



*Case Reports*

## **HYDROXYAPATITE AND BETA-TRICALCIUM PHOSPHATE IN THE SOCKET PRESERVATION: PRESENTATION OF CLINICAL CASES**

M. Di Girolamo<sup>1</sup>, D. Mazza<sup>2</sup>, S. Volpe<sup>3</sup>, G. Stelitano<sup>3</sup>, L. Volpe<sup>4</sup>, F. Cecchetti<sup>2</sup>

<sup>1</sup>Department of Biomedicine and Prevention, University of Rome “Tor Vergata”, Rome, Italy;

<sup>2</sup>Department of Social Dentistry and Gnathological Rehabilitation University of Rome “Tor Vergata”, Rome, Italy;

<sup>3</sup>Private Practice, Roma;

<sup>4</sup>Universidad Europea de Valencia Grado en Odontología, Valencia, Spain

*Correspondence to:*

Michele Di Girolamo, DDS

Department of Clinical Sciences and Translational Medicine,

University of Rome “Tor Vergata”,

Rome, Italy

e-mail: micheledigirolamo@tiscalinet.it

### **ABSTRACT**

The reduction in alveolar bone volume is a direct consequence of tooth loss. Hard and soft tissue remodelling is part of the normal physiological healing process of the residual edentulous ridge. This remodelling negatively affects the size of the ridge in the buccolingual and apicocoronal aspects. Ridge atrophy after tooth loss has been shown to follow specific patterns. In the Maxilla, the alveolar buccal wall tends to reabsorb more rapidly after tooth extraction. The ridge is gradually represented by the palatal wall (centripetal resorption). The purpose of this article is to describe two clinical cases of preservation of the alveolar ridge with the use of a biomaterial and a membrane.

**KEYWORDS:** *bone, alveolus, ridge, tooth, regeneration*

### **INTRODUCTION**

In a 12-month prospective study, Schropp et al. (1) analyzed 46 premolars and molars extraction sockets from 46 patients and found a 50% ridge loss with an average horizontal loss of 6.1 mm. Two-thirds of this loss of bone volume occurs within the first 3 months. This bone remodelling process may vary according to individual local and systemic factors, but it mainly affects the horizontal bucco-coronal thickness in the anterior sites (2-4). Su et al. observed an average loss of 3.87 mm in ridge width and 1.67 mm in ridge height in the anterior and premolar sites during the first three months after extraction

Received: 04 June 2023

Accepted: 20 July 2023

ISSN: 2038-4106

Copyright © by BIOLIFE 2023

This publication and/or article is for individual use only and may not be further reproduced without written permission from the copyright holder. Unauthorized reproduction may result in financial and other penalties. **Disclosure: All authors report no conflicts of interest relevant to this article.**

(5). The ridge is gradually resorbed by the palatal wall (centripetal resorption), compromising the sagittal and axial intermaxillary relationship (6-8). This phenomenon appears progressive and often translates into aesthetic and functional challenges during rehabilitation with dental implants.

To minimize the need for bone regeneration during implant placement “Alveolar Socket Preservation” (ASP), has been described in recent decades. This technique was born as a procedure aimed at preventing, or more appropriately limiting, the alteration of the post-extraction bone crest to have an optimal implant-prosthetic rehabilitation. The different alveolar preservation techniques proposed in the literature involve biomaterials, bone grafts, their combination, and barrier membranes (9, 10). The biomaterial used to fill the alveolus should ideally inhibit the growth within the alveolus of the epithelial and connective tissue, stabilize the clot and limit bacterial contamination (11).

In addition to resorbable and non-resorbable membranes, various biomaterials used for soft tissue augmentation, such as autogenous free gingival grafts, dermal allografts and collagen matrix xenografts, were also used to seal the alveolus (12). The purpose of this article is to describe two clinical cases of preservation of the alveolar ridge with the use of a biomaterial and a membrane.

#### *Clinical cases presentation*

Both patients underwent a medical history and control of any active pathological process. All therapeutic options were examined with patients, and socket preservation and subsequent implant prosthetic rehabilitation were chosen. Considering the risks and benefits of the proposed procedure, the signature of informed consent was obtained.

