evidence and Research*. 2015;8:485. doi:https://doi.org/10.2147/mder.s94885
- Strand N, D'Souza RS, Hagedorn JM, et al. Evidence-Based Clinical Guidelines from the American Society of Pain and Neuroscience for the Use of Implantable Peripheral Nerve Stimulation in the Treatment of Chronic Pain. *Journal of Pain Research*. 2022;15:2483-2504. doi:https://doi.org/10.2147/JPR.S362204
- Melzack R, Wall PD. Pain Mechanisms: A New Theory. Science. 1965;150(3699):971-978. doi:https://doi.org/10.1126/ science.150.3699.971
- 42. Goroszeniuk T, Pang D. Peripheral Neuromodulation: A Review. *Current Pain and Headache Reports*. 2014;18(5). doi:https://doi. org/10.1007/s11916-014-0412-9
- Meyer-Frießem CH, Wiegand T, Eitner L, et al. Effects of Spinal Cord and Peripheral Nerve Stimulation Reflected in Sensory Profiles and Endogenous Pain Modulation. *The Clinical Journal of Pain*. 2019;35(2):111-120. doi:https://doi.org/10.1097/ ajp.000000000000661
- Guentchev M, Preuss C, Rink R, Peter L, Sailer M, Jochen Tuettenberg. Long-Term Reduction of Sacroiliac Joint Pain With Peripheral Nerve Stimulation. *Operative Neurosurgery*. 2017;13(5):634-639. doi:https://doi.org/10.1093/ons/opx017
- 45. Navani A, Manchikanti L, Albers SL, et al. Responsible, Safe, and Effective Use of Biologics in the Management of Low Back Pain: American Society of Interventional Pain Physicians (ASIPP) Guidelines. *Pain Physician*. 2019;22(1S):S1-S74.
- 46. Choi WS, Kim JS, Ryu KS, Hur JW, Seong JH, Cho HJ. Endoscopic Radiofrequency Ablation of the Sacroiliac Joint Complex in the Treatment of Chronic Low Back Pain: A Preliminary Study of Feasibility and Efficacy of a Novel Technique. *BioMed Research International*. 2016;2016. doi:https://doi.org/10.1155/2016/2834259
- 47. Ibrahim R, Telfeian AE, Gohlke K, Decker O. Endoscopic Radiofrequency Treatment of the Sacroiliac Joint Complex for Low Back Pain: A Prospective Study with a 2-Year Follow-Up. *Pain Physician*. 2019;22(2):E111-E118.



BPI

Letter to the Editor

OZONIZED ORAL GEL AS AN ADJUVANT IN THE TREATMENT OF PERIODONTAL DISEASE: A PRELIMINARY REPORT

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ABSTRACT

Ozonized oils have been demonstrated to induce the reduction of many oral microorganisms. The aim of this study was to evaluate the efficacy of a new ozonized oil formulation for the treatment of periodontal disease. A total of 10 patients with a diagnosis of chronic periodontitis were randomly selected, and a split-mouth scheme was used. All patients underwent to support periodontal therapy at the baseline measurement. Microbial sampling and analysis were performed in each selected site before supporting periodontal therapy. The selected site corresponded to the deepest periodontal pocket of the oral cavity. *Aggregatibacter actinomycetemcomitans, Porphyromonas gingivalis, Tannerella forsythia, Treponema denticola, Fusobacterium nucleatum, Campylobacter rectus*, and Total Bacterial Loading were evaluated through the quantification of total bacterial genome copies by PCR. Then, support periodontal therapy was done using an ultrasonic scaler. After support periodontal therapy, each patient was given ozonized sunflower seed oil [Ozoral gel, Innovares SRL, Sant'Ilario d'Enza (RE), Italy]. The patients were instructed to apply the gel daily after evening oral hygiene at home. After 2 weeks, microbiological samples were collected again in each patient and analyzed. A statistically significant difference was detected between Total Bacterial Loading (p<0.014) and *Tannerella Forsythia* (p<0.012) pre and post-ozonized sunflower seed oil treatment. Ozoral has demonstrated antiseptic properties. Additional studies with larger sample sizes are needed to confirm this preliminary result.