#### *Case 1:*

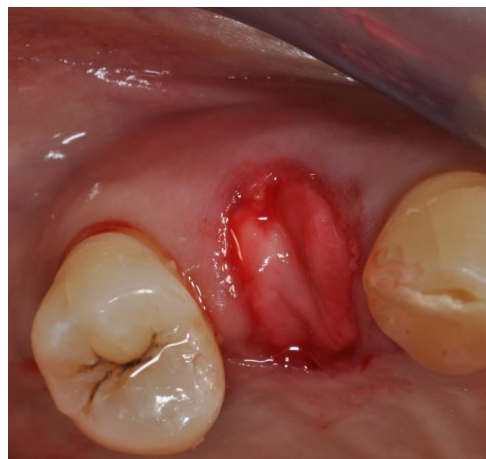
A 69-year-old patient complained of pain and mobility of 1.4. Intraoral physical examination revealed grade 2 dental mobility, associated with mesial and distal periodontal probing of about 7 mm with bleeding. The periapical X-ray confirmed the bone loss, which would not have allowed us an immediate post-extraction implantology (Fig. 1).

Under local anaesthesia (Mepivacaine 2% Sain-Maurdes-Fosses, France), the dental element was extracted, and the post-extraction alveolus was thoroughly cleaned and irrigated with a saline solution. The walls of the alveolus were examined to determine the morphology of the defect and the presence of the vestibular bone wall.

The alveolus was filled with biomaterial (Rigenera Biotek S.R.L. Pavolaro di Dueville, Vicenza, Italy) composed of 30% of Slow Resorption Hydroxyapatite (HA) and 70% of Rapidly Resorption Beta-Tricalcium phosphate. To cover the



**Fig. 1.** Case 1. Pre-operative X-ray.



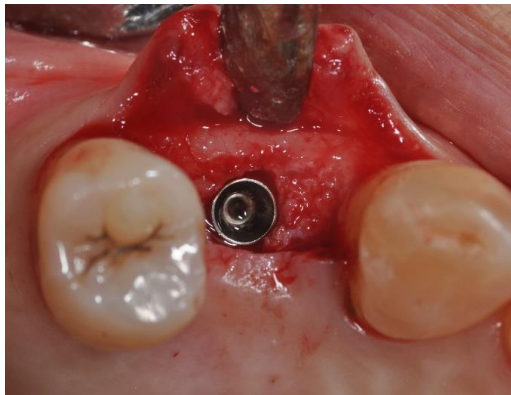
**Fig. 2.** Case 1. An occlusal vision of the exposed membrane to obtain second-intention healing.



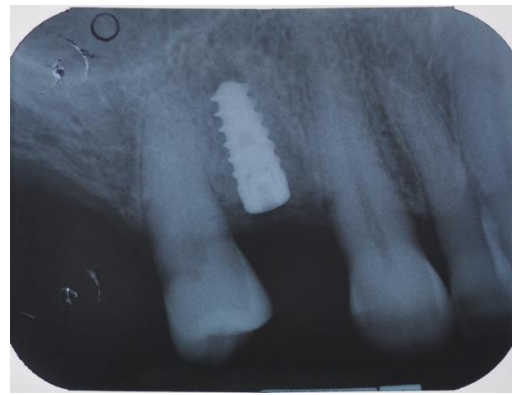
**Fig. 3.** Case 1. Radiographic control of regeneration.

biomaterial was inserted a resorbable membrane (Evolution Fine Tecnos®<sup>®</sup>, Giaveno, Italy) intentionally left exposed according to the socket Preservation technique. (Fig. 2). After 5 months of healing, clinically good preservation was evident. A control intraoral X-ray was performed (Fig. 3), and implant insertion was scheduled.

At the lifting of the flap, we noticed good preservation of the alveolus that allowed us to insert the implant 12 mm 3.75 Ø ( Safe BT Biotec srl Povolara di Dueville (VI) Italy) (Fig. 4, 5). Antibiotics (Amoxicillin 1 gr) were prescribed for 7 days. The patient underwent monthly check-ups to follow the recovery. After 4 months of osseointegration, a full-thickness flap was lifted, and the healing screw was placed (Fig. 6). Subsequently, prosthetic rehabilitation was carried out (Fig. 7-9).



**Fig. 4.** Case 1. Occlusal vision at the insertion of the implant (note the excellent preservation of the bone crest)



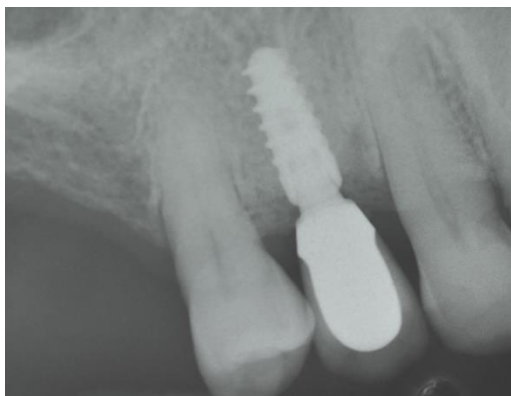
**Fig. 5.** Case 1. Control X-ray at 4 months.



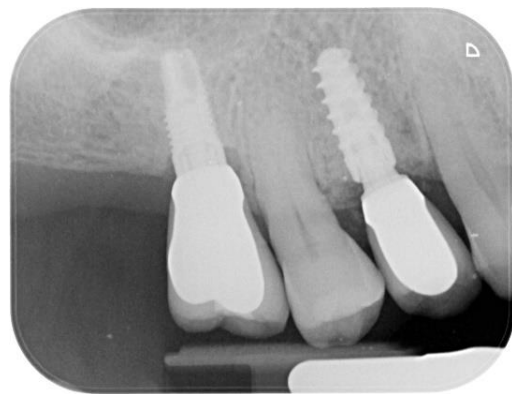
**Fig. 6.** Case 1. Insertion of the healing screw (we note how the vestibular draft has been maintained).



**Fig. 7.** Case 1. Prosthetic rehabilitation.



**Fig. 8.** Case 1. X-ray post-rehabilitation control.



**Fig. 9.** Case 1. X-ray follow-up of 2 years.

### Case 2

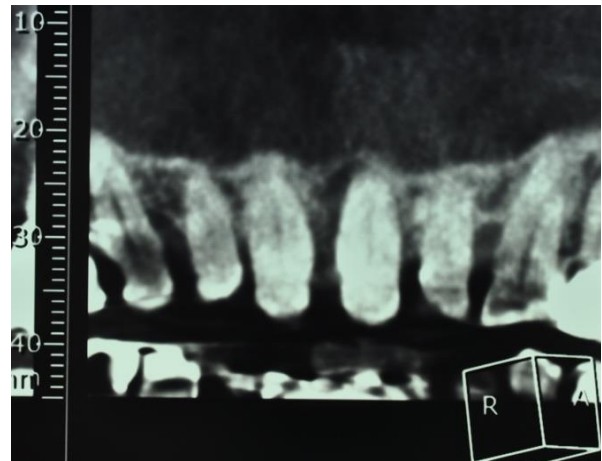
A 78-year-old man presented to our observation with a vestibular fistulized periodontal abscess of element 2.1 (Fig. 10). Periodontal probing and radiographic examination revealed mesial vertical bone resorption and the lack of part of the vestibular cortex of 2.1 (Fig. 11). With the patient's consent, it was decided to extract the dental element and carry out a subsequent implant-prosthetic rehabilitation.

As in the previous case, at the same time as the dental extraction, a reconstruction of the bone volume was carried out with a biomaterial (Rigenera Biotek S.R.L. Pavolaro di Dueville, Vicenza, Italy) and a resorbable membrane (Evolution Fine TecnoSS®, Giaveno, Italy) was inserted to cover the biomaterial. Due to the lack of much of the buccal plate, evidenced by the probing, a full-thickness pocket was prepared using a sharp blade and a microsurgical elevator. Subsequently, the membrane was shaped according to the "ice cream cone" technique (10) and partially inserted inside the pocket created previously (Fig. 12).