KEYWORDS: ozonized oil, ozone therapy, periodontal disease, periodontal pathogens

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INTRODUCTION

Ozonized oils, like ozonized sunflower oil, have been demonstrated to induce the reduction of many oral microorganisms (1). Ozonation of edible oil is performed by bubbling the gas mixture (O2/O3) into the oil under a controller reaction environment. This preparation is ideal for topical use in treating chronically infected cutaneous and mucosal areas of the body (2). Ozonized oils are widely recognized as one of the best bactericidal, antiviral and antifungal agents, and, therefore, it is profitably and practically employed in medicine and odontology. In this sense, studies have been carried out in peri-implant mucositis (3), caries prevention (4), periodontal diseases (5), regeneration and wound healing of the extraction socket and surgical site (6, 7).

Plaque biofilm is the main cause of both caries and periodontal disease. Ozonized oils have been proven useful in controlling oral infectious microorganisms in dental plaque (3). The antimicrobial property of ozonized oils effectively reduces the number of various periodontal bacteria (8). Ozonized oil seems to exert its antimicrobial action through different mechanisms, including: 1) Direct oxidation (germicide) (2, 9, 10); 2) Cytotoxicity (11); 3) Grow factors Release (12) and 4) Oxidative pre-conditioning (13).

Various etiological factors cause oral lesions, and microorganisms play a major role (14). Elimination of these microbial pathogens is the aim of most dental treatments. It has been demonstrated that ozonated sunflower oil effectively kills the biofilms formed by Candida species and the bacterium *Streptococcus mutans* (15).

The efficacy and safety of ozonized oil is closely linked to its quality control. The peroxide value is one of the basic parameters to define the dosage and its clinical application (16). This indicator is critical to define the proper indication. Lack of standardization and quality control of ozonized oils may cause variability when the germicide capacity is assayed. A new ozonized sunflower seed oil [Ozoral gel, Innovares SRL, Sant'Ilario d'Enza (RE), Italy] has recently been introduced in the market for periodontal treatments (3, 5). Ozoral® is a mucoadhesive hydrogel containing 15% of Ozonia3000® Sunflower, an ozonized sunflower seed oil registered to ECHA (European Chemical Agency https://echa.europa.eu/it/regulations/reach/legislation and classified as non-toxic, non-irritating and non-hazardous by ingestion. The muco-adhesiveness of Ozoral® is due to a polysaccharide of vegetable origin, which favours adhesion and permanence of the product on the oral mucosa despite the humidity.

In the present study, the antimicrobial properties of this new ozonized sunflower seed oil against oral and periodontal pathogens have been evaluated using the quantification method of total bacterial genome copies by PCR. The study's null hypothesis was that the ozonized sunflower seed oil did not demonstrate antibacterial effects; it does not affect antibacterial capabilities in addition to supporting periodontal therapy (SPT).

MATERIALS AND METHODS

The study was a single-centre clinical trial. A total of 10 patients with a diagnosis of chronic periodontitis were randomly selected. Patients enrolled in this study were 35-55 years old. Subjects had not previously received any surgical or non-surgical periodontal therapy. The patients were excluded from the study if they met the following criteria: pregnancy; a history of taking antibiotics or using antibacterial mouth rinses for the past 6 months; teeth with furcation involvement; smoking, and drug or alcohol abuse. Subjects participating in the study volunteered to follow a detailed verbal description of the procedure and signed consent forms. This trial was approved by the Albanian University Ethical Committee n 232.

A total of 10 patients were selected, and a split-mouth scheme was used. The patients were treated with SPT. Before SPT, microbial sampling and analysis were performed in each selected site. The selected site corresponded to the deepest periodontal pocket of the oral cavity. Microbiological samples were collected from each patient. For bacteria analysis, sites were isolated using cotton rolls. Sterile absorbable paper points (size 60) were used to collect subgingival samples and were immediately transferred to the microbiological laboratory for processing. *Aggregatibacter actinomycetemcomitans, Porphyromonas gingivalis, Tannerella forsythia, Treponema denticola, Fusobacterium nucleatum, Campylobacter rectus*, and Total Bacterial Loading were evaluated. Then SPT was done using an ultrasonic scaler. After SPT, ozonized oil (Ozoral®) was given to each patient for use on the left side of the oral cavity. The patient was instructed to apply the gel once a day after evening oral hygiene. After 2 weeks, microbiological samples were collected again in each patient.

Ozoral gel was supplied by Innovares SRL, Sant'Ilario d'Enza (RE), Italy. The manufacturer did a quality control report of the batch. The method to assay the peroxide values was ISCO3 (2016) (17). Peroxide Values in Ozonized Oils - www.isco3.org.