No primary healing intention was sought since the soft tissues would have migrated in the initial healing phase, creating a greater amount of keratinized gingiva. After 5 months, at the time of insertion of a 10 mm 3.75 Ø implant (Safe BT Biotec srl Povolario di Dueville, Vicenza, Italy), the mucosa and the underlying bone were well represented. (Fig. 13-17). After osseointegration, at 4 months of healing, the prosthetic rehabilitation was carried out (Fig. 18-20).



**Fig. 10.** Case 2. Pre-operative image shows a buccal fistula of 2.1.



**Fig. 11.** Case 2. Pre-operative CBCT shows bone loss mesially at 2.1 due to periodontal abscess.



**Fig. 12.** Case 2. Surgery-curettage of the alveolus and bone reconstruction.



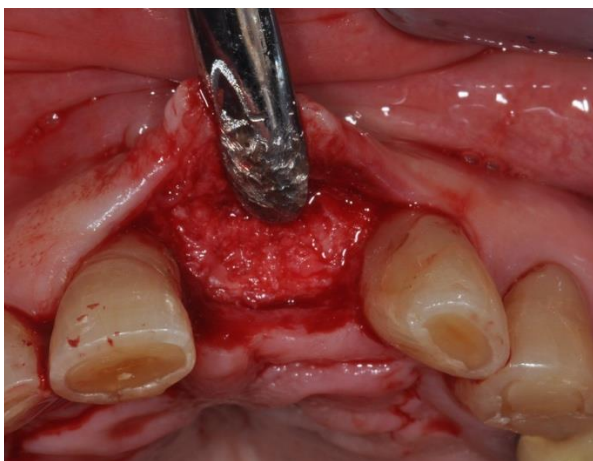
**Fig. 13.** Case 2. Healing after one week.



**Fig. 14.** Case 2. Healing after 2 weeks, the almost complete mucosal closure and the absence of inflammation are evident.



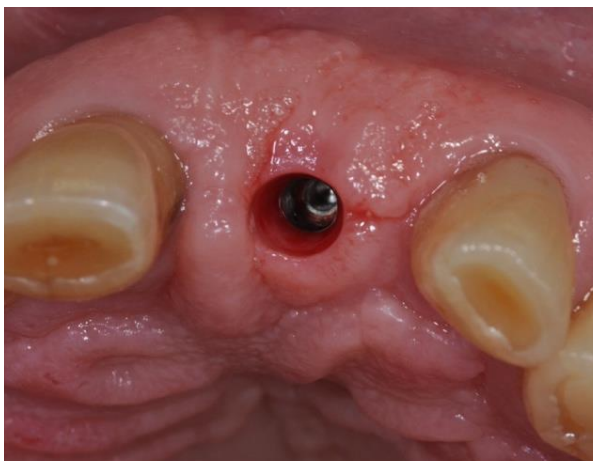
**Fig. 15.** Case 2. The appearance of the completely healed ridge.



**Fig. 16.** Case 2. When the flap is opened, excellent bone regeneration is evident which allowed us to insert an implant in the correct three-dimensional position.



**Fig. 17.** Case 2. Post-operative X-ray.



**Fig. 18.** Case 2. Detail of the buccal plate and implant position.



**Fig. 19.** Case 2. Prosthetic rehabilitation.

## DISCUSSION

The alveolus is an anatomical entity linked to the presence of the tooth in the arch. After a dental extraction, physiological healing is followed by a three-dimensional reduction of the alveolar volume (13). The tooth and its attachment apparatus, the root cement, the periodontal ligament, and the alveolar bone establish a functional unit.

The forces developed during the masticatory activity are transmitted from the tooth crown, through the root and attachment apparatus, to the hard tissues of the alveolar process, where they will be dispersed. This alveolar atrophy is the consequence of the teeth loss and the change in conditions inside or around the alveolus, and it will result in a series of adaptive alterations of the edentulous portion of the alveolar ridge.

One research article points out that the cause of the greater degree of reabsorption of the vestibular side is to be connected to the anatomy typical of the alveolar process: the buccal cortex, in fact, essentially consists of fasciculate bone, while the lingual one has a smaller fraction of it (14). The presence of vestibular tissue is functionally related to the dental element itself, and it gradually disappears after extraction leading to the reduction of the vertical and horizontal dimensions of the alveolar ridge. The purpose of socket preservation is to maintain the original alveolar dimensional contours, limiting the natural process of post-extraction resorption. A multicenter study underlined the importance and advantages of the socket preservation technique after tooth extraction of the anterior upper arch (15). These areas frequently have a thin vestibular bone plate, with a small amount of bundle bone that is inevitably lost during tooth extraction resulting in an early collapse of the labial plate and consequent imperfection (16, 17).

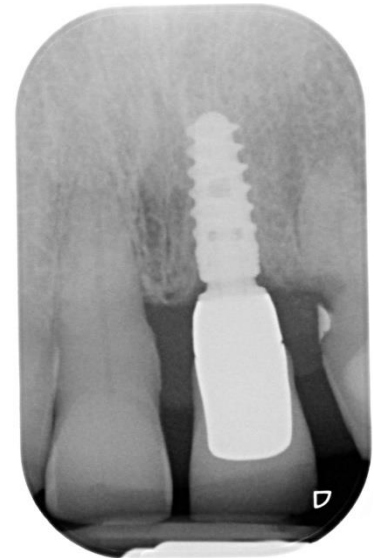
In the past years and still today, socket preservation has been the object of multiple studies with the use of different alveolar filling materials. These fall into one of four categories: autogenous bone grafts, allogeneic bone grafts, xenogenic bone grafts and alloplasts. Depending on which of these is used, the process can be osteoconductive, osteopromotor or osteoinductive (18, 19). Among the bone grafts used for alveolar preservation, biomaterials of synthesis present a growing diffusion in dental surgery thanks to their excellent bio-compatibility and the ability to act as a scaffold, stabilizing the clot in the different stages of healing.

Hydroxyapatite is one of the most commonly used biomaterials in bone regeneration techniques. Some authors underlined that hydroxyapatite synthesized in nanomolecular form, with dimensional characteristics similar to the molecules deposited by human osteoblasts, has better bone resorption and better bone new formation than traditional biomaterials (20). Furthermore, if these hydroxyapatite nano molecules are combined with rapidly resorbing substances, better results are obtained in terms of resorption of the nanohydroxyapatite, leaving space for bone regeneration (21, 22).

The material used as a filler in the two clinical cases is Rigenera BCP in 0.25-1.00 mm granules (Biotec S.R.L. Povaloro di Dueville, Vicenza, Italy) composed of 30% slow resorption hydroxyapatite and 70% Beta-tricalcium rapid resorption phosphate. The nanostructured surface promotes blood absorption by entering the granules of proteins and growth factors and promotes cell differentiation (23).

In another study (24), histological examinations were performed 6 months after surgery on bone samples in regenerated sites with materials based on hydroxyapatite, an amount of newly formed bone of 41% indicated a residual biomaterial quantity of 27% and a presence of medullary spaces of about 30%.

A fundamental role is played by the membranes that cover the grafted material. In the last decade, non-absorbable membranes, resorbable membranes, collagen sponges and non-cross-linked xenogenic collagen matrices have been used. The resorbable and non-absorbable membranes can be totally covered with a coronally advanced flap to obtain the primary closure following the biological principles of guided bone regeneration. However, this procedure inevitably changes the gingival architecture and location of the mucogingival junction. To avoid this, today, we tend to use crosslinked collagen membranes with different periods of resorption that can be intentionally left exposed to provide a transient barrier function (25). Histological studies of non-cross-linked collagen matrices in non-submerged or submerged environments revealed



**Fig. 20.** Case 2. Final X-ray.

complete integration with mature mucosal and submucosal tissues and membrane revascularization after 3 months (26).