Probes oligonucleotides were designed based on 16S rRNA gene sequences of the Human Oral Microbiome Database (HOMD 16S rRNA RefSeq Version 10.1), counting 845 entries. All the sequences were aligned to find either a consensus sequence or less conservated spots. Two real-time polymerase chain reaction (PCR) runs were performed for each sample. The first reaction quantified the total amount of bacteria using two degenerate primers and a single probe matching a highly conservated 16S ribosomal RNA gene sequence. The second reaction detected and quantified the following bacteria: *Aggregatibacter actinomycetemcomitans, Porphyromonas gingivalis, Tannerella forsythia, Treponema denticola, Fusobacterium nucleatum, Campylobacter rectus.* Oligonucleotide concentrations and PCR conditions were optimized to ensure sensitivity, specificity, and no inhibitions in case of unbalanced target amounts. Absolute quantification assays were performed using the Applied Biosystems 7500 Sequence Detection System. The amplification profile was initiated by a 10 min incubation period at 95°C to activate polymerase, followed by a two-step amplification of 15 s at 95°C and 60 s at 57°C for 40 cycles. All these experiments, including non-template controls, were performed to exclude reagent contamination.

Plasmids containing synthetic DNA target sequences (Eurofin MWG Operon, Ebersberg Germany) were standard for the quantitative analysis. Standard curves for each target were constructed in a triplex reaction using a mix of the same plasmids in serial dilutions ranging from 101 to 107 copies. There was a linear relationship between the threshold cycle values plotted against the copy number log over the entire range of dilutions. The copy numbers for individual plasmid preparations were estimated using the Thermo NanoDrop spectrophotometer; the absolute quantification of total bacterial genome copies in samples allowed for calculating a relative number of bacterial species. Plasmid purification and handling was performed in a separate laboratory with dedicated pipettes to prevent samples and polymerase chain reaction contamination.

Descriptive statistics (mean, standard deviation, minimum, median, and maximum) were calculated for each group and variable. The data normality of the distributions was calculated with the Kolmogorov–Smirnov test. The Friedman non-parametric test was then performed, followed by Dunn's post hoc test. Significance was predetermined as p < 0.05 for all the tests performed. SPSS program and paired simple statistic T-test were used to detect significant differences.

RESULTS

A statistically significant difference was detected between Total Bacterial Loading (p<0.014) and *Tannerella forsythia* (p<0.012) pre and post-ozonized oil treatment (Table I). When the p-value is less than 0.05, the difference between the two compared bacterial loadings is statistically significant.

	Pairwise differences					t	Degree of	р
							freedom	value
Couple	Media	Standard deviation	Standard error	Confidence	interval for the 95% ference			
				inferior	superior			
AA1 - AA2	130450	334141	105664	-108579	369481	1.235	9	.248
PG1 - PG2	-136	2700	854	-2068	1795	160	9	.877
TF1 - TF2	18552	18768	5934	5126	31978	3.126	9	.012
TD1 - TD2	19213	32478	10270	-4020	42446	1.871	9	.094
FN1 - FN2	1107097	1682268	531979	-96324	2310519	2.081	9	.067
CR1 - CR2	68025	187066	59155	-65793	201844	1.150	9	.280
TBL1 - TBL2	1365269	1429368	452005	342761	2387778	3.020	9	.014

 Table I. paired sample test.

AA: Aggregatibacter actinomycetemcomitans; PG: Porphyromonas gingivalis; TF: Tannerella forsythia; TD: Treponema denticola; FN: Fusobacterium nucleatum; CR: Campylobacter rectus; TBL: Total Bacterial Loading 1 pre-treatment, 2 post-treatment. The table reports data on the treated side only, pre-and post-treatment.

DISCUSSION

Ozonized oil seems to strongly inhibit the formation of dental plaque and reduce the number of pathogens, both Grampositive and Gram-negative organisms, including: *Staphylococci, Streptococci, Enterococci, Pseudomonas, Escherichia coli* and, above all, against Mycobacteria (18, 19) This effect of oxidation gives to its bactericidal, virucidal, and fungicidal activity. After contact with ozonized oil - microorganism, severe alteration of the cytoplasm was observed (20). In addition, applying ozonized oil reduces amylase, lipase, keratinase and urease enzyme activities in the microorganism significantly, in line with a reduction in nucleic acid content (11). This action seems not to damage human body cells; the reason attributed to this is the antioxidant ability of mammalian cells (21).