The membrane used (Evolution Fine, TecnoSS®, Giaveno, Italy) is obtained from heterologous mesenchymal tissue and is completely resorbable. Experimental studies have shown histological evidence of the prolonged barrier effect of this membrane, which lasts at least eight weeks, protecting the graft from external agents (27).

The possible beneficial effect on the amount of bone resorption that occurs after tooth extraction with flap or flapless is controversial. Although some studies have shown slightly less pronounced bone remodelling of the alveolar crest after flapless extraction (28), other studies have failed to find significant differences between the flap and flapless tooth extractions (29).

Although it is suggested that wound dehiscence and membrane exposure may cause infection and lack of bone formation, recent studies show that intentional exposure to bioresorbable barriers does not adversely affect alveolar preservation procedures (30, 31).

A study in 2016 showed that the dense collagen matrix Evolution protects the graft from infection in case of membrane exposure, which does not become infected, and the wound heals by second intention (32). In these two clinical cases, we used a flapless approach and left the membrane exposed during healing. Lifting and advancing a full-thickness flap can cause a marginal recession to adjacent teeth, alteration of the shape of the papillae, loss of keratinized mucosa, and greater post-operative discomfort for the patient.

In this study, we observed no adverse effects where membranes were left exposed to the oral cavity and sutures were mainly used to hold the membranes in place.

## CONCLUSIONS

Oral rehabilitation with dental implants requires sufficient bone architecture in both vertical and horizontal dimensions. The socket preservation after extraction plays a fundamental role in maintaining the alveolar ridge. The clinical results obtained in these two cases seem to indicate that the use of Beta-tricalcium phosphate as a filler, and a membrane of completely resorbable heterologous mesenchymal tissue, are able to convey and promote bone regeneration.

## REFERENCES

1. Schropp L, Wenzel A, Kostopoulos L, Karring T. Bone healing and soft tissue contour changes following single-tooth extraction: a clinical and radiographic 12-month prospective study. *The International Journal of Periodontics & Restorative Dentistry*. 2003;23(4):313-323.
2. Tan WL, Wong TLT, Wong MCM, Lang NP. A systematic review of post-extraction alveolar hard and soft tissue dimensional changes in humans. *Clinical Oral Implants Research*. 2011;23(S5):1-21. doi:<https://doi.org/10.1111/j.1600-0501.2011.02375.x>
3. MacBeth N, Trullenque-Eriksson A, Donos N, Mardas N. Hard and soft tissue changes following alveolar ridge preservation: a systematic review. *Clinical Oral Implants Research*. 2016;28(8):982-1004. doi:<https://doi.org/10.1111/clr.12911>
4. Di Girolamo M, Barlattani A, Grazzini F, et al. Healing of the post extractive socket: technique for conservation of alveolar crest by a coronal seal. *Journal of Biological Regulators and Homeostatic Agents*. 2019;33(6 Suppl. 1):125-135. DENTAL SUPPLEMENT.
5. Su Y, Tang J, Min S, et al. Alveolar ridge dimensional changes following ridge preservation procedure with novel devices: part 3 - histological analysis in non-human primate model. *Clinical Oral Implants Research*. 2017;28(11):e252-e261. doi:<https://doi.org/10.1111/clr.13010>
6. Schwarz F, Ramanauskaite A. It is all about peri-implant tissue health. *Periodontology 2000*. 2022;88(1):9-12. doi:<https://doi.org/10.1111/prd.12407>
7. Volpe S, Di Girolamo M, Pagliani P, Zicari S, Sennerby L. Osteotome-Induced Blood Clot and Subsequent Bone Formation with the Use of Collagen Sponge for Integration of Single Dental Implants into the Atrophied Posterior Maxilla: A Retrospective Follow-Up of 36 Implants after 5 to 13 years. *Int J Dent*. 2022;2022:1-8. doi:<https://doi.org/10.1155/2022/6594279>