Even when the exact action mechanism of the ozonized oil is not described, there is much pre-clinical and clinical evidence of its antimicrobial and wound healing beneficial efficacy. As an antimicrobial, the most sensible bacterium is *Staphylococcus aureus*, and the primary resistant is *Pseudomonas aeruginosa* (22). A recent *in vitro* study confirms the microorganism sensibility to ozonized oil in that way (from more to less sensibility): *Staphylococcus aureus* > *Candida albicans* > *Escherichia coli* > *Pseudomonas aeruginosa* > *Enterococcus faecalis* (23).

In general, a lethal effect of ozonized oil is evident when it is applied to a multi-resistant strain of *Staphylococcus* epidermis, *Staphylococcus aureus*, also when it is applied to fungi from the genus *Trichophyton*, *Epidermophyton* and *Microsporum*, yeast as *Candida albicans* and protozoan as *Giardia lamblia* (24, 25).

A comparison regarded the antimicrobial effectiveness of ozonized extra virgin olive oil (peroxide value of 560/590 mEq/kg) with 0.2% chlorhexidine digluconate and 10% povidone-iodine through a disk diffusion test was done recently (8). Ozonized oil showed a significantly better behaviour than the references. This effect on one of the main pathogens suggests its potential applicability for periodontal treatment (8).

However, the word *ozonized* is without scientific meaning if it is not associated with *how much* peroxides are present in the oil. Probably the leading cause of variability regarding the microbiological efficacy of these active components is closely connected with the lack of standardization. The few studies on the therapeutic effects of ozonized oils on acute cutaneous wound healing in animal models did not investigate the dose/effect response, expressed as the number of peroxides in the ozonized derivative used (26).

Our study evaluated the antibacterial properties of a standardized ozonized sunflower seed oil in a group of patients who used it as a home-care praesidium. Ozoral gel has been statistically significant (p<0.05) in reducing Total Bacterial Loading and *Tannerella forsythia* bacterial loading. This last bacterium belongs to Socransky's red complex, a periodontal pathogen (27-32). Thus, Ozoral gel demonstrated antibacterial effects. In contrast with previous studies, based on the only analysis of traditional microbiology, these results demonstrate for the first time the reduction of the total bacterial load by ozonized oils, using quantification of total bacterial genome copies by PCR; this should explore the significance of the results of the work, not repeat them. A combined Results and Discussion section is often appropriate. Avoid extensive citations and discussion of published literature.

CONCLUSIONS

It is our knowledge, however, that additional studies with a larger sample size and a higher number of home-care applications are needed to firmly demonstrate the effectiveness of ozonized oil as a viable antimicrobial agent in routine dental therapies.

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REFERENCES

- 1. Saini R. Ozone therapy in dentistry: A strategic review. J Nat Sci Biol Med. Jul 2011;2(2):151-153.
- 2. Valacchi G, Fortino V, Bocci V. The dual action of ozone on the skin. Br J Dermatol. Dec 2005;153(6):1096-1100.
- Butera A, Pascadopoli M, Gallo S, et al. Ozonized Hydrogels vs. 1% Chlorhexidine Gel for the Clinical and Domiciliary Management of Peri-Implant Mucositis: A Randomized Clinical Trial. J Clin Med. Feb 12 2023;12(4).
- 4. Nardi GM, Fais S, Casu C, et al. Mouthwash Based on Ozonated Olive Oil in Caries Prevention: A Preliminary In-Vitro Study. *Int J Environ Res Public Health.* Dec 6 2020;17(23).
- 5. Tete G, D'Amicantonio T, Polizzi E. Efficacy Ozone Therapy in Reducing Periodontal Disease. Materials (Basel). Mar 16 2023;16(6).
- 6. Anzolin AP, da Silveira-Kaross NL, Bertol CD. Ozonated oil in wound healing: what has already been proven? *Med Gas Res.* Jan-Mar 2020;10(1):54-59.
- 7. Lim Y, Lee H, Woodby B, Valacchi G. Ozonated Oils and Cutaneous Wound Healing. Curr Pharm Des. 2019;25(20):2264-2278.
- 8. Montevecchi M, Dorigo A, Cricca M, Checchi L. Comparison of the antibacterial activity of an ozonated oil with chlorhexidine digluconate and povidone-iodine. A disk diffusion test. *New Microbiol.* Jul 2013;36(3):289-302.
- 9. Guinesi AS, Andolfatto C, Bonetti Filho I, Cardoso AA, Passaretti Filho J, Farac RV. Ozonized oils: a qualitative and quantitative analysis. *Braz Dent J.* 2011;22(1):37-40.
- Lezcano I, Nuñez N, Espino M, Gómez M. Antibacterial Activity of Ozonized Sunflower Oil, Oleozón, Against Staphylococcus aureus and Staphylococcus epidermidis. Ozone: Science & Engineering: The Journal of the International Ozone Association. 2000 2000;22(2):207.
- 11. Neveen SI. Antifungal activity of ozonized Olive Oils (Oleozone). International J of Agriculture and Biology 2006;8(5):670-675.
- 12. Bocci V. Ozone A New Medical Drug. Dordrecht, The Nederlands: Springer; 2005.
- 13. Zamora Z, Gonzalez R, Guanche D, et al. Ozonized sunflower oil reduces oxidative damage induced by indomethacin in rat gastric mucosa. *Inflamm Res.* Jan 2008;57(1):39-43.
- 14. González-Munoz L, Flichy-Fernández AJ, Ata-Ali J, Pascual-Moscardó A, Penarrocha-Diago MA. Effect of ozone therapy upon clinical and bacteriological parameters of the oral cavity: an update. *Journal of Clinical and Experimental Dentistry*. 2011;3(4):e325-e327.
- 15. Higa B, Cintra BS, Alvarez CM, et al. Ozonated oil is effective at killing Candida species and Streptococcus mutans biofilmderived cells under aerobic and microaerobic conditions. *Med Mycol*. Aug 8 2022;60(8).
- ISCO3, Martínez-Sánchez G, Lozano ÓL. Physico-chemical characterization of ozonized oil. Peroxide Value. http://isco3.org/ officialdocs/#4. 2016;ISCO3/LAB/00/04
- 17. Zanardi I, Travagli V, Gabbrielli A, Chiasserini L, Bocci V. Physico-Chemical Characterization of Sesame Oil Derivatives. *Lipids*. 2008;43(9):877-886. doi:https://doi.org/10.1007/s11745-008-3218-x
- 18. Sechi LA, Lezcano I, Nunez N, et al. Antibacterial activity of ozonized sunflower oil (Oleozon). *J Appl Microbiol*. Feb 2001;90(2):279-284.
- 19. Rodrigues KL, Cardoso CC, Caputo LR, Carvalho JCT, Fiorini JE, Schneedorf JM. Cicatrizing and antimicrobial properties of an ozonized oil from sunflower seeds. *Inflammopharmacology*. 2004;12(3):261-270.
- 20. Sechi LA. Antibacterial activity of ozonized sunflower oil (Oleozon). Journal of Applied Microbiology. 2001;90:279-284.
- 21. Izzotti A, Fracchia E, Rosano C, et al. Efficacy of High-Ozonide Oil in Prevention of Cancer Relapses Mechanisms and Clinical Evidence. *Cancers (Basel)*. Feb 24 2022;14(5).
- 22. Menéndez s, González R, Ledea O. Ozono, aspectos básicos y aplicaciones clínicas. La Habana: CENIC; 2008.
- 23. Zanardi I, Burgassi S, Paccagnini E, Gentile M, Bocci V, Travagli V. What is the best strategy for enhancing the effects of topically applied ozonated oils in cutaneous infections? *Biomed Res Int.* 2013;2013:702949.
- 24. Menendez S, Falcon L, Simon DR, Landa N. Efficacy of ozonized sunflower oil in the treatment of tinea pedis. *Mycoses*. Oct 2002;45(8):329-332.

- 25. Hernandez F, Hernandez D, Zamora Z, et al. Giardia duodenalis: effects of an ozonized sunflower oil product (Oleozon) on in vitro trophozoites. *Exp Parasitol*. Mar 2009;121(3):208-212.
- 26. Kim HS, Noh SU, Han YW, et al. Therapeutic effects of topical application of ozone on acute cutaneous wound healing. *J Korean Med Sci.* Jun 2009;24(3):368-374.
- 27. Haffajee AD, Socransky SS. Microbial etiological agents of destructive periodontal diseases. Periodontol 2000. Jun 1994;5:78-111.
- 28. Lauritano D, Carinci F, Palmieri A, Cura F, Caruso S, Candotto V. Reuterinos((R)) as adjuvant for peri-implant treatment: A pilot study. *Int J Immunopathol Pharmacol.* Jan-Dec 2019;33:2058738419827745.
- 29. Roncati M, Lauritano D, Cura F, Carinci F. Evaluation of light-emitting diode (LED-835 NM) application over human gingival fibroblast: an in vitro study. *J Biol Regul Homeost Agents*. Apr-Jun 2016;30(2 Suppl 1):161-167.
- Lopez MA, Andreasi Bassi M, Confalone L, Carinci F, Ormianer Z, Lauritano D. The use of resorbable cortical lamina and micronized collagenated bone in the regeneration of atrophic crestal ridges: a surgical technique. Case series. J Biol Regul Homeost Agents. Apr-Jun 2016;30(2 Suppl 1):81-85.
- 31. Scarano A, Murmura G, Carinci F, D. Lauritano D. Immediately loaded small-diameter dental implants: Evaluation of retention s, and comfort for the edentulous patient (2012) European Journal of Inflammation, 10 (1), pp. 19-23. Immediately loaded small-diameter dental implants: Evaluation of retention, stability, and comfort for the edentulous patient. *Eur J Inflamm*. 2012;10(1):19-23.
- 32. Carinci F, Girardi A, Palmieri A, et al. LAB®-Test 1: Peri-Implantitis and bacteriological analysis Eur J Inflamm. 2012;10(1):91-93.