8. Avila-Ortiz G, Chambrone L, Vignoletti F. Effect of alveolar ridge preservation interventions following tooth extraction: A systematic review and meta-analysis. *Journal of Clinical Periodontology*. 2019;46(S21):195-223. doi:<https://doi.org/10.1111/jcpe.13057>
9. Stumbras A, Kuliesius P, Januzis G, Juodzbaly G. Alveolar Ridge Preservation after Tooth Extraction Using Different Bone Graft Materials and Autologous Platelet Concentrates: a Systematic Review. *Journal of Oral and Maxillofacial Research*. 2019;10(1). doi:<https://doi.org/10.5037/jomr.2019.10102>
10. Natto ZS, Parashis AO, Steffensen B, Ganguly R, Finkelman M, Y. Natalie Jeong. Efficacy of collagen matrix seal and collagen sponge on ridge preservation in combination with bone allograft: A randomized controlled clinical trial. *J Clin Periodontol*. 2017;44(6):649-659. doi:<https://doi.org/10.1111/jcpe.12722>
11. Ghanaati S, Schlee M, Webber MJ, et al. Evaluation of the tissue reaction to a new bilayered collagen matrix *in vivo* and its translation to the clinic. *Biomed Mater*. 2011;6(1):015010-015010. doi:<https://doi.org/10.1088/1748-6041/6/1/015010>
12. Walker CJ, Prihoda TJ, Mealey BL, Lasho DJ, Noujeim ME, Huynh-Ba G. Evaluation of Healing at Molar Extraction Sites With and Without Ridge Preservation: A Randomized Controlled Clinical Trial. *J Periodontol*. 2017;88(3):241-249. doi:<https://doi.org/10.1902/jop.2016.160445>
13. Lyu C, Shao Z, Zou D, Lu J. Ridge Alterations following Socket Preservation Using a Collagen Membrane in Dogs. *Biomed Res Int*. 2020;2020:1-9. doi:<https://doi.org/10.1155/2020/1487681>
14. Zhao L, Xu T, Hu W, Chung KH. Preservation and augmentation of molar extraction sites affected by severe bone defect due to advanced periodontitis: A prospective clinical trial. *Clinical Implant Dentistry and Related Research*. 2018;20(3):333-344. doi:<https://doi.org/10.1111/cid.12585>
15. Keil C, Gollmer B, Zeidler-Rentzsch I, Gredes T, Heinemann F. Histological evaluation of extraction sites grafted with Bio-Oss Collagen: Randomized controlled trial. *Annals of Anatomy - Anatomischer Anzeiger*. 2021;237:151722. doi:<https://doi.org/10.1016/j.aanat.2021.151722>
16. Fugazzotto PA. GBR using bovine bone matrix and resorbable and nonresorbable membranes. Part 1: histologic results. *The International Journal of Periodontics & Restorative Dentistry*. 2003;23(4):361-369.
17. Xu J, Aoki H, Shohei Kasugai, Otsuka M. Enhancement of mineralization on porous titanium surface by filling with nano-hydroxyapatite particles fabricated with a vacuum spray method. *Mater Sci Eng C Mater Biol Appl*. 2020;111:110772-110772. doi:<https://doi.org/10.1016/j.msec.2020.110772>
18. Loukelis K, Papadogianni D, Chatzinikolaidou M. Kappa-carrageenan/chitosan/gelatin scaffolds enriched with potassium chloride for bone tissue engineering. *International Journal of Biological Macromolecules*. 2022;209:1720-1730. doi:<https://doi.org/10.1016/j.ijbiomac.2022.04.129>
19. Khanna R, Katti KS, Katti DR. Bone nodules on chitosan–polygalacturonic acid–hydroxyapatite nanocomposite films mimic hierarchy of natural bone. *Acta Biomaterialia*. 2011;7(3):1173-1183. doi:<https://doi.org/10.1016/j.actbio.2010.10.028>
20. Ballouze R, Marahat MH, Mohamad S, Saidin NA, Kasim SR, Ooi JP. Biocompatible magnesium-doped biphasic calcium phosphate for bone regeneration. *Journal of Biomedical Materials Research Part B: Applied Biomaterials*. 2021;109(10):1426-1435. doi:<https://doi.org/10.1002/jbm.b.34802>
21. Sigusch BW, Dietsch S, Berg A, et al. Antimicrobial photodynamic active biomaterials for periodontal regeneration. *Dental Materials*. 2018;34(10):1542-1554. doi:<https://doi.org/10.1016/j.dental.2018.06.026>
22. 22.J. Jiménez Garcia, S. Berghezán, João Caramês, Dard M, Marques D. Effect of crosslinked vs non-cross-linked collagen membranes on bone: A systematic review. *J Periodontol Res*. 2017;52(6):955-964. doi:<https://doi.org/10.1111/jre.12470>
23. 23.Sbricoli L, Guazzo R, Annunziata M, Gobbato L, Bressan E, Nasti L. Selection of Collagen Membranes for Bone Regeneration: A Literature Review. *Materials*. 2020;13(3):786. doi:<https://doi.org/10.3390/ma13030786>
24. 24.Romasco T, Tumedei M, Inchingolo F, et al. A Narrative Review on the Effectiveness of Bone Regeneration Procedures with OsteoBiol® Collagenated Porcine Grafts: The Translational Research Experience over 20 Years. *J Funct Biomater*. 2022;13(3):121-121. doi:<https://doi.org/10.3390/jfb13030121>