Letter to the Editor

OSTEONECROSIS OF THE MANDIBLE ASSOCIATED WITH ZOLEDRONATE THERAPY

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INTRODUCTION

Bisphosphonates are synthetic compounds that mimic the structure of a naturally occurring substance called inorganic pyrophosphate (1). They have a high affinity for hydroxyapatite, the mineral component of bone tissue, and bind to regions of bone with high turnover (2); this allows them to effectively inhibit the activity of osteoclasts, which are the cells responsible for breaking down and resorbing bone tissue (3). As a result, bisphosphonates can be used to treat various bone disorders by preventing bone resorption at the molecular, cellular, and tissue level (4). The standard of care for treating osteopenia and osteoporosis, as well as Paget's disease and Osteogenesis imperfecta, still involves oral bisphosphonates (5). Additionally, intravenous bisphosphonates such as pamidronate (Aredia) and zoledronic acid (Zometa) are also used for these conditions (6).

Multiple myeloma and metastatic bone lesions are commonly treated with pamidronate and zoledronic acid, effectively preventing skeletal complications like pathologic fractures and hypercalcemia of malignancy (7). The action of bisphosphonates involves several mechanisms, such as inhibiting the differentiation of osteoclast precursors, promoting apoptosis of osteoclasts, and stimulating the release of osteoclastic inhibitory factors to osteoblasts (8). Additionally, these compounds can interfere with cellular metabolism by resembling adenosine triphosphate (ATP), disrupting cellular processes and further reducing osteoclast activity (9).

While bisphosphonates offer various therapeutic benefits, a notable complication that can arise in some patients receiving these drugs is bisphosphonate-related osteoradionecrosis of the jaws (BRONJ) (10). This condition, first identified and reported by Marx in 2003 (11), can have significant consequences for affected individuals. The onset of symptoms in BRONJ can be unpredictable, making it difficult for clinicians to diagnose and manage the condition promptly (12). This

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variability in presentation often means that the disease is only identified once it has become symptomatic, posing a challenge for effective diagnosis and management (13). The following report describes a case of BRONJ involving the lower maxilla in a patient suffering from multiple myeloma.

CASE REPORT

A 74-year-old female patient reported pain and swelling in the left lower jaw that has been present for several weeks.

The patient's medical history reveals that she has a history of diabetes, hypertension, and kidney failure, in addition to the previous history of multiple myeloma and treatment with intravenous zoledronate for two years. Upon local examination, it was observed that the patient had partial edentulism and alveolar mucosal dehiscence in the left body of the mandible. In addition, there was evidence of an exposed bone section that appeared yellowish-white in colour close to the posterior mandibular region, next to the roots of the first left lower molar (Fig. 1).

The orthopantomogram revealed the presence of diffuse osteolytic lesions and erosion on the left side of the mandible, affecting both the buccal and lingual cortical plates (Fig. 2). The size of the exposed bone gradually expanded, and the area became increasingly more sensitive to pain. The first-line conservative measures, comprising administration of chlorhexidine 0.2% mouthwash and oral antibiotics, resulted in a reduction of pain but failed to bring about resolution of the denuded bone.

Considering the progressive nature of the BRONJ, surgical intervention of sequestrectomy was subsequently planned. Using a large round burr and saline irrigation, the necrotic maxillary alveolus was excised until healthy, actively bleeding bone was achieved. Subsequently, a biopsy was performed to affirm the bone's vitality and exclude any malignancy. One year later, the patient reported discomfort, swelling, and pain, prompting the need for further clinical evaluation. Cone beam computed tomography revealed loss of integrity and diffused erosion of the lingual cortical bone. (Fig. 3).