25. 25.Stoppenbrink D, Daratsianos N, Kutschera E, et al. Dimensional changes of the alveolar ridge contour of the premolar extraction site in adolescents. *Journal of Orofacial Orthopedics / Fortschritte der Kieferorthopädie*. 2019;80(4):205-215. doi:<https://doi.org/10.1007/s00056-019-00182-w>
26. 26.Araújo MG, Silva CO, Misawa M, Sukekava F. Alveolar socket healing: what can we learn? *Periodontology 2000*. 2015;68(1):122-134. doi:<https://doi.org/10.1111/prd.12082>
27. 27.Parashis AO, Hawley CE, Stark PC, Ganguly R, Hanley JB, Steffensen B. Prospective Clinical and Radiographic Study of Alveolar Ridge Preservation Combining Freeze-Dried Bone Allograft With Two Xenogeneic Collagen Matrices. *Journal of Periodontology*. 2016;87(4):416-425. doi:<https://doi.org/10.1902/jop.2016.150500>
28. 28.Yu K, Liu W, Wang H, Tan Z. New Incision and Flap Designs in Autogenous Bone Ring Grafting with Simultaneous Implant Placement and an Evaluation of the Effect of First-Stage Wound Dehiscence in Dogs. *Int J Oral Maxillofac Implants*. 2020;35(4). doi:<https://doi.org/10.11607/jomi.8010>
29. 29.Jambhekar S, Kern F, Bidra AS. Clinical and histologic outcomes of socket grafting after flapless tooth extraction: a systematic review of randomized controlled clinical trials. *The Journal of Prosthetic Dentistry*. 2015;113(5):371-382. doi:<https://doi.org/10.1016/j.prosdent.2014.12.009>
30. 30.M Di Girolamo, Cecchetti F, Volpe L, et al. Critical investigation of the pre-surgical anatomic factors that influence the aesthetic result in post-extractive implantology and immediate prosthetic. *J Biol Regul Homeost Agents*. 2021;35(3S1):127-137. doi:<https://doi.org/10.23812/21-3supp1-15>
31. 31.M Di Girolamo, Arullani CA, Calcaterra R, Manzi J, Arcuri C, L Baggi. Preservation of extraction socket in immediate implant placement: a clinical study. *Oral Implantol (Rome)*. 2017;9(4):222-232. doi:<https://doi.org/10.11138/orl/2016.9.4.222>
32. 32.DI Girolamo M, Calcaterra R, DI Gianfilippo R, Arcuri C, Baggi L. Bone level changes around platform switching and platform matching implants: a systematic review with meta-analysis. *ORAL & implantology*. 2016;9(1):1-10. doi:<https://doi.org/10.11138/orl/2016.9.1.001>