The therapeutic approach was based on administering amoxicillin + clavulanic ac 1gr x 2 /day, metronidazole 1000 mg/day for 15 days, and oral chlorhexidine (0.12%) rinses three times a day. Intravenous Zoledronate was stopped. Spontaneous bone sequestration eventually occurred a few months later, followed by the mandibular bone's stable and painless mucosal coverage. At follow-up after one year, the patient was disease-free (Fig. 4).

DISCUSSION

There are several possible causes of swelling in the jaws.



Fig. 1. *Exposed bone section that appeared yellowishwhite in color close to the posterior mandibular region, next to the roots of the first left lower molar.*



Fig. 2. *The orthopantomogram revealed the presence of diffuse osteolytic lesions on the left side of the mandible.*



Fig. 3. Cone beam computed tomography after one year.



Fig. 4. Follow-up after one year.
An infection in the jaw can cause swelling, pain, and tenderness due to poor oral hygiene, dental caries, or a periodontal abscess (14). Furthermore, disorders of the salivary glands, such as sialadenitis or sialolithiasis, can cause swelling in the jaw (15). Benign or malignant tumors and cysts can develop in the jaw and lead to swelling (16). Also, swelling and tenderness may occur in patients with temporomandibular joint (TMJ) disorders (17). Lastly, a fracture, an injury, or other trauma to the jaw can cause swelling (18).

In the abovementioned case, the patient's medical history reveals a 2-year treatment course of intravenously administered bisphosphonates, specifically zoledronic acid, after diagnosing multiple myeloma. The patient presented with clinical symptoms of swelling and exposed non-vital bone in the left alveolar region. This presentation is consistent with the possibility of jaw osteonecrosis, a rare but known complication associated with the long-term use of bisphosphonates.

An oral-maxillofacial surgeon first reported osteonecrosis of the maxilla and mandible as a complication of intravenous bisphosphonate treatment in 2003 (11). Following this, osteonecrosis of the jaw was also reported in patients taking oral bisphosphonates for osteoporosis (19). Since then, the number of reported cases of BRONJ has increased, but the estimated incidence varies considerably from less than 1% to 18.6% (20). The drug's potency, administration route, and therapy duration are determining factors, with zoledronate (Zometa) having the highest reported incidence and the oral forms having a relatively low incidence (21). Both dental and oncological practitioners must possess a thorough awareness of the significant risk of developing osteonecrosis of the jaw (ONJ) in patients who are receiving bisphosphonate therapy (22).

Hence, the reported case re-affirms that dental and oncological professionals must exercise caution when treating patients receiving BPs and provide appropriate education and monitoring to minimize the risk of ONJ development.

REFERENCES

- De Rosa G, Misso G, Salzano G, Caraglia M. Bisphosphonates and Cancer: What Opportunities from Nanotechnology? *Journal of Drug Delivery*. 2013;2013:1-17. doi:https://doi.org/10.1155/2013/637976
- Carvalho MS, Cabral JMS, da Silva CL, Vashishth D. Bone Matrix Non-Collagenous Proteins in Tissue Engineering: Creating New Bone by Mimicking the Extracellular Matrix. *Polymers*. 2021;13(7):1095. doi:https://doi.org/10.3390/polym13071095
- Florencio-Silva R, Sasso GR da S, Sasso-Cerri E, Simões MJ, Cerri PS. Biology of Bone Tissue: Structure, Function, and Factors That Influence Bone Cells. *BioMed Research International*. 2015;2015(421746):1-17. doi:https://doi.org/10.1155/2015/421746
- Drake MT, Clarke BL, Khosla S. Bisphosphonates: Mechanism of Action and Role in Clinical Practice. *Mayo Clinic Proceedings*. 2008;83(9):1032-1045. doi:https://doi.org/10.4065/83.9.1032
- Golu MV, Paşcanu I, Togănel C, et al. What Do Prescribers of Bone Modifying Agents Know about Medication-Related Osteonecrosis of the Jaw? Is Current Prevention Enough? *Applied Sciences*. 2022;12(18):9224. doi:https://doi.org/10.3390/app12189224
- Tanvetyanon T, Stiff PJ. Management of the adverse effects associated with intravenous bisphosphonates. *Annals of Oncology*. 2006;17(6):897-907. doi:https://doi.org/10.1093/annonc/mdj105
- Rosen LS, Gordon D, Kaminski M, et al. Zoledronic acid versus pamidronate in the treatment of skeletal metastases in patients with breast cancer or osteolytic lesions of multiple myeloma: a phase III, double-blind, comparative trial. *Cancer Journal (Sudbury, Mass)*. 2001;7(5):377-387.
- Bellido T, Plotkin LI. Novel actions of bisphosphonates in bone: Preservation of osteoblast and osteocyte viability. *Bone*. 2011;49(1):50-55. doi:https://doi.org/10.1016/j.bone.2010.08.008
- Billig H, Rosberg S, Johanson C, Ahrén K. Adenosine as substrate and receptor agonist in the ovary. *Steroids*. 1989;54(5):523-542. doi:https://doi.org/10.1016/0039-128x(89)90045-7
- Nicolatou-Galitis O, Schiødt M, Mendes RA, et al. Medication-related osteonecrosis of the jaw: definition and best practice for prevention, diagnosis, and treatment. *Oral Surgery, Oral Medicine, Oral Pathology and Oral Radiology*. 2019;127(2):117-135. doi:https://doi.org/10.1016/j.oooo.2018.09.008
- 11. Marx RE. Pamidronate (Aredia) and zoledronate (Zometa) induced avascular necrosis of the jaws: a growing epidemic. *Journal of Oral and Maxillofacial Surgery*. 2003;61(9):1115-1117. doi:https://doi.org/10.1016/s0278-2391(03)00720-1

- 12. Kishimoto H, Noguchi K, Takaoka K. Novel insight into the management of bisphosphonate-related osteonecrosis of the jaw (BRONJ). *Japanese Dental Science Review*. 2019;55(1):95-102. doi:https://doi.org/10.1016/j.jdsr.2018.09.002
- Vogt-ferrier NB, Hugentobler MA, Uebelhart B, Tramèr MR, Rollason V. Interventions for treating osteonecrosis of the jaw bones associated with bisphosphonates. *Cochrane Database of Systematic Reviews*. Published online April 14, 2010. doi:https://doi. org/10.1002/14651858.cd008455
- 14. Duangthip D, Chu CH. Challenges in Oral Hygiene and Oral Health Policy. *Frontiers in Oral Health*. 2020;1. doi:https://doi. org/10.3389/froh.2020.575428
- 15. Wilson KF, Meier JD, Ward PD. Salivary gland disorders. American Family Physician. 2014;89(11):882-888.
- Puri N, Ahuja US, Dhillon M, Rathore A. Ultrasonography as a Diagnostic Aid in Evaluating Cystic Lesions, Benign Tumors and Malignancies of Maxillofacial Region: A Clinical Study. *The Open Dentistry Journal*. 2018;12(1):1050-1058. doi:https://doi. org/10.2174/1874210601811131050
- 17. Reed LS, Foster MD, Hudson JW. Synovial Chondromatosis of the Temporomandibular Joint: A Case Report and Literature Review. *CRANIO* (2013;31(4):309-313. doi:https://doi.org/10.1179/crn.2013.31.4.009
- Follmar KE, Marklieke DeBruijn, Alessio Baccarani, et al. Concomitant Injuries in Patients With Panfacial Fractures. *J Trauma*. 2007;63(4):831-835. doi:https://doi.org/10.1097/ta.0b013e3181492f41
- 19. Hewitt C, Farah CS. Bisphosphonate-related osteonecrosis of the jaws: a comprehensive review. *Journal of Oral Pathology & Medicine*. 2007;36(6):319-328. doi:https://doi.org/10.1111/j.1600-0714.2007.00540.x
- Walter C, Al-Nawas B, Grötz KA, et al. Prevalence and Risk Factors of Bisphosphonate-Associated Osteonecrosis of the Jaw in Prostate Cancer Patients with Advanced Disease Treated with Zoledronate. *European Urology*. 2008;54(5):1066-1072. doi:https:// doi.org/10.1016/j.eururo.2008.06.070
- Dalle Carbonare L, Mirko Zanatta, Adriano Gasparetto, Maria Teresa Valenti. Safety and tolerability of zoledronic acid and other bisphosphonates in osteoporosis management. *Drug, Healthcare and Patient Safety*. 2010;2:121. doi:https://doi.org/10.2147/ dhps.s6285
- 22. Hewson I, Syme D, Bruscino-Raiola F. Radical surgical treatment of bisphosphonate related osteonecrosis of the jaw. *Australian Dental Journal*. 2012;57(2):227-230. doi:https://doi.org/10.1111/j.1834-7819.2012.01675.x